



National Institute for Public Health
and the Environment
Ministry of Health, Welfare and Sport

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Interpretation of the 2014 experiment with
the Substance Emission Model

RIVM Report 2016-0063

A.M.A. van der Linden et al.



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Colophon

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Synopsis

Fate of plant protection products in soilless cultivations after drip irrigation: measured vs. modelled concentrations

Interpretation of the 2014 experiment with the Substrate Emission Model

The Greenhouse Emission Model has recently been adopted as a model package for assessing emissions to and concentrations in groundwater and surface water after use of plant protection products in greenhouse crops. Stakeholders advised that the model be tested against experimental data.

In October 2014, facilities of WUR Plant Research were used to perform a pilot experiment in which cucumber plants on stone wool substrate were treated with three plant protection products, using a drip irrigation method. Concentrations of the active substances were measured in both the water flowing to and draining from the substrate. GEM was tailored to the experimental conditions and used to predict concentrations in parts of the experimental system. Measured and simulated concentrations of imidacloprid and fluopyram were comparable from approximately 36 hours after the start of the experiment onwards. Prior to this, concentrations in the inflowing water were underestimated and concentrations in the drain water were overestimated, probably because of incomplete mixing. For dimethomorph, agreement between the measured and calculated concentrations was reached after approximately 80 hours. This more lengthy period may be due to exceeding the solubility of the substance, causing precipitation or settling on the tube walls, and redissolving later on; the model does not account for these processes.

Degradation of all three substances was found to be negligible over the duration of the experiment. Plant uptake was the major dissipation process. Experimental results show that uptake of substances was lower than uptake of water, thereby supporting the transpiration stream concentration approach proposed by Briggs et al. (1982); this approach is often applied however experimental evidence is scarce. Transpiration stream concentration factors far below one were found to fit experimental results best.

Keywords: authorisation, GEM, greenhouse, plant protection products, stone wool, substrate, validation status

Publiekssamenvatting

Gedrag van gewasbeschermingsmiddelen in substraatteelten na toepassing via het druppelsysteem

Interpretatie van het 2014 pilot-experiment

Door het gebruik van gewasbeschermingsmiddelen in kassen kunnen restanten van deze middelen in het nabijgelegen oppervlaktewater terechtkomen en het waterleven aantasten. Bij de toelatingsbeoordeling worden de effecten op het waterleven geëvalueerd met het zogeheten Greenhouse Emission Model (GEM). Dit model is door het RIVM ontwikkeld in samenwerking met Wageningen Environmental Research en Wageningen Plant Research.

Het RIVM heeft het model getoetst in een 'semi-praktijkexperiment' met drie stoffen. Hieruit blijkt dat het vertrouwen waarmee dit model de concentraties in het teeltsysteem kan voorspellen, aanzienlijk is vergroot. Om te kunnen berekenen in welke mate de stoffen in oppervlaktewater terechtkomen als water wordt geloosd, is het van belang om de concentraties in het teeltsysteem goed te kunnen voorspellen.

In het experiment zijn de gewasbeschermingsmiddelen met het gietwater aan een komkommernog op substraat toegediend. Vervolgens zijn zes dagen lang de concentraties in het gietwater en het drainwater gemeten. Dit is voldoende lang om belangrijke processen in het teeltsysteem, het hergebruik van water en de opname van stoffen door planten, te onderzoeken. Van twee stoffen blijken de berekende concentraties in het water dat uit het teeltsysteem stroomt vanaf ongeveer 36 uur na aanvang van het experiment in de buurt van gemeten concentraties te liggen. Bij de derde stof was dit na ongeveer 80 uur het geval. De belangrijkste reden waardoor de stoffen uit het water verdwenen was dat ze door de planten worden opgenomen; afbraak speelde in dit experiment geen rol van betekenis.

Kernwoorden: GEM, gewasbeschermingsmiddelen, kas, toelating, steenwol substraat, validatiestatus

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Summary

In October 2014, an experiment was performed in which cucumber plants on stone wool substrate were treated with three plant protection products, using a drip irrigation method. Concentrations of the active substances, dimethomorph, fluopyram and imidacloprid were measured in both the water flowing to and draining from the substrate for six days following application. The water circulated through the system throughout the experiment; fresh water was supplied to the system several times, discharge from the system did not occur.

Results of the experiment were used to test the Substance Emission Model (SEM), the part of the Greenhouse Emission Model (GEM) that is used to predict behaviour of substances in substrate growing systems and emissions from these systems to surface water. The major concepts underlying the model are that the solution in the various parts of the system can be considered well mixed, that sorption in substrate systems with stone wool is absent, and that the dissipation of the substance is by plant uptake and degradation in the water phase. The model's scenario was adapted to the lay-out of the experiment. Water flows in the system were derived from recorded volumes and other observations during the experiment. However, drain water volumes were not recorded for three days; target irrigation surplus was used to fill this gap.

Simulations with SEM were performed for the applied amounts using two approaches. In the first, the amount was derived from measured concentrations in the application solution, whereas in the second, this was derived from the weighted masses. The two approaches were followed because of uncertainties about the amounts that effectively arrived in the system. Measured and simulated concentrations of imidacloprid and fluopyram were comparable from approximately 36 hours after the start of the experiment onwards. Prior to that, concentrations in the inflowing water were underestimated and concentrations in the drain water overestimated, probably because of incomplete mixing. For dimethomorph, agreement between measured and calculated concentrations was reached after approximately 80 hours. This more lengthy period may be due to exceeding the solubility of the substance, causing precipitation, or settling on the tube walls and redissolving later on; the model does not account for these processes.

It cannot be concluded from this experiment whether sorption plays a role or not. Degradation of all three substances was found to be negligible over the experiment's duration; half-lives are above six days; more precise values cannot be derived. Plant uptake was the major dissipation process: 54%, 34% and 45% uptake was calculated for dimethomorph, fluopyram and imidacloprid, respectively. Experimental results show that uptake of substances was relatively lower than uptake of water, thereby supporting the transpiration stream concentration approach that is often applied but for which experimental evidence is scarce. Transpiration stream concentration factors far below one were found to fit experimental results best.

1 Introduction and problem definition

Emissions of plant protection products (PPP) from greenhouses are currently a major source of water contamination in areas with a high greenhouse density. The Greenhouse Emission Model (GEM) was developed and introduced in 2008 to simulate emissions of PPPs from greenhouses and to calculate predicted environmental concentrations in groundwater and surface water (Cuijpers et al. 2008, Vermeulen et al. 2010, van der Linden et al. 2015, Wipfler et al. 2015). The model package consists of three models: WATERSTREAMS (Voogt et al. 2012), the Substance Emission Model (van der Linden et al. 2015), and TOXSWA (Adriaanse 1996). The WATERSTREAMS model simulates water flows within the greenhouse including the discharge moments and volumes. The Substance Emission Model simulates the fate of PPP within the greenhouse including degradation and plant uptake, and TOXSWA calculates the fate of PPPs in the ditch.

The Substance Emission Model consists of three sub modules for different applications, which are:

1. Application of PPP through drip irrigation.
2. Spraying or fogging PPP in systems with drip irrigation.
3. Spraying or fogging PPP in systems with an ebb/flow system.

Detailed information is available from Van der Linden et al. (2015).

In 2012, two experiments were conducted to test the Greenhouse Emission Model (GEM) for soilless cultivations (Van der Maas et al. 2015). In the first experiment, gerbera and bell pepper were grown in two greenhouses at two commercial companies. Pymetrozine and propamocarb-hydrochloride were added through drip irrigation and water samples were taken at irregular time intervals for a period of 30 days. The second experiment was conducted in the laboratory with pot plants, focussing on the processes in and around the crop (plant uptake, sorption and degradation in soil). Measured concentrations in different compartments of the greenhouses (experiment 1) and in the in- and outgoing water fluxes of the pots (experiment 2) were compared with modelled concentrations. In the first experiment, the concentrations were lower than those estimated by the model. However, the cause of the difference between measured and modelled concentrations could not be assessed because of the limited number of available measurements and insufficiently precise information on water flows in the greenhouses. A follow-up experiment was conducted in the autumn of 2014 at the Wageningen UR Greenhouse horticulture site in Bleiswijk. The greenhouse at this site is equipped with a controlled irrigation and pesticide application system, which in principle should enable a better understanding of the processes at hand. A new experiment was designed to measure water fluxes and pesticide concentrations in a soilless cultivation, using drip irrigation; results were partly reported by Van der Maas et al. (2015). We interpret the results of this experiment in the current report. The results of an additional experiment aiming to validate the GEM model with bell pepper on stone wool, conducted in 2016, will be reported separately (Wipfler et al., in prep.)

The objective of this study was twofold: (i) interpretation of the results of the 2014 experiment using the Substance Emission Model, (ii) identification of essential processes and state parameters to be measured as a basis for the experimental design of the validation experiment for GEM.

Reading guidance

This report starts with a description of the 2014 experimental set-up and the data gathered during the experiment (chapter 2). The lay-out of the experiment required adaptations to the simulation model, i.e. changes to the size and number of tanks. The most important assumptions and changes to the model are described in chapter 3. Chapter 4 describes the interpretation of the experimental results and compares measured and simulated concentrations. Chapter 5 presents conclusions of the research and recommendations for further investigation. Recommendations, specifically regarding the set-up of new experiments, are given in chapter 6.

2 Overview of the experiment and available information

The experiment was performed in a standing cucumber crop used earlier for other experiments unrelated to the fate of PPPs. The crop received normal treatment, including treatment with PPPs and the crop management approach was continued; the only changes were related to the application of PPP. Crop management changes made shortly before the experiment may have influenced the results of the experiment. The experiment was the only reason for applying the PPPs, i.e. there was no pest to be treated. PPPs were applied on 22 October (see section 2.1) further referred to in this report as $t=0$.

2.1 Lay-out of the experimental system

The experiment was conducted in October 2014 in one unit (unit 6.10) of a greenhouse at Wageningen UR Greenhouse Horticulture in Bleiswijk, the Netherlands. Unit 6.10 has a total net surface area of 120 m², giving space to 276 fruit-carrying cucumber plants in 12 rows (23 plants per row). In each row, 11.5 stone wool slabs were laid down with 2 plants per slab. During the experiment, the cucumber plants were fully grown and carrying fruit.

Water was pumped from a supply tank (mixing tank) to the plants via a series of tubes. Each plant had a dripper to supply water, nutrients and, during the experiment, PPP (see Figure 2-1). The drippers open due to the increased pressure each time the pump is activated. Upon opening, the water supply from each dripper was approximately 50 ml per minute. Water and dissolved substances are then taken up by the crop, and any surplus water runs to the drain tank via a drain opening on the lowest side of each slab, and via a trough and PVC pipe system. Troughs were positioned at a slope of approximately 1% to ensure fast collection of the drain water. The drain solution was pumped back from the drain tank into the mixing tank.

The mixing tank had a variable content, controlled by two sensors preventing the system from flooding and from falling dry. During the experiment, the minimum water level was set at approximately 49 cm (volume 472 L) and the maximum level at 90 cm (volume 872 L). During operation, the lower sensor triggered the automatic delivery of 400 L fresh nutrient solution to the mixing tank. The automatic water supply may be overruled; for example, prior to and during the supply of the PPP the system was switched to manual (see section 2.3).

Irrigation (in blue) and drain system (in red):

- drip irrigation lines: length 13.2 m, diameter 16 mm
- main pipe: length 9 m, diameter 32 mm
- 12 rows with slabs, 12 troughs
 - slab size: 100 * 15 * 7.5 cm
 - per slab 2 plants and 2 drippers, 23 plants per row, 276 plants in total
- drippers: adjustable about 50 ml per minute
- mixing tank: 114 * 85 * 110 cm, maximum level by floater at 90 cm from bottom

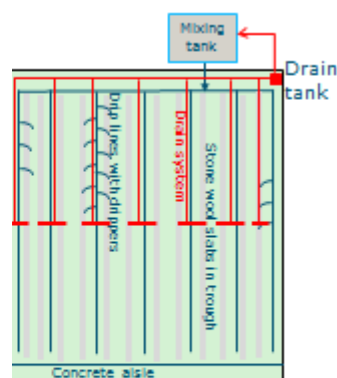
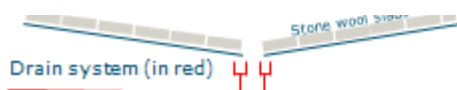


Figure 2-1 Layout of drip irrigation and drain system of the greenhouse unit in which the experiment was conducted. Green lines indicate the water supply, red lines the drain pipes, and grey lines the slabs and the troughs.

2.2 Operational measurements during the experiment

A number of measurements were performed to monitor and check crop growth, starting from the beginning of the cropping period and continued during the experiment. Measurements of radiation, temperature, water content of the slabs and electrical conductivity were logged every 5 minutes. Figure 2-2 gives an example of the measurements.

During the experiment, water supply to the crop was automatically supplied between 10.00 and 15.00, based on the water requirement of the crop as calculated from measured radiation and a target drain ratio (35% of the supplied water). Cumulative radiation triggered activation of the water pump and water supply for a fixed duration of 2.5 minutes. Each time water is supplied, the water content of the slabs rises (Figure 2-2). After each event, the water content drops due to water uptake by the crop and drainage to the troughs. Drainage lags behind water supply; usually no drainage was observed after the first water supply of the day, and none to little drainage after the second water supply. Drainage was measured occasionally during the experiment, not continuously (see Table 2-1); if not measured, the target value is given.

Events preceding the application of the PPPs may have affected experimental conditions during the experiment. An example is that the water supply was discontinued the day before the application of the PPPs in order to lower the water content of the slabs, which is prescribed in the 'Druppelprotocol (drip application protocol)'¹. For this reason, information on water management prior to application is included.

Figure 2-3 shows two days of the data given in Figure 2-2 in more detail: the day before and the day of the application of the PPPs. The watering regime was switched to manual control on 21 October, the day

¹ 'Druppelprotocol' is the guideline for applying PPP with drip irrigation in soilless cultivation.

before the PPPs were applied to the system. After 12.00, no water was provided until the application of the PPPs on 22 October. The effect is visible in Figure 2-2; the water content drops to 53%, about 5% lower than the minimum water contents in the previous period.

The first watering event of 22 October lasted 6 minutes, where 74 L of the PPP containing solution was applied in one event. After this, the mixing tank was emptied to discard surplus PPP solution, and refilled with fresh nutrient solution to the maximum level, i.e. 872 L. The water supply was switched to the automatic position after this event, however the fourth watering event was manually forced. The following day, the first watering event was also manual; for unknown reasons it was not started automatically.

