

Environmental risk limits for phenanthrene

RIVM Letter Report 601357007/2011 E.M.J. Verbruggen | R. van Herwijnen



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This investigation has been performed by order and for the account of Directorate-General for Environmental Protection, Directorate Environmental Safety and Risk Management, within the framework of the project 'International and National Environmental Quality Standards for Substances in the Netherlands' (INS)

Abstract

Environmental risk limits for phenanthrene

RIVM has derived environmental risk limits (ERLs) for phenanthrene. This derivation has been performed because the current ERLs have not been derived according to the current valid methodology. Phenanthrene is a substance belonging to the group of PAHs and is included in the Dutch decree on water quality objectives in the context of the Water Framework Directive (WFD). The ERLs in this report are advisory values that serve as a scientific background for the Dutch Steering Committee for Substances, which is responsible for setting those standards.

The maximum permissible concentration in water (MPC $_{water}$) is the level at which no harmful effects are expected, based on annual concentrations. This MPC is based on three routes: direct toxicity, secondary poisoning and consumption of fish by humans. Direct toxicity is the most critical of these three routes and determines the overall MPC for fresh- and saltwater (1.1 microgram per liter). The Maximum Acceptable Concentration (MAC $_{water, eco}$), that protects the ecosystem from effects of short term concentration peaks, is 6.7 microgram per liter for fresh- and saltwater.

The newly derived ERLs for water, suspended matter and sediment are higher than the currently valid ERLs. These differences are due to a more extensive dataset on ecotoxicology in combination with the more recent methodology for derivation of ERLs. Monitoring data indicate that the new MPC and MAC $_{\rm eco}$ for water, suspended matter and sediment are not being exceeded. In this observation, mixture toxicity for the total of PAHs has not been included.

Trefwoorden / Key words: environmental risk limits, maximum permissible concentration, negligible concentration, phenanthrene

Rapport in het kort

Milieurisicogrenzen voor fenantreen

Het RIVM heeft in opdracht van het ministerie van Infrastructuur en Milieu (I&M), milieurisicogrenzen voor fenantreen bepaald. Dit was nodig omdat de huidige norm voor fenantreen voor waterkwaliteit niet is afgeleid volgens de meest recente methodiek. Fenantreen is een stof die behoort tot de stofgroep PAK's. De stof is opgenomen in de Regeling Monitoring Kader Richtlijn Water, waarin staat aan welke eisen oppervlaktewater in Nederland moet voldoen. De Stuurgroep Stoffen stelt de nieuwe normen vast op basis van de wetenschappelijke advieswaarden in dit rapport.

Het Maximaal Toelaatbaar Risiconiveau (MTR) is de concentratie in water waarbij geen schadelijke effecten te verwachten zijn, gebaseerd op jaargemiddelde concentraties. Hiervoor zijn drie routes onderzocht: directe effecten op waterorganismen, indirecte effecten op vogels en zoogdieren via het eten van prooidieren, en indirecte effecten op mensen via het eten van vis. De eerste van de drie levert de laagste waarde en bepaalt daarmee het MTR voor zoet- en zoutwater (1,1 microgram per liter). De Maximaal Aanvaardbare Concentratie (MAC_{water, eco.)}, die het ecosysteem beschermt tegen kortdurende concentratiepieken, is 6,7 microgram per liter.

De nieuw afgeleide milieurisicogrenzen voor water, in water zwevend stof en sediment zijn hoger dan de nu geldende milieurisicogrenzen. Dit komt doordat er meer gegevens beschikbaar zijn over de directe effecten van fenantreen op waterorganismen en door het gebruik van een nieuwere methodiek. Gebaseerd op monitoringsgegevens worden de nieuwe MTR en MAC_{eco} voor water, zwevend stof en sediment naar verwachting niet overschreden. Bij deze beoordeling is de giftige aard voor het totaal aantal PAK's nog niet in beschouwing genomen.

Trefwoorden:

milieukwaliteitsnormen, milieurisicogrenzen, maximaal toelaatbaar risiconiveau, verwaarloosbaar risiconiveau, fenantreen

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Summary

Environmental risk limits are derived using ecotoxicological, physicochemical, and human toxicological data. They represent environmental concentrations of a substance offering different levels of protection to man and ecosystems. It should be noted that the ERLs are scientifically derived values. They serve as advisory values for the Dutch Steering Committee for Substances, which is appointed to set the Environmental Quality Standards (EQSs) from these ERLs. ERLs should thus be considered as preliminary values that do not have an official status.

This report contains ERLs for phenanthrene in water, groundwater, sediment and soil. The following ERLs are derived: Negligible Concentration (NC), Maximum Permissible Concentration (MPC), Maximum Acceptable Concentration for ecosystems (MAC $_{\rm eco}$), and Serious Risk Concentration for ecosystems (SRC $_{\rm eco}$). The risk limits were mostly based on data presented in the RIVM report "Environmental risk limits for polycyclic aromatic hydrocarbons (PAHs)" (Verbruggen, in prep.).