The water content of the slabs increased during the application of the PPPs, but remained below the minimum values observed the days before. No drainage was observed during and shortly after this event and also not after the following watering event. After the second watering event, a small amount of drainage was observed, but not everywhere in the system. After the next watering event, some drainage was observed at all observation points (see Figure 2.4 for the location of the sampling and observation points in the system). The drainage amounts were considered too low, which led to the manually forced water supply. The volumes of applied water were registered daily on a computer (Table 2-1).

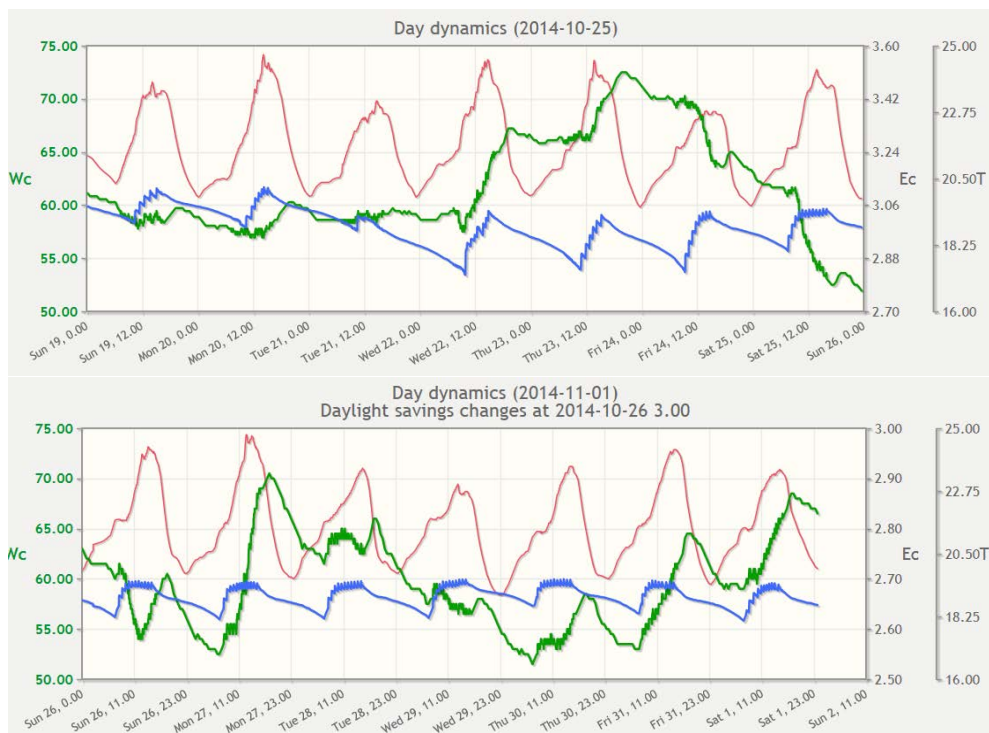


Figure 2-2 Data from the data logger in the slabs from 19 October to 2 November. WC=water content (%), T=temperature (°C), EC= electric conductivity (mS/cm). Note the difference in scale between upper and lower panel for EC.

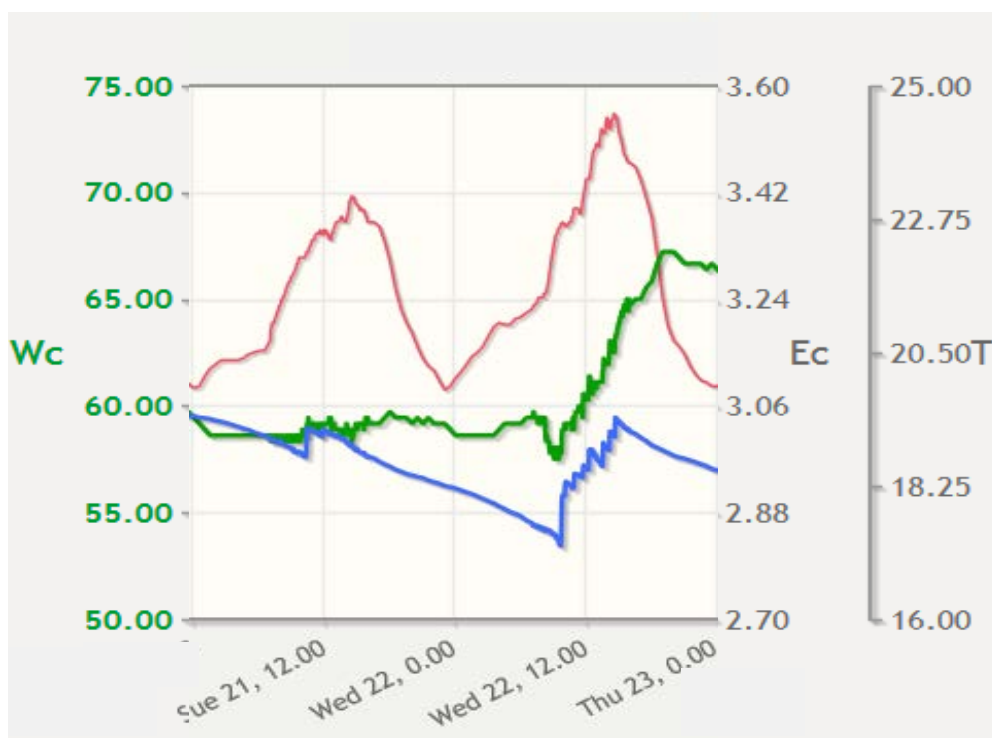


Figure 2-3 Data from the data logger in the slabs for 21 and 22 October.
 WC=water content (% , blue, left axis), T=temperature (°C, red, 2nd right axis),
 EC= electric conductivity (mS/cm, green, 1st right axis).

Table 2-1 Details on the daily water supply during the experimental period.

date	volume of water (L)	number of watering events	drain (%)
20 October	259	7	40
21 October	74	2	6
22 October	259	7	15
23 October	222	6	15
24 October	296	8	31
25 October	444	12	35 [#]
26 October	444	12	35 [#]
27 October	518	14	35 [#]
28 October	444	12	27
29 October	444	12	35

[#] estimated value; missing information prevented accurate calculation.

Temperature is important as it may influence the behaviour of the substances in the system. Temperature was recorded during the entire season, thus also during the experiment (see Figure 2-2 and Figure 2-3). The temperature varied between approximately 19 and 25 °C.

2.3 Application of the plant protection products

The experiment was performed using three PPPs: Paraat, Luna and Admire with active substances dimethomorph, fluopyram and imidacloprid, respectively. Label instructions on the dosage of these PPP are given in Appendix 1. The weighed amounts of the PPPs and nominal concentrations in the mixing tank are given in Table 2.2. In addition to the three PPPs, a tracer (Fe-EDDHA, 40 mmol/l) was added so that the movement of the solution in the system could be followed visually. Fe-EDDHA is readily taken up by plants and therefore cannot be considered a quantitative tracer.

The PPP were applied on 22 October at 10.00. The required amounts of the three PPPs were determined, added to 2 L water and thoroughly mixed. The mixture was added to the mixing tank containing nutrient solution to reach a total volume of 166 L and thoroughly mixed. The PPP were applied by drip irrigation to the crop. The pump was activated manually and run for 6 minutes. In this way, 74 L of the solution was pumped to the cucumber plants and the rest was discarded by draining the mixing tank. Hereafter, the mixing tank was filled with fresh nutrient solution and the system was reset to automatic application.

Table 2-2 Applied masses of plant protection products, nominal concentrations of the active substances (a.s.) in the mixing tank (MT), and solubility. The nominal concentrations were calculated from the weighed masses and the batch volume. The applied amounts were calculated from the volume supplied to the system (74 L).

PPP	active substance	applied mass a.s. (mg)	nominal a.s. concentration (mg/L)	solubility a.s. (mg/L)
Paraat	dimethomorph	8360	113	47.2, 10.7 [#]
Luna	fluopyram	2270	30.7	15
Admire	imidacloprid	1120	15.1	613

[#] solubility for E and Z isomers respectively

2.4 Sample collection and analyses

Samples were collected between 22 October and 28 October 2014:

- On 22 October, prior to application of the PPP, samples were taken (in duplicate) from both the mixing tank and the drain water tank to establish background concentrations.
- Samples were taken from the mixing tank directly after preparing the PPP-solution, and again after the PPPs were supplied to the cultivation.
- During application, samples were taken from the drippers at three locations in the greenhouse unit, positioned close to the yellow dots in Figure 2-4.
- On 22 October, following the primary application of PPPs, samples were taken at the same locations during the first three watering events and during the last watering event of the day. Replicates were taken when possible. Immediately after taking samples from the drippers, samples were taken from the drain water (blue rectangles in Figure 2-4), when drainage occurred.
- On the following days, samples were taken at the same locations, but only once a day after the last watering event.

All samples were transferred into dark green 1 L glass bottles and sent to a commercial laboratory where they were analysed for the three substances.

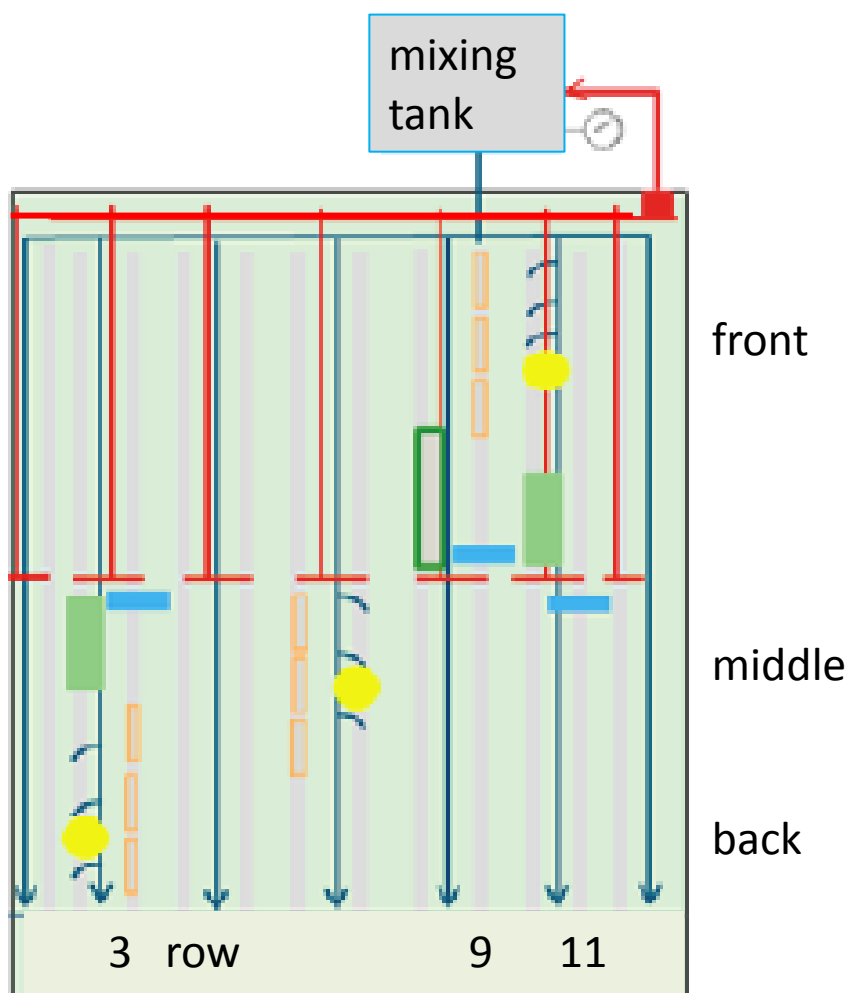


Figure 2-4 Sampling locations in the greenhouse unit. The yellow dots indicate the position of the sampled drippers. Drainage samples were collected at the blue rectangles (troughs in rows 3, 9 and 11).

2.5 Analytical results

Samples were brought to the laboratory and analysed for the active substances of the three PPPs. Selected samples were also analysed for the Fe content; elevated concentrations were due to the use of the FeEDDHA colour tracer. The active substance of Paraat is dimethomorph: a mixture of the isomers E-dimethomorph and Z-dimethomorph. The laboratory did not distinguish between the two isomers; therefore, total dimethomorph contents were used for the interpretation of the experiment.

Background concentrations of the active substances were low, but all three substances were found in the nutrient solution and in the drain water sampled prior to application of the PPPs (Table 2-3).

Table 2-3 Background concentrations ($\mu\text{g/L}$) in water in the cultivation system

sampled water	concentration		
	dimethomorph	fluopyram	imidacloprid
mixing tank before application	0.014	0.068	0.011
	0.018	0.061	0.011
fresh supply mixing tank after application	0.11	0.048	#
	0.094	0.043	#
drain water before application	0.025	0.076	0.033
	0.03	0.064	0.022
average	0.049	0.060	0.019

not measured

Table 2-4 lists the concentrations of the three substances in the application solution. Samples of this solution were taken in duplicate from the mixing tank at the start of the application event and from the remaining application solution following application. The remaining application solution was discarded after taking the samples, and the mixing tank was refilled with fresh nutrient solution.

Table 2-4 Concentrations (mg/L) in PPP application solution

	dimethomorph	fluopyram	imidacloprid
before dose 1 [#]	26	22	30
before dose 2	18	26	30
after dose 1	19	30	36
after dose 2	21	30	42
Average	21	27	34.5

1 and 2 are replicates

Figure 2-5 to Figure 2-6 show the concentrations in the solutions sampled from the drippers in the glasshouse unit in time, for dimethomorph, fluopyram and imidacloprid, respectively. At the first sampling moment, concentrations are equal to the levels measured in the application solution, except for the drippers at the back which lagged behind. Concentrations decreased rapidly, as fresh nutrient solution was supplied. Later, the levels stabilise or even increase due to recirculation of the nutrient solution containing PPPs; from the fourth watering event on October 22 onwards (see Appendix 3).

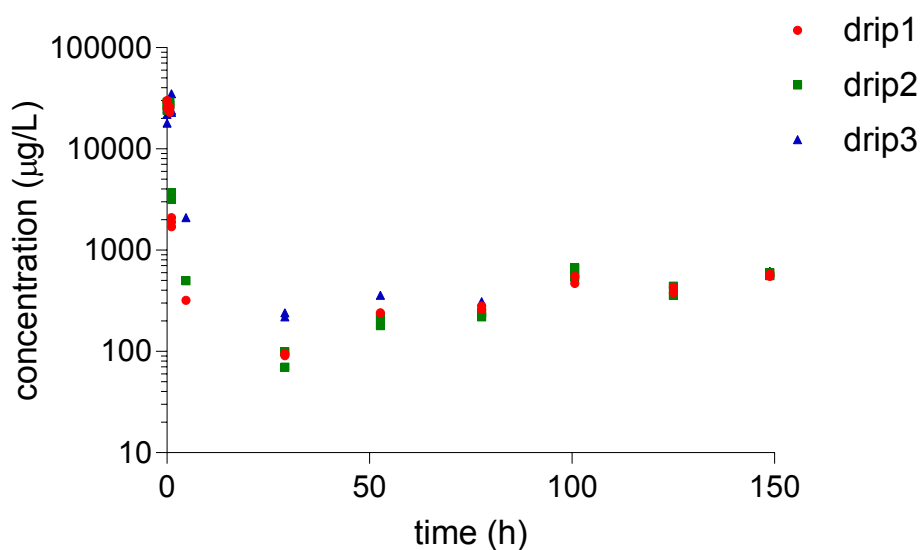


Figure 2-5 Dimethomorph concentrations in samples taken from the drippers. Drippers 1, 2 and 3 are positioned in the front, middle and back of the unit respectively. Note the logarithmic scale of the Y-axis. Application of substance is at $t=0$ h.

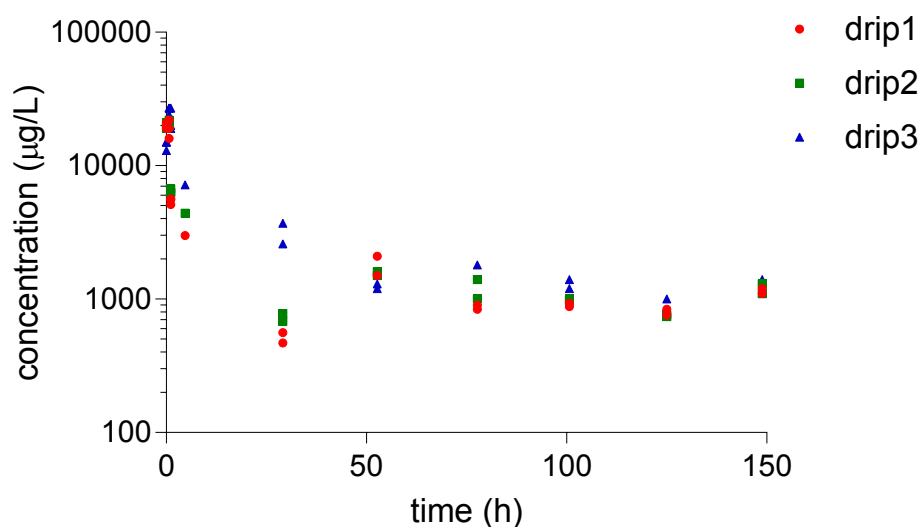


Figure 2-6 Fluopyram concentrations in samples taken from the drippers. Drippers 1, 2 and 3 are positioned in the front, middle and back of the unit respectively. Note the logarithmic scale of the Y-axis. Application of substance is at $t=0$ h.

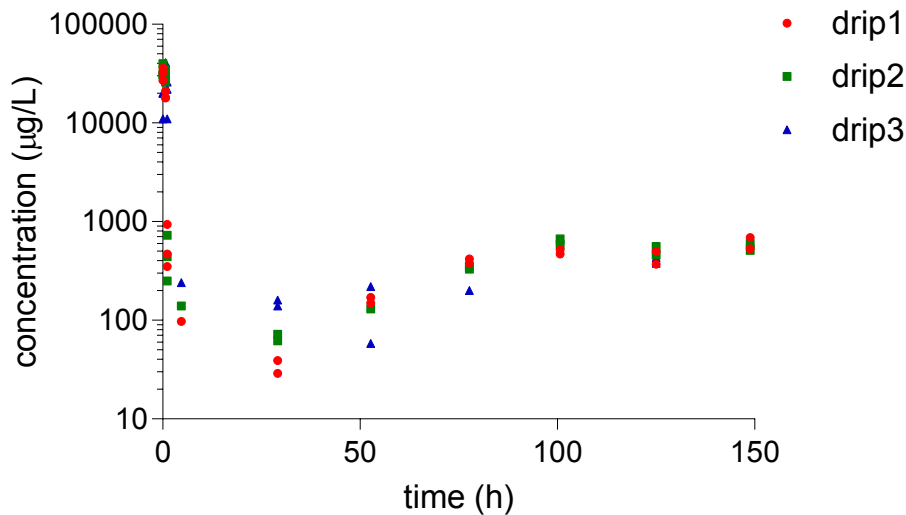


Figure 2-7 Imidacloprid concentrations in samples taken from the drippers. Drippers 1, 2 and 3 are positioned in the front, middle and back of the unit respectively. Note the logarithmic scale of the Y-axis. Application of substance is at $t=0$ h.

Figure 2-8 to Figure 2-10 show concentrations of dimethomorph, fluopyram and imidacloprid in the water draining from the slabs. The first moment drain water appeared from all slabs was after 3.15 h (3rd watering event after application of the PPPs, but the volume was too low to take duplicate samples). Concentrations in this drain water were at background level. Concentrations increased more rapidly in the front end of the greenhouse unit than in the middle and the back. Concentrations of all three substances increased up to three to four days after the application, depending on the substance.

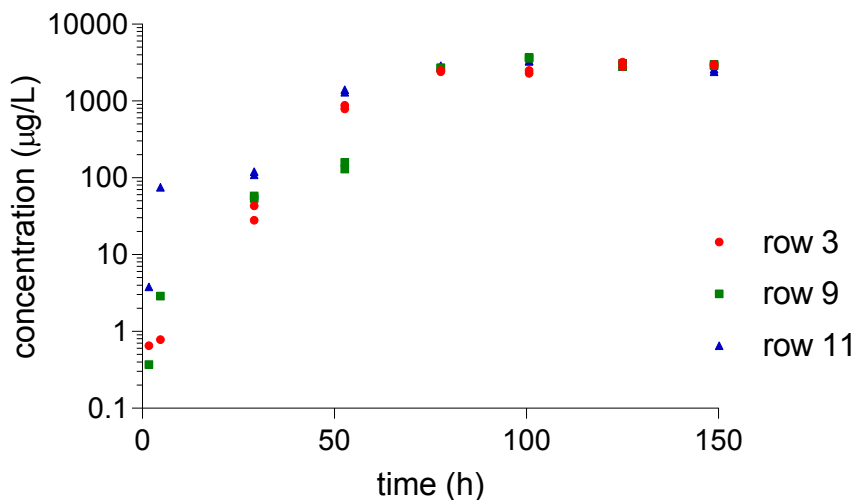


Figure 2-8 Dimethomorph concentrations in samples taken from the troughs. Row 11 is in the front of the greenhouse unit, row 9 in the middle and row 3 in the back. Note the logarithmic scale of the Y-axis.

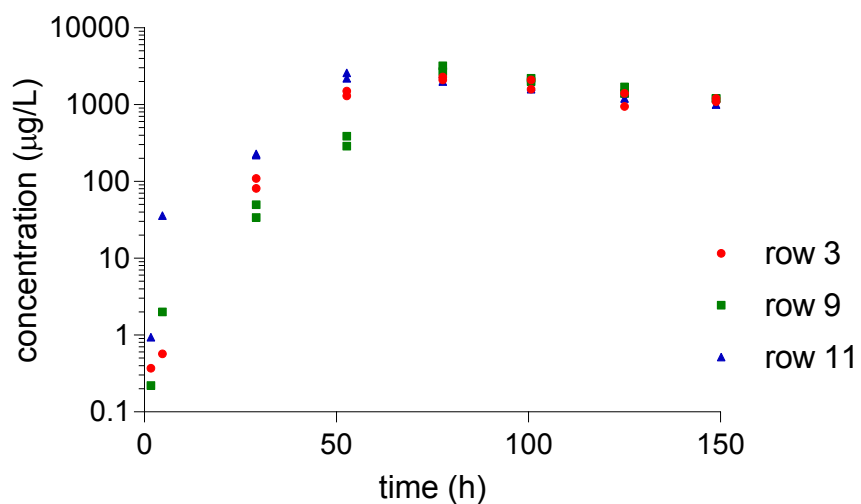


Figure 2-9 Fluopyram concentrations in samples taken from the troughs. Row 11 is in the front of the greenhouse unit, row 9 in the middle and row 3 in the back.

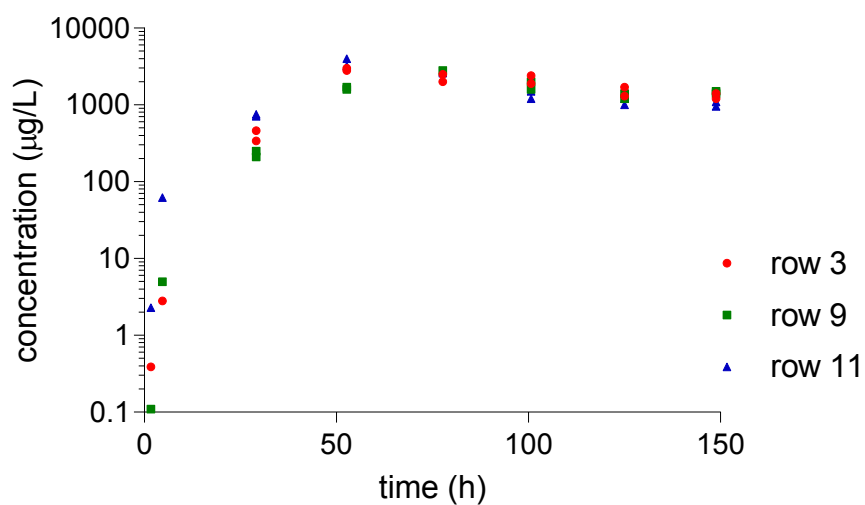


Figure 2-10 Imidacloprid concentrations in samples taken from the troughs. Row 11 is in the front of the greenhouse unit, row 9 in the middle and row 3 in the back.

3 Adaptations to the model and generation of input files

In GEM (Wipfler et al. 2015, van der Linden et al. 2015), three models are used to simulate the fate and behaviour of substances in substrate cultivations. The WATERSTREAMS model (Voogt et al. 2012) simulates the water flows in the greenhouse including the water supply to the crop and the amounts discharged from the system to the surface water. The substance emission model uses the water flows generated by the WATERSTREAMS model and calculates the behaviour of a substance in the greenhouse system, including concentrations of the substance in various tanks, uptake by the crop, and the discharge to the surface water. TOXSWA calculates the fate and behaviour of the substance in the ditch into which the water leaving the greenhouse is discharged. The results are further used in the authorisation procedure.

This report only addresses the second model; the Substance Emission Model (SEM). In order to interpret the results of the experiment and identify recommendations for a future validation experiment, the model had to be adapted to mimic the experimental set-up (see section 3.1). During the experiment, several water flows were recorded and these were transferred into input files for the Substance Emission Model. Thus the measured water flows in this case replace the water flows calculated with the WATERSTREAMS model (see section 3.2). During the experiment, no discharge to surface water occurred.

Major assumptions in SEM are that the solution in the various parts of the system is well-mixed, that sorption in substrate systems with stone wool is absent, and that the dissipation of the substance is by plant uptake and degradation in the water phase.

3.1 Adaptations of the substance fate model

GEM scenarios were developed to resemble greenhouse conditions typical of current growing conditions. The scenarios are for a production area of 1 ha, a fair estimate of net production areas within one greenhouse in the Netherlands. The schematisation of the greenhouse in GEM is different from the lay-out of the experiment in the greenhouse unit in Bleiswijk. The greenhouse unit production area was much smaller, fewer tanks were used, and the relative sizes of the tanks were different. The principle of the growing system, however, can be regarded as identical to the principle in the GEM scenarios for crops grown on substrate slabs, i.e. the crop received its nutrient solution from the mixing tank and the surplus water drained to the drain water tank.

The mixing tank in the experiment combined the functions of the drain water tank, the clean water tank, and the mixing tank in the GEM scenarios. This – triple – function required the mixing tank to be relatively large, much larger than its relative size in the GEM scenarios. Figure 3-1 presents a simplified schema of the experimental system.

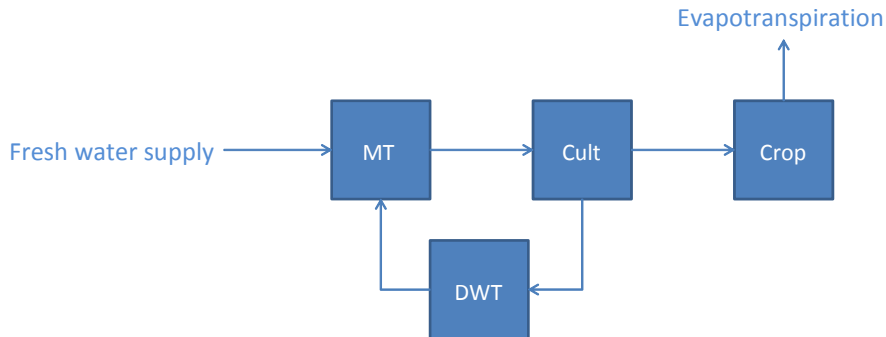


Figure 3-1 Backbone schematisation of the experimental system for use in a simulation package. Water flows are indicated with arrows. MT=mixing tank, Cult= cultivation consisting of slabs including the plant roots, Crop= stems, leaves and fruits of a greenhouse crop, DWT=drain water tank.

The following section addresses the most important assumptions and processes pertinent to and occurring in the system. A full description is provided by Van der Linden et al. (2015). One of the most important concepts in the Substance Emission Model for soilless cultivation in GEM is that the system can be described as a series of connected tanks with a known constant volume of water, which is assumed to be perfectly mixed in each tank. The concentration of a substance in a tank is determined from inflow and outflow, and processes taking place in the tank. There is no check in the model on whether the solubility of the substance is exceeded.

Degradation in all tanks is described according to a first-order process:

$$\frac{dC}{dt} = f_T k_{deg} C$$

where

C concentration of the substance, µg/L

f_T factor, -, for the influence of temperature on the degradation, f_T is a function of time and may be different for each tank

k_{deg} first-order degradation rate coefficient, d⁻¹

with

$$k_{deg} = \frac{\ln(2)}{DegT_{50}}$$

where

DegT₅₀ half-life of the substance, d, at reference conditions. The half-life may be specific for each tank.

The half-life may be specific to a tank and its operating conditions. For example, the half-life can be different in a disinfection tank because of addition of a disinfectant or application of UV light. This was not tested here as there was no disinfection unit operational during the experiment.

Degradation is dependent on the temperature. In GEM, this influence is calculated from the Arrhenius equation:

$$f_T = \exp\left(\frac{-E_{act}}{R}(T_{tank}^{-1} - T_{ref}^{-1})\right)$$

where

E_{act} Arrhenius activation energy, J mol⁻¹

R molar gas constant, 8.314 J mol⁻¹ K⁻¹

T_{tank} actual temperature in the tank, K

T_{ref} reference temperature, usually 293.15 K (20 °C)

One dissipation process is the uptake of substances by the crop. Passive uptake along with water is assumed, but, compared to water uptake, reduced dependent on substance properties. Currently this is implemented in GEM via the Transpiration Stream Concentration Factor (TSCF) which is calculated using Briggs' formula (Briggs et al. 1982):

$$TSCF = 0.784 \times e^{-\frac{(\log K_{ow} - 1.78)^2}{2.44}}$$

where

K_{ow} the octanol water partition coefficient, (-)

As noted earlier, GEM assumes constant water volumes for the various tanks. In the experimental situation, the water content of the mixing tank fluctuated. The mixing tank was equipped with floats which were connected to the computer. The lower float signalled the computer to deliver fresh water, while the upper float signalled the computer to stop delivery of fresh water. Additional safety floats attached to alarms, were installed, but safety levels were not breached during the experiment. The delivery of fresh water was assumed to be a pulse, so the fresh water was added instantly to the mixing tank. The added volume was assumed equal to the volume of the tank between the two floats.

Float levels and the surface area of the mixing tank were used to calculate the water content of the mixing tank. The water contents of the other tanks were assumed to be constant, although in practice some variation in time was noted.

During the experiment, for practical reasons, samples were taken from the drippers delivering nutrient solution to the slabs, and from the troughs collecting the drain water returning to the mixing tank. This was accounted for by considering the tubes between the mixing tank and cultivation and the tubes and troughs between cultivation and drain water tank as separate tanks, with known (small) volumes (Figure 3-2).

The model code used for the interpretation of the experiment is given in Appendix 2.

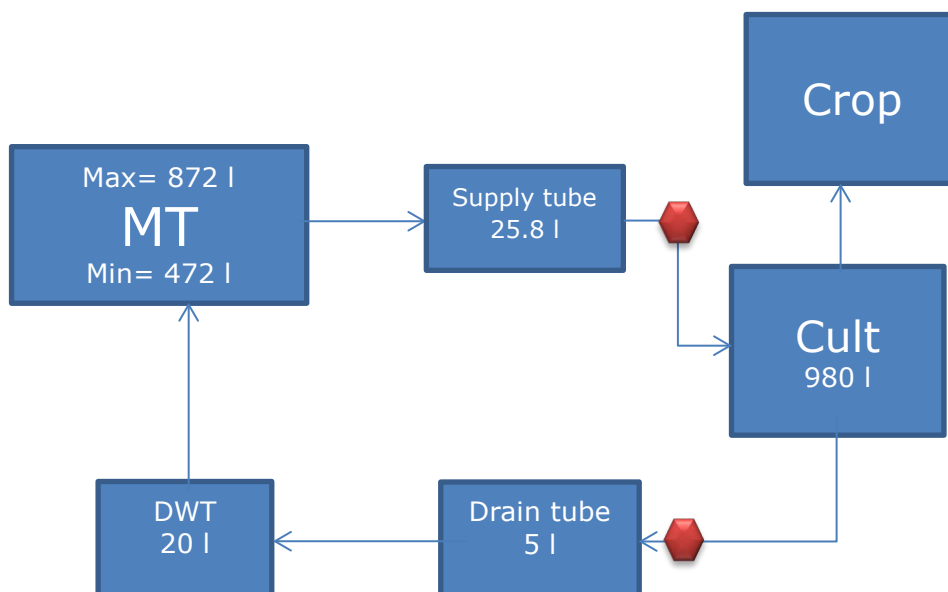


Figure 3-2 Overview of the experimental greenhouse used in the simulation model. MT= mixing tank, Cult=cultivation consisting of stone wool slabs and plant roots, Crop= plant stems, leaves and fruits. DWT=drain water tank. Supply tube and drain tube are the tube from the mixing tank to the cultivation and the troughs from the cultivation to the drain water tank, respectively. All volumes (litres) were calculated from the total length of the tubes and their inner diameters. The diamonds represent the sampling points. The volume of drain tube was not known and was estimated to be five litres.

3.2 Water input file for the calculations

From the experimental conditions, it was considered most appropriate to use an hour as basis for the input files for the calculations.

All tanks, except the mixing tank, were assumed to have constant water content over time, although some fluctuations occurred. This assumption is equal to the approach taken in GEM in the scenarios used for the authorisation evaluations. The assumption of constant water content is poor for the drain water tank, but the volume of this tank is so small that this has little influence on the calculation results.

The volume of the mixing tank fluctuates, as fresh nutrient solution is supplied in batches triggered by the floats in the tank. The content of the mixing tank is calculated from the water flows into (drainage water and fresh water) and out of the tank (water supply to the crop). Immediately after application of the PPPs, before the first watering event, the mixing tank was filled to the maximum level. This was used as the starting volume of the mixing tank. A float at 50 cm, i.e. at a water content of 472 L, signalled the delivery of 400 L fresh nutrient solution.

The volumes of the tubes were calculated from the lengths and diameters of the tubes present in the system. The water content of the cultivation was estimated to be 980 L. The average water content of the stone wool slabs in the first 8 days after the application of the substances was 58% (V/V), which amounted to a water content of 900 L in the slabs. At the start of the cultivation, i.e. months before the start of the experiment, plants were transferred to the slabs in a volume of

1 L stone wool substrate. Water content of this additional substrate was unknown (not measured). For the calculations, it was assumed that the average water content was 29%, i.e. 50% of the measured water content of the slabs. Water content was expected to be lower than in the slabs because of evaporation at the top and the vertical gully that was used to supply the solution, leading to lower water contents at the top.

The water supply from the mixing tank to the crop was registered daily by the central control system. However, in practice, the water was supplied in a limited number of watering events. In each event, nutrient solution was pumped for 2.5 minutes at a constant rate, equivalent to a volume of 30.8 L (about 10% lower than indicated in Figure 2.1). The water supply per minute (12.33 L) was calculated from the measured volume supplied during application of the PPPs (74 L) and the duration of the application event (6 minutes). The starting time of each event was read from the registration of the water content of the slabs; the water content increased immediately at each watering event. Each watering event was assigned to the nearest hour.

Surplus water drains from the substrate slabs under gravity. Usually, drainage starts each day either during or immediately after the second watering event and occasionally during or after the third watering event. Because of water uptake by the crop between the last watering event on the previous day and the first watering event on the current day, there is a shortage, and the surplus of the first watering event counterbalances this. Thus, the number of drainage periods on each day is mostly equal to the number of watering events minus one.

The drainage situation on the day of application was different from the other days. Due to the application of the 'Druppelprotocol (Drip application protocol)', the water content in the slabs was relatively low at the moment of application of the PPP. The volume of water used for applying the PPP, 74 L, was not sufficient to fully replenish this lower level, resulting in a low overall drainage percentage. The drainage pattern was closely followed and the observed pattern used to construct the drainage file for this day.

The volumes of water draining from the slabs were measured for a part of the experimental period only and assigned to the appropriate drainage periods. For the other periods, i.e. periods without measured drainage volume, it was assumed that 35% drainage was achieved (approximately the target value).

Temperatures were recorded at three different locations in the slabs. The average temperature as a function of time was used in the calculations.

4 Interpretation of the concentrations

4.1 Applied amounts of PPP

Nominal concentrations in the application solution differ from measured concentrations and therefore there is uncertainty with respect to the amounts effectively supplied to the system (see also Table 4-1):

- For dimethomorph, the nominal concentration (113 mg/L) was approximately five times the measured concentration (average of measurements in the application solution before and after the application is 21.3 mg/L). The reason for this difference may be due to exceeding the solubility of dimethomorph in water. The solubilities of the E- and the Z-isomer are 47.2 and 10.7 mg/L respectively, so below the nominal concentration.
- For fluopyram, nominal and measured concentrations in the application solution differ by approximately 10%. The average measured concentration (27 mg/L) is about twice the solubility of the substance in water (15 mg/L).
- For imidacloprid, the average measured concentration is 29.3 mg/L, which is about twice the nominal concentration.

The variation coefficients of the measurements of the concentrations in the application solution are 14%, 11% and 26% for dimethomorph, fluopyram and imidacloprid, respectively; this is high for these concentration levels.

Concentrations greater than the water solubility may be caused by formulation additives which enhance the dispersion of the substance in the application solution. Neither, the identity of the formulation additives, nor the behaviour of these additives in the system was known. It is therefore unclear for how long and where concentrations above water solubility can occur.

Given the uncertainty in the applied amounts, two approaches were followed in the simulations (section 4.3): one in which the applied amounts were based on the measured concentrations of the PPPs (MC approach), and one in which the applied amounts were based on the nominal concentrations of the PPPs in the application solution (WA approach), i.e. concentrations derived from weighted amounts.

Table 4-1 Amounts and concentrations of PPP applied to the system. In the MC approach, the mass applied is based on the measured concentrations in the application solution. Variation coefficients are given in between brackets. In the WA approach, the mass applied is derived from the weighted amounts.

	MC approach		WA approach	
	mg	mg/L	mg	mg/L
dimethomorph	1574 (14%)	21.3	8358	113
fluopyram	2004 (11%)	27.0	2273	30.7
imidacloprid	2172 (26%)	29.3	1123	15.1

4.2 Measured concentrations in the system

Figure 4-1 shows the averages of the concentrations of the active substances of the PPPs and their ranges in water flowing from the drippers towards the cultivation, in time. The range in concentrations was not only caused by variability due to the measurement method, but also to the response time of the drippers to changes in concentrations in the mixing tank. The drippers at the front end are closer to the mixing tank and therefore respond more quickly to changes in concentrations in the mixing tank than drippers in the middle and back. After application of the PPP solution, times for the PPP solution to arrive were 80, 130 and 190 seconds for the front, middle and back drippers respectively (see Appendix 3). The differences in response time are ignored in the figures in this report. Concentrations of all three substances declined rapidly, average concentrations at the second sampling time ($t = 1$ h) were at least one order of magnitude below the initial concentrations. Fresh nutrient solution replaced the contents of the tubes. The total volume of the tubes is smaller than the amount of water delivered during one watering event. However, due to the differences in response time because of the positions of the drippers in the greenhouse, a high variation in concentration was observed, especially in the first few hours after the application. Imidacloprid declined most rapidly, followed by fluopyram; dimethomorph declined most slowly. After the first few hours, the behaviour of fluopyram and imidacloprid was similar. The difference during the first few hours was possibly caused by fluopyram being slightly above its ideal solubility level. Dimethomorph declined more slowly during the first day and had a fairly constant concentration around 1 mg/L thereafter.

The lowest concentrations for fluopyram and imidacloprid were observed at $t = 29.1$ h. Concentrations of these two substances increased after that moment because the drainage water containing the substances was recycled to the mixing tank (see Appendix 3).

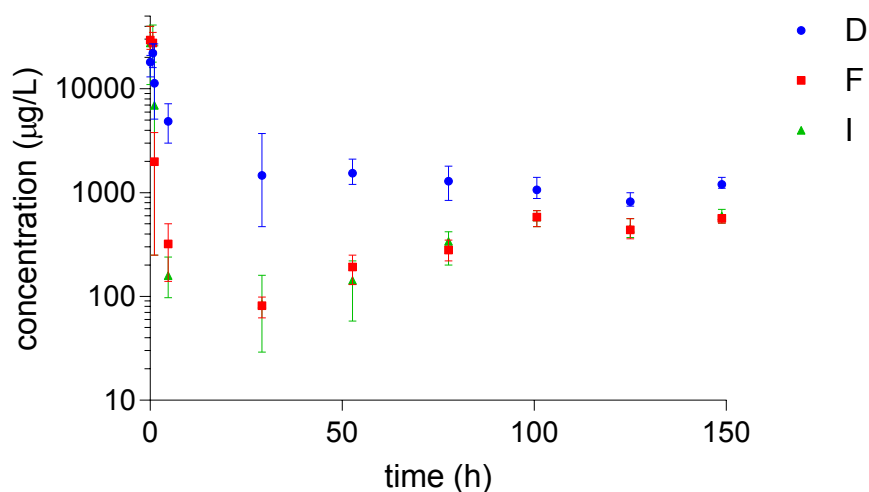


Figure 4-1 Measured concentrations of dimethomorph (D), fluopyram (F) and imidacloprid (I) in water sampled from the drippers. The error bars indicate the range of concentrations. In many cases, there is overlap in ranges observed for fluopyram and imidacloprid.

Concentrations of the three substances in the drain water stayed well below the solubilities. All three substances were present in the first drain water sample after application, but concentrations were at background level. The following samples had concentrations at least one order of magnitude above the background level. After the second sampling point in time, concentrations of imidacloprid increased most quickly, followed by fluopyram and dimethomorph. Concentrations of imidacloprid declined after 53 h and those of fluopyram after 78 h. Dimethomorph concentrations increased until 78 h and stayed fairly constant thereafter.

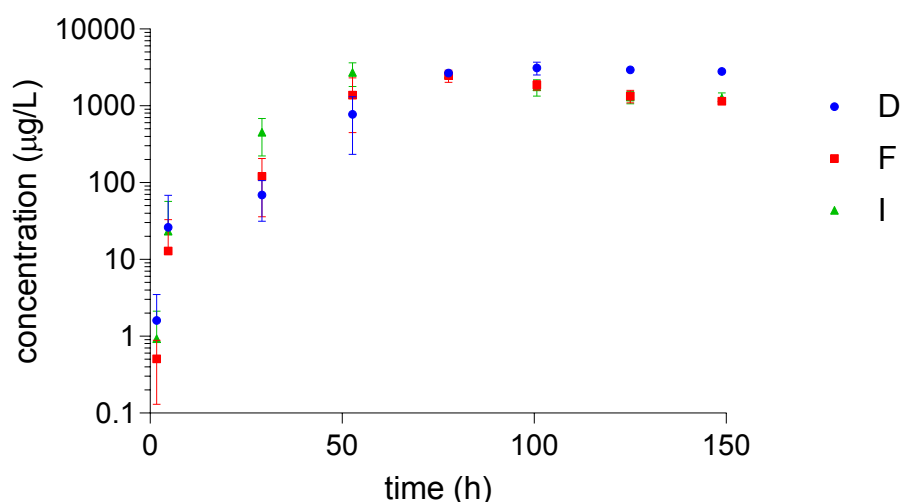


Figure 4-2 Concentrations of dimethomorph (D), fluopyram (F) and imidacloprid (I) in samples taken from the troughs. The error bars indicate the range of measured concentrations. In many cases, there is an overlap in ranges observed for the three substances, mostly for fluopyram and imidacloprid.

Figure 4-3 to Figure 4-5 show the concentrations in water sampled from the drippers and in the drain tube for the individual substances as a function of time. The concentrations in the drain tube were higher than the concentrations in the water sampled from the drippers after 29 h (imidacloprid), 53 h (fluopyram) and 78 h (dimethomorph). The lower concentrations in water sampled from the drippers, as compared to the concentrations in the drain tube, may have been caused by both dilution with fresh water added to the mixing tank (see Figures 3-1 and 4-6) and the uptake factor (TSCF) of the substances being less than one. However, the fact that the concentrations of the three substances in the water draining from the cultivation system in the second half of the experiment were consistently at least a factor two higher than in the water flowing towards the cultivation system, can only be explained by the TSCF being substantially less than one.

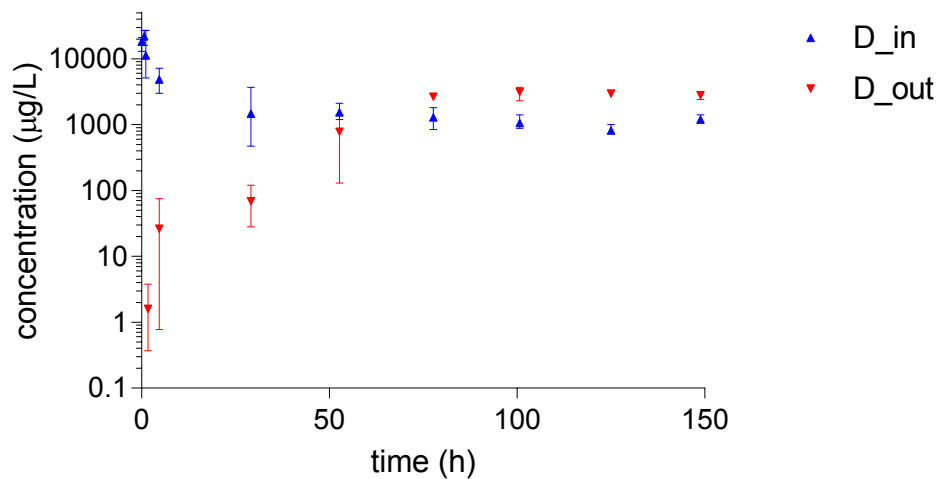


Figure 4-3 Measured dimethomorph concentrations in water sampled from the drippers (D_in) and water sampled from the troughs (D-out). The error bars indicate the range of measured concentrations.

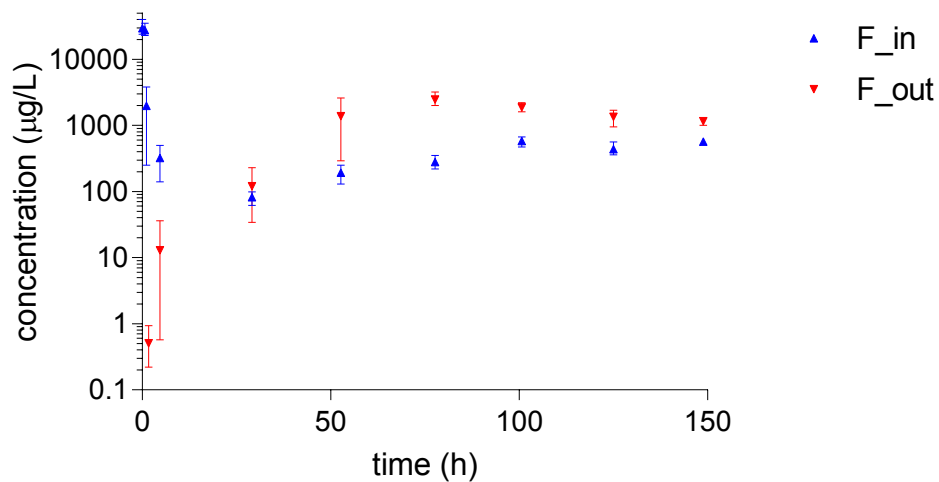


Figure 4-4 Measured fluopyram concentrations in water sampled from the drippers (F_in) and water sampled from the troughs (F_out). The error bars indicate the range of measured concentrations.

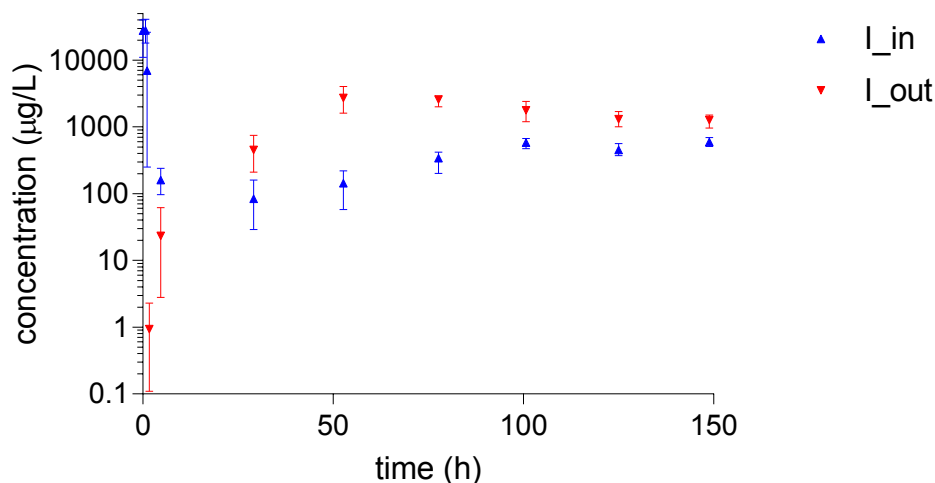


Figure 4-5 Measured imidacloprid concentrations in water sampled from the drippers (I_{in}) and water sampled from the troughs (I_{out}). The error bars indicate the range of measured concentrations.

4.3 Simulations and comparison with measurements

4.3.1 Model input parameters and simulated water balance

Measurements showed that all three substances were present in the system before the PPPs were applied. As these background concentrations were low, at least four orders of magnitude below initial concentrations in the mixing tank, they were assumed zero in the calculations.

The Substance Emission Model modified for this experiment requires as input: the lay-out of the experiment, the water flows between the different units as a function of time (see previous chapters), the applied amount of PPP and the temperature as a function of time and place (see chapter 2). It was assumed that the measured temperatures in the slabs also represent temperatures of the other elements.

The following dissipation processes are included in the SEM model for drip-application: plant uptake, degradation in the water in the various tanks, and discharge from the system. No discharge occurred during the experiment. The reference half-lives of the substances, i.e. the half-life in water at 20 °C, were assumed applicable to the whole system. However, in absence of information on degradation of the substances in the system, in most of the simulations a reference half-life of 1000 days (24000 h) was assumed for all three substances, indicating negligible degradation during the experimental period. A default Arrhenius activation energy of 65400 J/mol was assumed for all three substances. When the degradation is negligible, the value chosen for the activation energy has a negligible effect on the calculated concentrations.

SEM assumes passive uptake of the substances by plants, but influenced by the octanol water partition coefficient according to Briggs et al. (1982). Octanol water partition coefficients were taken from the dossiers (see Appendix 4). The calculated TSCF values were 0.58 (dimethomorph E-isomer), 0.54 (dimethomorph Z-isomer), 0.30 (fluopyram) and 0.43

(imidacloprid). The expected plant uptake is thus relatively high for dimethomorph and relatively low for fluopyram. All three TSCF values indicate that relative enrichment of the substances in the cultivation will occur, in increasing order dimethomorph, imidacloprid and fluopyram.

Figure 4-6 shows the cumulative water fluxes from the mixing tank to the cultivation, from the cultivation to the crop, from the cultivation back to the mixing tank, and the water contents of the mixing tank, across time. The cumulative fluxes and the fresh water supply were calculated from the information given in section 3.2. Total fresh water supply to the mixing tank was calculated at 1200 L, i.e. 3 times 400 L after 48 h, 92 h and 121 h. The total water delivery to the cultivation system was 2230 L plus the volume of the application solution. Evaporation amounted to 1592 L, and 658 L drained from the cultivation back to the mixing tank.

As can be seen in Figure 4-6, the movement of water occurs during day time, and no water movement is simulated during the night (horizontal line segments). After approximately 92 h, a volume equal to the average wet contents of the slabs is supplied to the slabs. In the second half of the experiment, growing conditions were better than in the first half, resulting in a higher water demand of the crop. The pattern of water flows influences simulated concentrations of the PPPs (see following sections); at night, no water flows in the tubes and troughs.

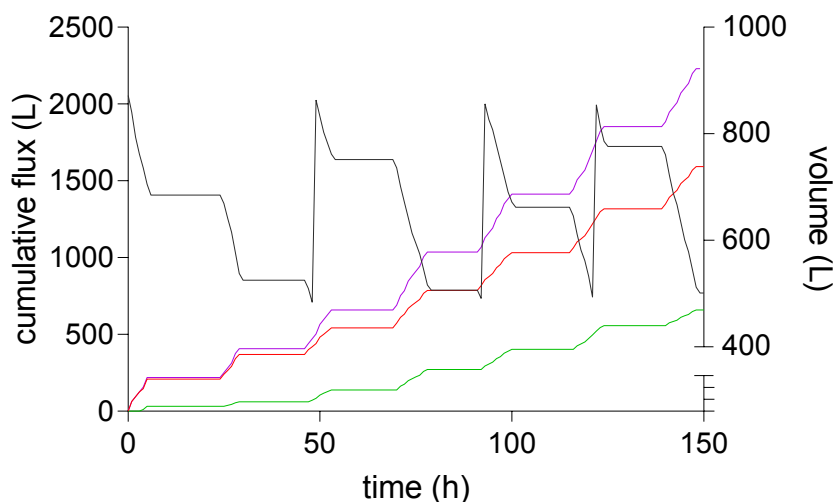


Figure 4-6 Calculated cumulative water fluxes (left axis) and water content of the mixing tank (right axis) during the experiment. Purple: water supply to the cultivation system, red: water taken up by the crop, green: water drained from the cultivation system, black: water content of the mixing tank.

Due to the uncertainty about the amounts of PPP applied to the system (section 4.1), two approaches were followed for the simulations. In the first (approach MC), the amounts of substances applied were calculated from application volume and the measured concentrations in the application solution. In the second (approach WA), the amounts of substances applied were calculated from the application volume and the

nominal concentrations in the application solution. Results of both approaches are discussed in the following sections.

4.3.2 *Dimethomorph*

Figure 4-7 shows measured (dots) and simulated (solid lines) dimethomorph concentrations in the nutrient solution sampled from the water flowing towards the cultivation, in time. The model simulates a drop in the concentration of approximately three orders of magnitude in the first few hours after the start of the experiment. The water in the tubes is refreshed several times with water containing background concentrations. The drop in measured concentrations is less than one order of magnitude. The differences between calculated and measured concentrations are largest at the last sampling time on the day of application, i.e. 5 h after the start of the experiment.

On the second day, simulated dimethomorph concentrations in the tubes started to increase. This is due to incoming water with higher concentrations than the background concentrations, as a result of drainage water containing dimethomorph being returned to the mixing tank. Concentrations increase during the day on days 1 to 3, after which the concentrations level at approximately 0.3 mg/L (approach MC) respectively 1.0 mg/L (approach WA). In contrast, measured concentrations continued to decline for the first 4 days of the experiment, reaching the lowest concentration of approximately 1 mg/L. Hardly any influence of the water dynamics could be observed in the measurements. From 78 h after the start onwards, simulated concentrations according to the WA approach are close to the ranges observed in the measurements.

The difference between measured and simulated concentrations in the first few days cannot be explained with the available information. A possibility is that some dimethomorph settled in solid form on the wall of the tubes and was released back into the solution once the concentrations dropped below solubility.

Dimethomorph dissipated in the simulations from the system almost exclusively via uptake by the crop. At the end of the experiment, 54% of the applied amount was taken up by the crop. At the same time, only 0.36% of the applied amount had degraded. The negligible effect of degradation was expected because of the assumed dimethomorph half-life of 1000 days. The potential effect of degradation is discussed in section 4.3.4.

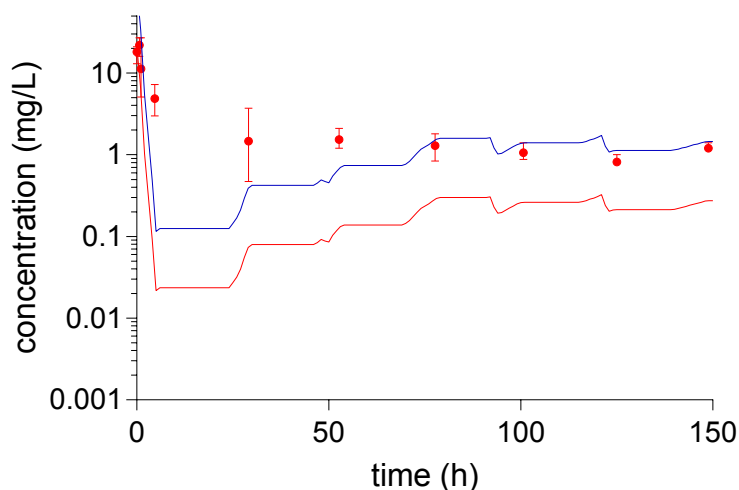


Figure 4-7 Measured (dots) versus calculated (lines) dimethomorph concentrations in water sampled from the drippers. The bars indicate the range in measured concentrations. Red line: MC approach, blue line: WA approach.

Figure 4-8 gives measured (dots) and simulated (solid lines) dimethomorph concentrations in the solution draining from the cultivation, in time. The model simulates a fast rise in concentrations shortly after the start of the experiment, from the moment draining starts. Measured concentrations also increased shortly after the start of the experiment, but slower than the simulated concentrations. In the MC approach, simulated concentrations are above measured concentrations for the first two days, and below measured concentrations afterwards. The pattern of measured concentrations was not observed in the simulations. Simulated concentrations are above measured concentrations in the WA approach for the first three days, but match afterwards. The WA approach appears to reflect measurements better than the MC approach.

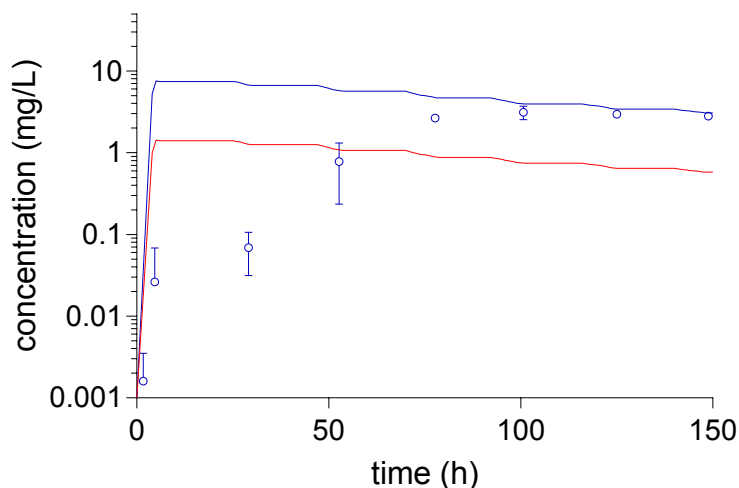


Figure 4-8 Measured versus calculated dimethomorph concentrations in water sampled from the troughs. The bars indicate the range in measured concentrations. Red line: MC approach, blue line: WA approach.

4.3.3

Fluopyram

Figure 4-9 gives measured (dots) and simulated (solid lines) fluopyram concentrations in the nutrient solution flowing towards the cultivation, in time. As seen for dimethomorph, the model simulates a drop in the concentrations of approximately three orders of magnitude in the first few hours. The water in the tubes is refreshed several times with water not influenced by added PPPs, as drainage has not yet occurred. The drop in measured concentrations is about a factor 20, depending on the location in the system (see also Figure 2-6).

On the second day, simulated concentrations in the tubes start to rise. This is due to incoming water with higher concentrations than the background as a result of drainage water containing fluopyram being returned to the mixing tank. The simulated concentrations on the second day are in the range of the measured concentrations. Simulated concentrations rise slightly after the second day, except shortly after fresh water is added to the system at approximately 2, 4 and 5 days after the start of the experiment (see also Figure 4-6). In general, measured and simulated concentrations are similar; simulation results are within the range of the measurements in three of six sampling times after the first day. Differences between the two approaches, MC and WA, are small; initial amounts differ by 13%.

Like dimethomorph, fluopyram dissipates from the system in the simulations almost exclusively via uptake by the crop. At the end of the experiment, 34% of the applied amount is taken up by the crop. At the same time, only 0.42% of the applied amount had degraded. The negligible effect of degradation was expected because of the assumed half-life of fluopyram of 1000 days. The potential effect of degradation is further discussed in section 4.3.4.

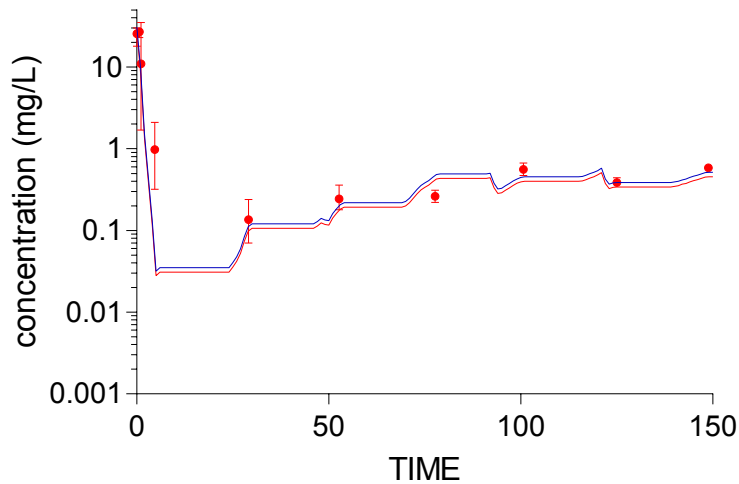


Figure 4-9 Measured (dots) versus calculated (line) fluopyram concentrations in water sampled from the drippers. The bars indicate the range in measured concentrations. Red line: MC approach, blue line: WA approach.

Figure 4-10 gives measured and simulated fluopyram concentrations in the solution draining from the cultivation, in time. The model simulates a fast rise in concentrations shortly after the start of the experiment, from the moment drainage starts. Measured concentrations also rise shortly after the start of the experiment, but slower than the simulated concentrations. Simulated concentrations are above measured concentrations during the first and second day, but are within the range of measured concentrations on three out of five sampling times afterwards. In the drain water, differences between both approaches, MC and WA, are also small.

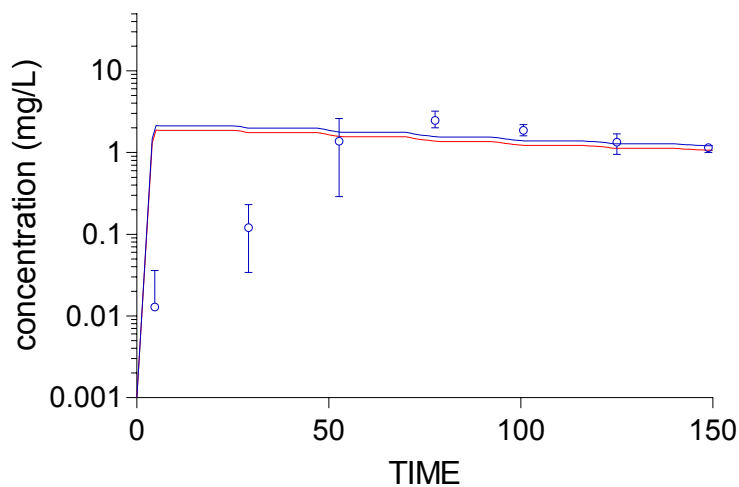


Figure 4-10 Measured (dots) versus calculated (line) fluopyram concentrations in water sampled from the troughs. The bars indicate the range in measured concentrations. Red line: MC approach, blue line: WA approach.

4.3.4 Imidacloprid

Figure 4-11 shows measured (dots) and simulated (solid lines) imidacloprid concentrations in the nutrient solution flowing towards the cultivation, in time. The model simulates a drop in the concentration slightly lower than three orders of magnitude in the first few hours. The water in the tubes is refreshed several times with water not influenced with added PPPs, as drainage has not yet occurred. The drop in measured concentrations is about two orders of magnitude depending on the location in the system (see also Figure 2-7), so slightly lower than simulated. On days 2–4, simulated concentrations in the tubes are within the range observed in the measurements, in both the MC and the WA approach. Later on in the experiment, simulated concentrations are slightly below observed concentrations in both approaches, but in the MC approach they are closer to the measurements than in the WA approach.

As with dimethomorph and fluopyram, imidacloprid dissipates from the system almost exclusively via uptake by the crop. At the end of the experiment, 45% of the applied amount is taken up by the crop. At the same time, only 0.38% of the applied amount has degraded. The negligible effect of degradation was expected because of imidacloprid's assumed half-life of 1000 days. The potential effect of degradation is discussed in section 4.3.4.

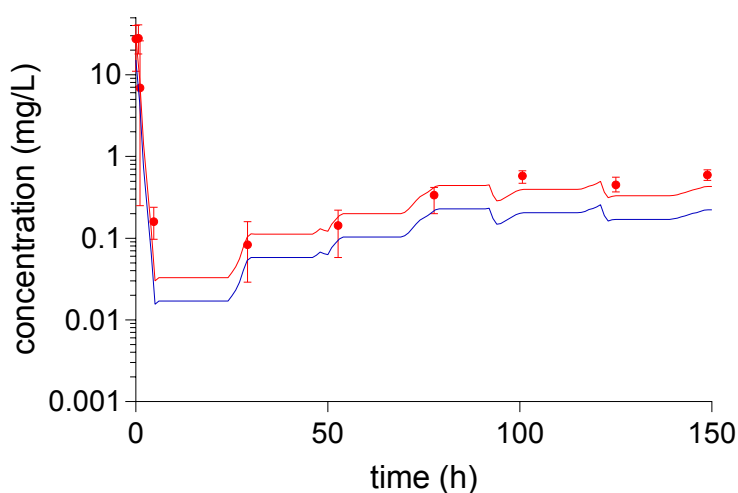


Figure 4-11 Measured (dots) versus calculated (line) imidacloprid concentrations in water sampled from the drippers. The bars indicate the range in measured concentrations. Red line: MC approach, blue line: WA approach.

Figure 4-12 shows measured (dots) and simulated (solid lines) imidacloprid concentrations in the solution draining from the cultivation, in time. As for dimethomorph and fluopyram, the model simulates a fast rise in concentrations shortly after the start of the experiment, from the moment draining starts. Measured concentrations also rise shortly after the start of the experiment, but slower than the simulated concentrations. Simulated concentrations are above measured concentrations in the first and second day for both approaches. After the second day, simulated concentrations are similar to the ranges of the measurement on 4 out 5 days (MC approach) but slightly below measured concentrations at all

sampling times in the WA approach. The MC approach appears to reflect measurements better than the WA approach.

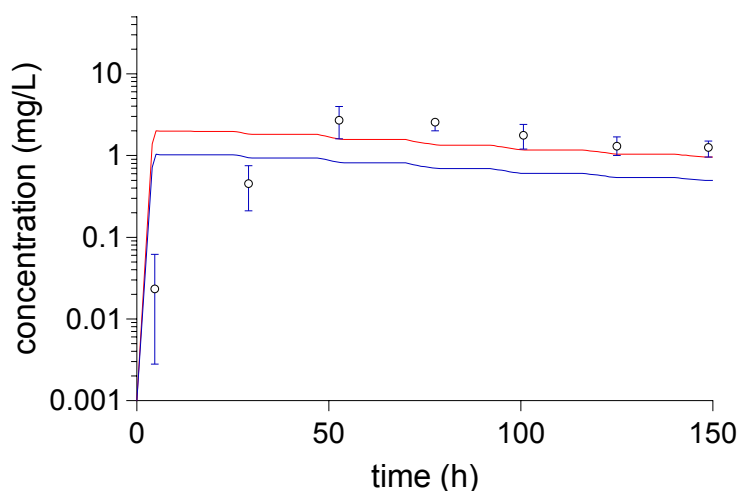


Figure 4-12 Measured (dots) versus calculated (line) imidacloprid concentrations in water sampled from the troughs. The bars indicate the range in measured concentrations. Red line: MC approach, blue line: WA approach.

4.3.5 Influence of TSCF and DegT50

Simulations in the previous sections used TSCF values based on octanol-water partition coefficients using the Briggs formula (Briggs et al. 1982). For fluopyram, the calculated TSCF is 0.3. Figure 4-13 shows the influence of varying the TSCF from 0 to 1 on simulated concentrations in nutrient solution flowing towards the cultivation and Figure 4-14 on simulated concentrations in the drain tubes. Higher TSCF values result in more uptake of substance by the crop and therefore lower concentrations in the system. The simulations suggest that a value in the low range is more in line with the observations. The experiment should have been extended in time to determine an appropriate value for the TSCF. It is likely that the same conclusion can be derived for dimethomorph and imidacloprid, should those simulations be performed.

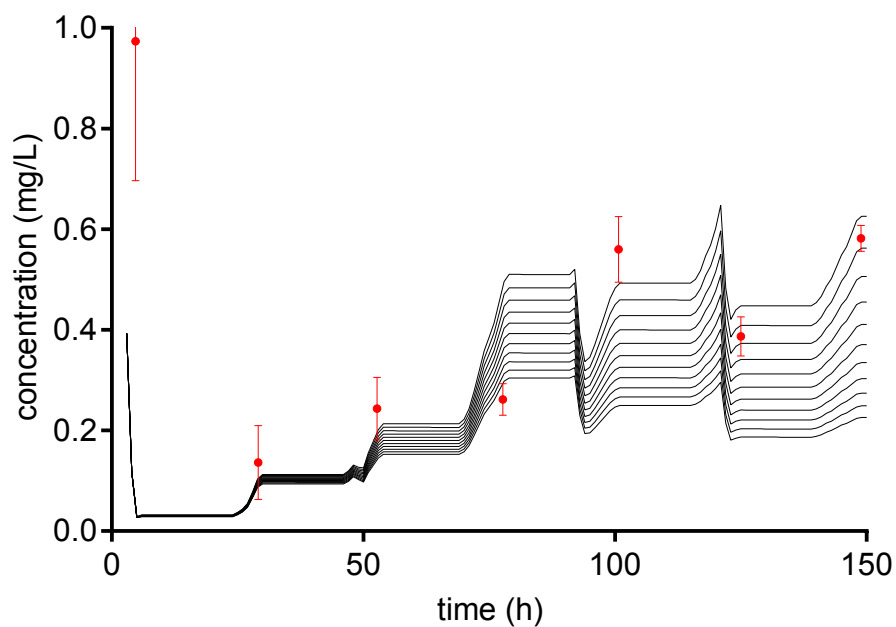


Figure 4-13 Influence of TSCF on simulated fluopyram concentrations in the nutrient solution flowing towards the cultivation. TSCF increases from 0 (top line) to 1 (bottom line) with increments of 0.1. The dots are the measured concentrations.

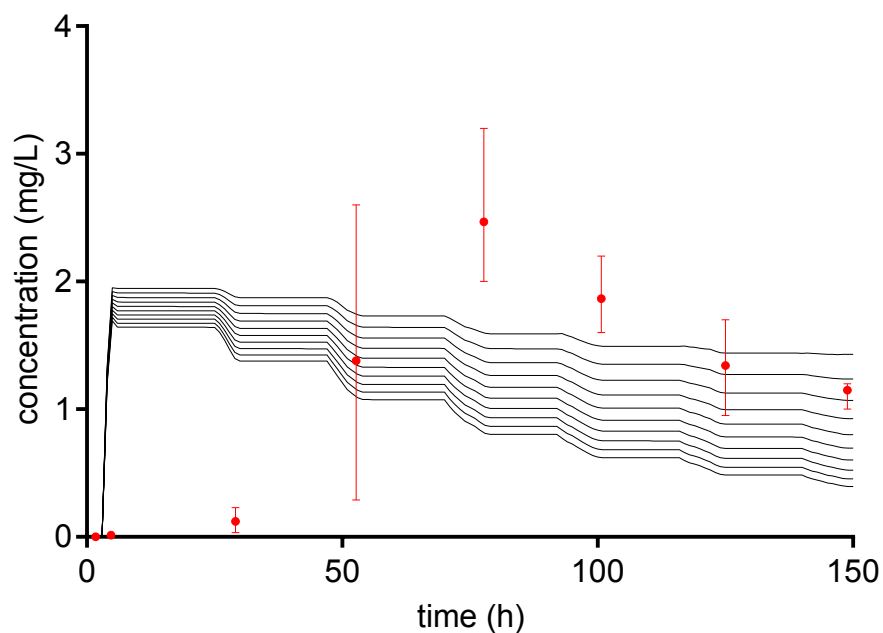


Figure 4-14 Influence of TSCF on simulated fluopyram concentrations in water draining from the cultivation. TSCF increases from 0 (top line) to 1 (bottom line) with increments of 0.1. The dots are the measured concentrations. Applied amount according to the MC approach.

The simulations in the previous sections assumed negligible transformation of the substances in the system; DegT50s were assumed to be 1000 d = 24000 h. Figure 4-15 and Figure 4-16 show the effect of shorter half-lives on simulated concentrations. With a half-life of 12 h, a substance disappears almost completely from the system within 6 days, i.e. the duration of the experiment. Results from simulations with half-lives above 200 h cannot be distinguished from each other in the graphs; the influence of degradation is negligible. The measurements indicate that degradation was indeed negligible during the experiments. Simulations (not presented) indicated that degradation is also negligible for imidacloprid. Similar simulations for dimethomorph were not performed.

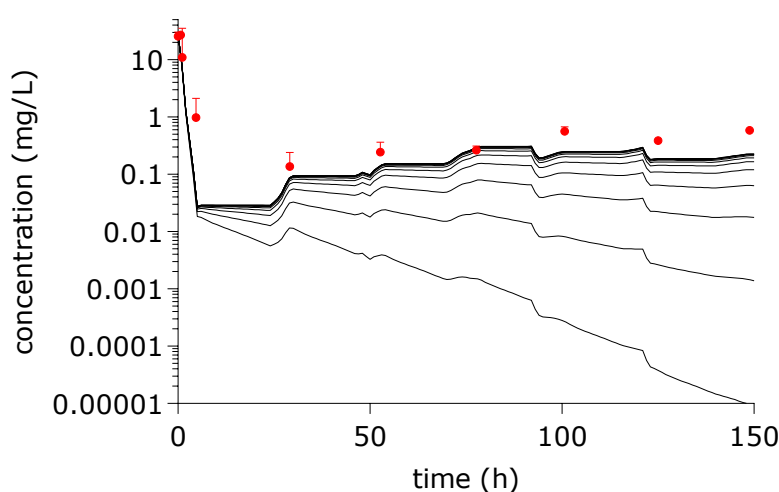


Figure 4-15 Influence of DegT50 on simulated fluopyram concentrations in nutrient solution flowing towards the cultivation. DegT50 decreases from 24000 h (top line) to 12 h (bottom line). The dots are the measured concentrations. Applied amount according to the MC approach.

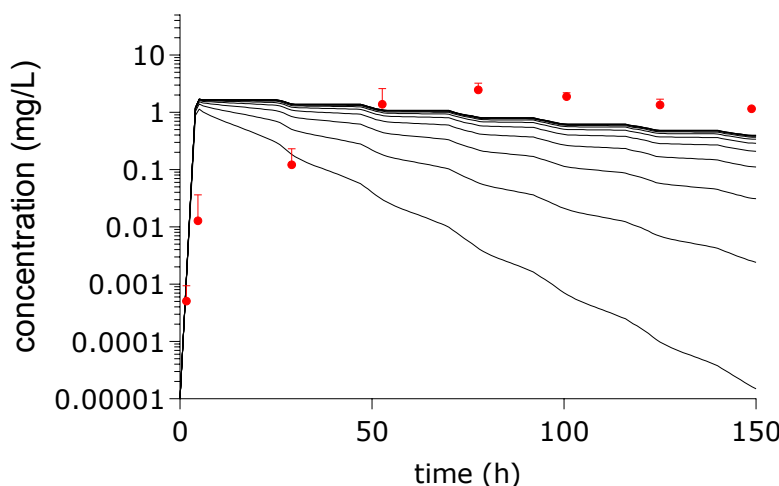


Figure 4-16 Influence of DegT50 on simulated fluopyram concentrations in the drain tubes. DegT50 decreases from 24000 h (top line) to 12 h (bottom line). The dots are the measured concentrations. Applied amount according to the MC approach.

4.4 Concentrations in relation to the cumulative drain water volume

Figure 4-17 shows the concentrations of dimethomorph, fluopyram and imidacloprid in relation to the cumulative amount of water that drained from the cultivation. As discussed in chapter 2, there is some uncertainty about the volume of water draining from the system; however, the amount of drainage is the same for the three substances. Fluopyram and imidacloprid show a similar pattern. Possibly imidacloprid is somewhat faster, but the ranges in measured concentrations are large and scaling to measured concentrations in the application solution would bring the curves closer to each other. More measurements shortly after the start of the experiment, i.e. up to a cumulative drain volume of 300 L in this experimental set-up, would have been necessary to be able to draw conclusions on differences between the substances. The pattern shown by dimethomorph is clearly different. As discussed earlier, this may be caused by exceeding its solubility in the application solution.

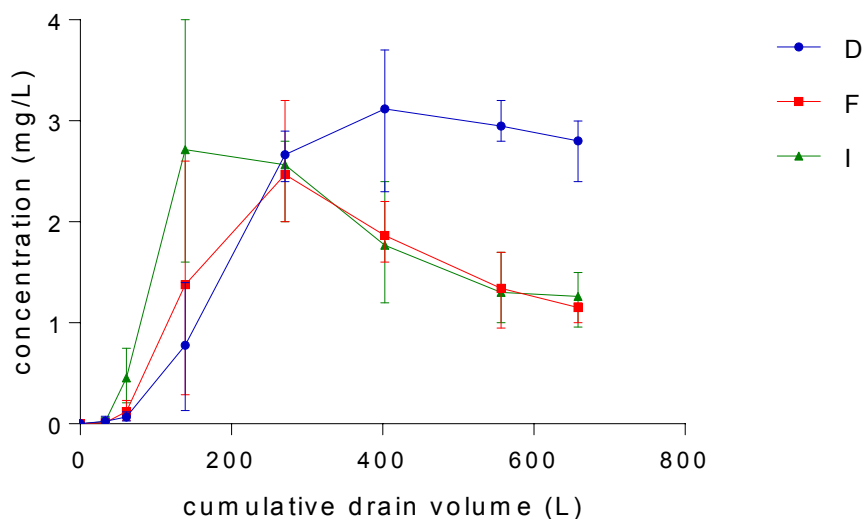


Figure 4-17 Concentrations of dimethomorph (D), fluopyram (F) and imidacloprid (I) in drain water in relation to the cumulative volume. The error bars indicate the ranges in measured concentrations.

5 Discussion and conclusions

One experiment, using three PPPs, was performed in order to test assumptions underlying the substance emission model (SEM) in GEM. Furthermore, the experiment was reviewed with the intention of improving the set-up for a new experiment. Improvements in the experimental set-up are discussed in chapter 6.

The experiment was performed in a standing cucumber crop in October 2014. As is apparent from greenhouse management registrations (for example Figure 2-2), the crop appeared to respond dynamically to changes in weather conditions, indicating that crop conditions during the experiment were acceptable.

The water input file used in the calculations was constructed based on registrations during the experiment. Unfortunately, drainage volumes were not measured for three days; target drainage values were used instead. This is only expected to have a limited effect on the results.

Major assumptions in SEM are that the solutions in the various parts of the system are well-mixed, that sorption in substrate systems with stone wool is absent, and that the dissipation of the substance is by plant uptake and degradation in the water phase. With respect to these three concepts, the following conclusions have been drawn.

Predicted and measured fluopyram and imidacloprid concentrations in water flowing to the cultivation and water draining from the cultivation compare well, except in drain water in the first 36-48 hours after application of the substances. In this period, predicted concentrations in the drain water are higher, indicating that mixing is not perfect (as is assumed in GEM) and displacement may occur to some extent. This indicates that emissions from the system will be overpredicted in the first few days after application if discharge of nutrient solution is necessary in that period. For dimethomorph, predicted and measured concentrations in both inflowing and draining water did not compare well for the first four days of the experiment. This was possibly due to concentrations exceeding the solubility, which is not accounted for in SEM. Further investigations are needed to determine whether it is worthwhile accounting for similar situations in SEM.

Degradation half-lives of the three substances appeared to be much longer than the duration of the experiment (six days). It is not possible to derive reliable degradation half-lives for the substances from this experiment. The concentrations are consistent with negligible degradation, which for the system is a DegT50 value above eight days, i.e. results are visually not distinguishable. Extending the experimental period would make it possible to derive longer reliable half-lives, but only to a limited extent because of competing dissipation processes, in this case plant uptake.

The disappearance of the three substances from the system can be attributed to plant uptake. The results clearly indicate the occurrence of

a concentration process, as concentrations in the drain water are consistently higher than in the inflowing water after the initial phase of the experiment. Based on the assumption that the plant uptake can be predicted from the octanol water partition coefficient (Briggs et al. 1982), calculated plant uptake was 54%, 34% and 45% of the applied amounts for dimethomorph, fluopyram and imidacloprid respectively, using transpiration stream concentration factors (TSCF) of 0.56, 0.3 and 0.43. The observed fluopyram concentrations in the system are consistent with a low value for the TSCF.

It cannot be concluded from this experiment whether or not sorption plays a role.

6 Recommendations for future experiments to validate GEM

In order to enlarge the validation status of the Greenhouse Emission Model – Application of PPP through drip irrigation, we recommend in future experiments to:

- Measure water contents of the mixing tank and the drain water tank continuously or on an hourly basis.
- Also measure water flows to and from the mixing tank and the drain water tank continuously or as frequently as possible. Detailed information about the water flows in the system is essential as the water flows are the driving forces of the substances in the systems.
- Calculate the expected concentrations, in course of time, before executing the experiment and develop a sampling scheme accordingly. In this way sampling moments that are crucial for model validation (such as breakthrough moments) can be determined beforehand.
- Apply, when possible, masses of PPP that lead to concentrations that are below the solubility; these might be lower than the recommended dosage.
- Try to treat all plants at the same time; i.e. avoid delays due to transport time in the system.
- Take water samples from the tanks, i.e. at least the mixing tank and the drain water tank, instead of from the tubes so that model adaptations can be avoided.
- Verify that substances/products can be used in a tank mixture if more than one substance is to be used in the experiment. Interactions between substances, for example precipitation or flocculation, may influence the delivery of substance to the crop with time. If substances can be used as tank mixtures, it may be better to add the substances/products separately to the mixing tank.

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Abbreviations and glossary

Cult	Cultivation system, consisting of stone wool slabs and plant roots
a.s.	active substance
drain tube	water pipes between cultivation and drain water tank
DWT	Drain Water Tank
GEM	Greenhouse Emission Model
MT	Mixing Tank
PPP	Plant Protection Product
PVC	Poly Vinyl Chloride, part of the tubes consists of this material.
SEM	Substance Emission Model, part of GEM
Tube	water pipes between mixing tank and cultivation
TOXSWA	TOXic Substances in Surface WAters
WATERSTREAMS	Model for calculating, a.o., water supply to, water distribution in and water emission from greenhouses.

Appendix 1 Instructions of use and solubility of applied substances

Label instructions as included in the dossier (www.ctgb.nl/toelatingen). Three substances were used in the experiment:

- dimethomorph (product: Paraat, 50 g/L wettable powder)
- fluopyram (product: Luna privilege, 500 g/L suspension concentrate)
- imidacloprid (product: Admire, 70% water dispersible granules).

The application rates for drip irrigation are included in Table A 1.

Table A 1 Label instructions and actual use of plant protection products in the experiment.

PPP	active substance	use instructions		actual use
		label	experimental conditions	
Paraat	dimethomorph	50–250 g / 1000 plants	13.8–69 g / 276 plants	37.5 g
Luna	fluopyram	40 ml / 1000 plants	11 ml / 276 plants	10.2 ml
Admire	imidacloprid	3.5–14 g / 1000 plants	1–3.9 g / 276 plants	3.6 g

Appendix 2 Substance Emission Model for application of PPP through drip irrigation in Berkeley Madonna

METHOD Euler

STARTTIME = 0

STOPTIME=150

DT = 0.02

DTout=1

{fixed length timestep}

{input section}

Vmtmin = 472 ; (L), lower level water volume in mixing tank

Vmtmax = 872 ; (L), upper level water volume in mixing tank

Vtube=26

Vcult =980 ; (L), water volume in substrate

Vdraintube = 25 ; (L)

Vdwt = 15 ; (L), water volume drain water tank

TSCF =0.26 ; TSCF Dimet=0.26, Fluo=0.30, Imid=0.47

Degt50mt = 24000 ; (h), = 1000 d

Degt50tube = 24000 ; (h), = 1000 d

Degt50cult = 24000 ; (h), = 1000 d

Degt50draintube = 24000 ; (h), = 1000 d

Degt50dwt = 24000; (h), = 1000 d

Tempdif = 0 ; (degrees C), difference in temperature between substrate and mixing tank

{Water part}

d/dt(Vmt) = refill - Wmt2tube + Wdwt2mt ; (L), water volume in mixing tank (variable in time)

Init Vmt = 872

refill = if (Vmt<Vmtmin) then pulse((Vmtmax-Vmt),DT,DT) else 0 ; (L), external water supply, triggered by Vmtmin

Wmt2tube = #W2cult(time) ; (L), water flow from mixing tank to substrate, read from file

Wtube2cult = #W2cult(time) ; (L)

Wdwt2mt = #W2drain(time) ; (L), water flow from drain water tank to mixing tank, read from file

W2crop = #W2crop(time) ; (L), water flow from substrate to crop, read from file

Wcult2draintube = #W2drain(time) ; (L), water flow from substrate to drainwater tank, read from file

Wdraintube2dwt = #W2drain(time) ; (L)

{Substance part}

Sbalans = Mmt + Mcult + Mdwt + M2crop + Mdeg + Mtube +
Mdraitube ; balance equation

{Percentage of mass in tanks, tubes, 2crop and degraded. Sbalans is
either 0 or mass applied.}

Pmt = if Sbalans > 0. then 100 * Mmt / Sbalans else 0

Pcult = if Sbalans > 0. then 100 * Mcult / Sbalans else 0

Pdwt = if Sbalans > 0. then 100 * Mdwt / Sbalans else 0

Pcrop = if Sbalans > 0. then 100 * M2crop / Sbalans else 0

Pdeg = if Sbalans > 0. then 100 * Mdeg / Sbalans else 0

Ptube = if Sbalans > 0. then 100 * Mtube / Sbalans else 0

Pdraitube = if Sbalans > 0. then 100 * Mdraitube / Sbalans else 0

d/dt(Mmt) = -Jmt2tube + Jdwt2mt - Jdegmt

Init Mmt = 0 ; (mg), substance mass in mixing tank

d/dt(Mtube) = +Jmt2tube - Jtube2cult - Jdegmtube

Init Mtube = 518 ; (mg), substance in tube, assumed added at t=0 to
the tube. Dim=518, Fluo=666, Imi=851

d/dt(Mcult) = Jtube2cult - Jcult2draintube - Jcult2crop - Jdegcult

Init Mcult = 1036 ; (mg), substance mass in substrate, Dim=1036,
Fluo=1332, Imi=1702

d/dt(Mdraintube) = Jcult2draintube - Jdraintube2dwt - Jdegdraintube

Init Mdraitube = 0 ; (mg) substance in draintube

d/dt(Mdwt) = Jdraintube2dwt - Jdwt2mt - Jdegdwt

Init Mdwt = 0 ; (mg), substance mass in drainwater tank

d/dt(M2crop) = Jcult2crop

Init M2crop = 0 ; (mg), substance mass taken up by crop

d/dt(Mdeg) = Jdegmt + Jdegcult + Jdegdwt + Jdegmtube + Jdegdraintube

Init Mdeg = 0 ; (mg), substance mass lost by degradation

{fluxes}

Jmt2tube = Wmt2tube * Cmt ; (mg), substance flow from mixing tank
to tube

Jtube2cult = Wtube2cult * Ctube ; (mg), substance flow from tube to
cultivation

Jcult2draintube = Wcult2draintube * Ccult ; (mg), substance flow from
substrate to draintube

Jdraintube2dwt = Wdraintube2dwt * Cdraitube ; (mg), substance flow
from draintube to drainwater tank

Jdwt2mt = Wdwt2mt * Cdwt ; (mg), substance flow from drainwater
tank to mxing tank

Jcult2crop = W2crop * Ccult * TSCF ; (mg), substance flow from
substrate to crop

Jdegmt = kdegmt * Mmt ; degradation rate in mixing tank

Jdegmtube = kdegmtube * Mtube ; degradation rate in tube

Jdegcult = kdegcult * Mcult ; degradation rate in substrate

Jdegdraintube = kdegdraintube * Mdraitube ; degradation rate in
draintube

Jdegdwt = kdegdwt * Mdwt ; degradation rate in drainwater tank

{concentrations}

Cmt = Mmt / Vmt ; (mg/L), concentration in mixing tank

$C_{tube} = M_{tube} / V_{tube}$; (mg/L)
 $C_{cult} = M_{cult} / V_{cult}$; (mg/L), concentration in substrate
 $C_{draintube} = M_{draintube} / V_{draintube}$; (mg/L)
 $C_{dwt} = M_{dwt} / V_{dwt}$; (mg/L), concentration in drainwater

{reaction rate constants}

$k_{degmt} = f_{Tmt} * \ln(2) / Degt50mt$; (d-1), degradation rate coefficient in mixing tank

$k_{deg tube} = f_{Ttube} * \ln(2) / Degt50tube$; (d-1), degradation rate coefficient in tube

$k_{degcult} = f_{Tcult} * \ln(2) / Degt50cult$; (d-1), degradation rate coefficient in substrate

$k_{degdrain tube} = f_{Tdrain tube} * \ln(2) / Degt50drain tube$; (d-1), degradation rate coefficient in draintube

$k_{degdwt} = f_{Tdwt} * \ln(2) / Degt50dwt$; (d-1), degradation rate coefficient in drainwater tank

$f_{Tmt} = \exp((-E_{act}/R_{gas}) * (1/(T_{mt}+T_0)-1/T_{ref}))$; (-), temperature correction factor in mixing tank

$f_{Ttube} = \exp((-E_{act}/R_{gas}) * (1/(T_{tube}+T_0)-1/T_{ref}))$; (-), temperature correction factor in tube

$f_{Tcult} = \exp((-E_{act}/R_{gas}) * (1/(T_{cult}+T_0)-1/T_{ref}))$; (-), temperature correction factor in substrate

$f_{Tdrain tube} = \exp((-E_{act}/R_{gas}) * (1/(T_{drain tube}+T_0)-1/T_{ref}))$; (-), temperature correction factor in draintube

$f_{Tdwt} = \exp((-E_{act}/R_{gas}) * (1/(T_{dwt}+T_0)-1/T_{ref}))$; (-), temperature correction factor in drainwater tank

{temperatures}

$T_{mt} = \#Temp(time) - Tempdif$; (degrees C), temperature in mixing tank

$T_{tube} = \#Temp(time)$; (degrees C), temperature in tube

$T_{cult} = \#Temp(time)$; (degrees C), temperature in substrate

$T_{drain tube} = \#Temp(time)$; (degrees C), temperature in draintube

$T_{dwt} = \#Temp(time)$; (degrees C), temperature in drainwater tank

{general constants}

$E_{act} = 65400$; (J/mol), Arrhenius activation energy

$R_{gas} = 8.314$; (J/(mol K)), gas constant

$T_{ref} = 293.15$; (K), reference temperature

$T_0 = 273.15$; (K), zero degrees Celsius

Appendix 3 Experimental diary (in Dutch)

Logboek meedruppelen middelen

Datum: 22 oktober 2014

Aanwezig: Marieke, Jim, Bram, Erik, op afroep Johan

Dinsdag 21 oktober

- Klaarzetten monsterflessen
- Stickers maken
- Maatbekers in kas klaarzetten
- Watergift om 12 uur stilgezet (Figuur1)
- Lampen opgehangen

Woensdag 22 oktober

- Maar bekers onder draingoot plaatsen
- Druppelaars in maatbekers plaatsen
- Afwegen middelen
 - imidacloprid (3.6 g),
 - dimethomorf (37.5 g),
 - pymetrozine (10.2 mL)
 - 6 g Fe-chelaat, FeEDDHA
- Matten markeren voor drain
- Achtergrond monsters genomen
- Spui oorspronkelijke voedingsoplossing uit voorraadbak in corridor; 47 cm: 455 liter
- Planten drainopvang, rij 3: 12 st; rij 9 11 st; rij 11 10 st en 2 afgeknipt.
- Al het water dat niet voor bemonstering nodig was, is op betreffende plaats teruggestort.



Giet-beurt	Start	Duur (min.s)	Eerste drupp	Midden drupp	Achterste drupp		Drain
			Erik	Bram	Jim		
1	9.55	6	Na 1.20 min	2.10	3.10	Roze kleuring drupp water	Geen drain
					5.00 laatste drupp	Gift 74 liter	
						164 liter aangemaakt (kleinste batch); 77 l afgevoerd	
2	11.02	2.30	Roze Ietsje drain	Roze Geen drain	Roze Laatste mat ietsje drain	Gift 30 l	
3	11.50	2.30	Wit	Wit	roze	Gift 33 l	Overall drain uit mat (heldere = wit opl) Per goot een beetje drain
4	13.15	2.30	2 drupp Code: 13.1 rij 11 erik	2 drupp Code: 13.2 rij 9 bram	2 drupp Code; 13.3, rij 3 Jim	Alles wit	Handstart gegeven om voldoende drain te krijgen. Ca. 1 liter in totaal

- Na 13.30 uur gift op automatisch. Totaal 7 beurten, zie Figuur. 1 en 2

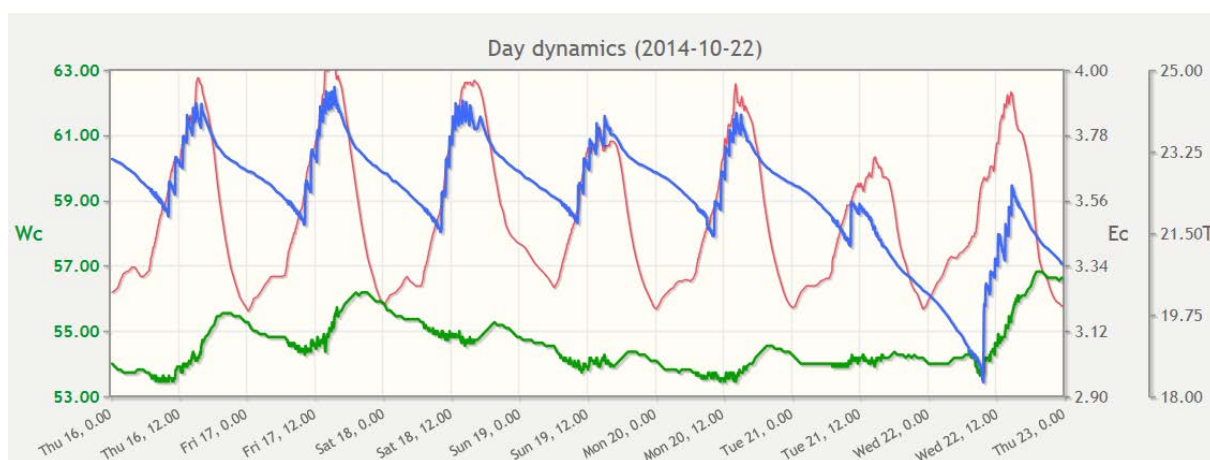


Figure C - 1 overzicht incl 21 okt (stilzetten gift) en 22 okt (uitvoering proef)

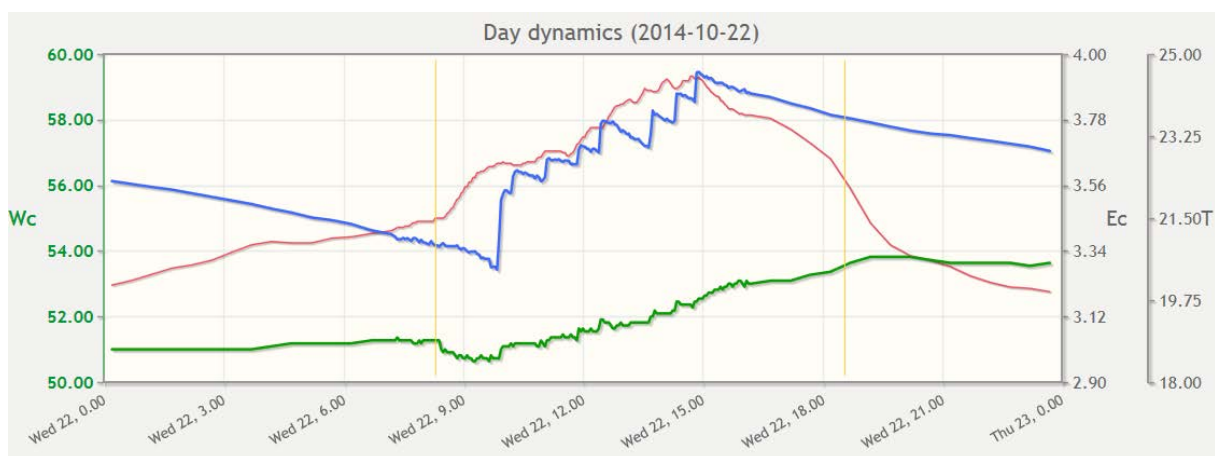


Figure C - 2 Watergeefbeurten op 22 okt in reguliere kas (6.10)



Figure C - 3 overzicht gietbeurten 1 (links), 2 (midden), 3 rechts). 1^e rij voorste druppelaars; 2^e rij middelste drupp; 3^e rij achterste drupp. In 1^e gietbeurt is voor en midden rood en achteraan nog roze; in 2^e gietbeurt is voor en midden alweer roze en achter nog rood; in 3^e gietbeurt is voor en midden weer helder (wit) en achter nog roze.

- Einde van de dag gift en drainmonsters verzameld
- 2 matten uit rij 6 en 10 verzameld voor monsters folie (boven en onder); planten (vruchten en blad apart), 4 st; steenwol 2 blokken van 10 cm per mat; wortels onder aan mat (2 per mat)
 - o Voor organische stof in mat 5 monsters per mat naar Chris en 2 stukken nieuwe mat (zelfde batch); Figuur 4

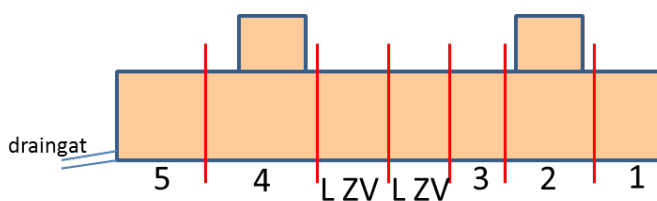


Figure C - 4 indeling mat in blokken en nummering voor organische stofbepaling.



Figure C - 5 links: draingat zit links. Folie gescheiden in onder en boven (met plantgaten); midden: in blokken gesneden; rechts: monsterflesjes GAC



Figure C - 6 plantmonster

Donderdag 23 oktober

- Strategie watergift aangepast om 10.30 uur, omdat er nog geen beurt gegeven was. Eerst een handstart, daarna elk uur een watergift tot vier uur voor zonsondergang.
- Gerard van den Broek (weekenddienst) laten zien welke monsters er genomen moeten worden op zondagochtend.
- 15.30 uur (na de laatste gietbeurt) gift en drainmonsters verzameld. Roze kleur is niet meer terug te vinden.

Vrijdag 24 oktober

- Strategie watergift is door Johan wederom aangepast, navragen wat er nu precies is gedaan.
- Monsternamen 17.15 uur (na de laatste gietbeurt)

Appendix 4 Substance properties

Substance properties (source Ctgb.nl, accessed 24-08-2016)

	dimethomor ph E[@]	dimethomor ph Z[@]	fluopyram	imidacloprid
molar mass (g/mol)	387.9	387.9	396.72	255.7
vapour pressure (Pa)	9.7E-7 (25 °C)	1.0E-6 (25 °C)	1.2E-6 (20 °C)	4E-10 (20 °C)
solubility (mg/L)	47.2	10.7	15	613
log Pow	2.63 (20 °C)	2.73 (20 °C)	3.3 (20 °C)	0.57
hydrolysis	stable	stable	stable	stable (pH 5-7), DT50 1 y (pH 9)
photolysis DT50 (d)	28-107 (22 °C, pH5)	28-107 (22 °C, pH5)	57-89 (25 °C)	0.3-0.4 (summer) 3.1-6.7 (autumn)
DegT50 soil lab (d)	56.7 [#]	56.7 [#]	-	
DegT50 soil field (d)	17-38	34-78	120	
DegT50 water (total system, d)	16 - 59	16 - 59	648 - 1470	21 (water) 1000 (sediment)
Kom (L/kg)	841 [#]	841 [#]	480	123
1/n	0.81-0.92 [#]	0.81-0.92 [#]	0.827	0.783

[@] substance under re-evaluation, new end-points not yet agreed

[#] for mixture of E- and Z-isomers

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