For the derivation of the MPC and MAC_{eco} for water and the MPC for sediment, the methodology used is in accordance with the Water Framework Directive. For the derivation of ERLs for air, no specific guidance is available. However, as much as possible the basic principles underpinning the ERL derivation for the other compartments are followed for the derivation of atmospheric ERL. For the MPCs for soil, and the NCs and the $SRCs_{eco}$ in general, the guidance developed for the project 'International and National Environmental Quality Standards for Substances in the Netherlands' was used (Van Vlaardingen and Verbruggen, 2007). An overview of the derived environmental risk limits is given in Table 1. The MPCs for water and suspended matter are higher than the current EQSs, in which the routes secondary poisoning and fish consumption were not included. The MPC for sediment is higher than the current EQS. These differences are due to a more extensive dataset on ecotoxicology in combination with the more recent methodology for derivation of ERLs.

Table 1 Derived MPC, NC, MAC_{eco}, and SRC_{eco} values for phenanthrene

ERL	unit	value			
		MPC	NC	MAC_{eco}	SRC_{eco}
freshwater ^a	μg.L ⁻¹	1.1	1.1 x10 ⁻²	6.7	43
freshwater susp. matter ^b	mg.kg _{dwt} -1	2.5			
drinking water human health ^c	mg.L ⁻¹	0.14			
saltwater	μg.L ⁻¹	1.1	1.1 x10 ⁻²	6.7	43
saltwater susp. Matter	mg.kg _{dwt} -1	2.5			
freshwater sediment ^d	mg.kg _{dwt} -1	0.78	7.8 x10 ⁻³		63
saltwater sediment ^d	mg.kg _{dwt} -1	0.78	7.8 x10 ⁻³		63
soil ^e	mg.kg _{dwt} -1	1.9	1.9 x10 ⁻²		90
groundwater	μg.L ⁻¹	1.1	1.1 x10 ⁻²		43
air	ma.m ⁻³	n.d.			

 $[^]a$ From the MPC $_{fw,\;eco}$, MPC $_{fw,\;secpois}$ and MPC $_{fw,\;hh\;food}$ the lowest one is selected as the 'overall' MPC $_{water}$

^b Expressed on the basis of Dutch standard suspended matter.

 $^{^{\}rm c}$ As stated in the new WFD guidance, the MPC $_{\rm dw,\,hh}$ is not included in the selection of the final MPC $_{\rm fw}$. Therefore, the MPC $_{\rm dw,\,hh}$ is presented as a separate value.

^d Expressed on the basis of Dutch standard sediment.

e Expressed on the basis of Dutch standard soil.

n.d. = not derived.

Monitoring data for phenanthrene indicate that currently the NC_{water} derived in this report is exceeded in all Dutch surface waters and that the $MPC_{susp, water}$ and $MPC_{susp, sw}$ are likely to be exceeded at many locations. Also, the $MPC_{sediment}$ could be exceeded in some cases and the $NC_{sediment}$ is likely to be exceeded in most cases. For this observation, the additive mixture toxicity for all PAHs has not been taken into account.

1 Introduction

1.1 Project framework

In this report environmental risk limits (ERLs) for surface water (freshwater and marine), soil, groundwater and air are derived for phenanthrene. Phenanthrene is listed in the Dutch decree on WFD-monitoring (*Regeling monitoring Kaderrichtlijn water*) as a specific pollutant. The aim of this report is to present updated risk limits that can be used to set water quality standards in accordance with the WFD. Phenanthrene is relevant for other compartments as well, therefore, ERLs for soil and air have also been derived. MPCs for direct ecotoxicity have already been derived by Verbruggen (in prep.). Additional ERLs, including those considering secondary poisoning and human health through fish consumption, are derived in this report. The derivation of the ERLs is performed in the context of the project National Policy on Substances. The following ERLs are considered:

- Maximum Permissible Concentration (MPC) defined in VROM (1999, 2004) as the standard based on scientific data which indicates the concentration in an environmental compartment for which:
 - 1 no effect to be rated as negative is to be expected for ecosystems;
 - 2a no effect to be rated as negative is to be expected for humans (for non-carcinogenic substances);
 - 2b for humans no more than a probability of 10^{-6} per year of death can be calculated (for carcinogenic substances). Within the scope of the Water Framework Directive (WFD), a probability of 10^{-6} on a life-time basis is used.

The MPCs for water and soil should not result in risks due to secondary poisoning (considered as part of the ecosystem in the definition above) and/or risks for human health aspects. These aspects are therefore also addressed in the MPC derivation. Separate MPC-values are derived for the freshwater and saltwater environment.

- Negligible Concentration (NC) the concentration at which effects to ecosystems are expected to be negligible and functional properties of ecosystems are safeguarded fully. It defines a safety margin which should exclude combination toxicity. The NC is derived by dividing the MPC by a factor of 100.
- Maximum Acceptable Concentration (MAC $_{eco}$) for aquatic ecosystems the concentration protecting aquatic ecosystems from effects due to short-term exposure or concentration peaks. The MAC $_{eco}$ is derived for freshwater and saltwater ecosystems.
- Serious Risk Concentration for ecosystems (SRC_{eco}) the concentration in water at which possibly serious ecotoxicological effects are to be expected. This value should be compared with the Serious Risk Concentration for humans (SRC_{human}), which is not derived elsewhere (Lijzen et al., 2001).
- Maximum Permissible Concentration for surface water that is used for drinking water abstraction (MPC $_{dw, hh}$). This is the concentration in surface

water that meets the requirements for use of surface water for drinking water production. The $MPC_{dw,\ hh}$ specifically refers to locations that are used for drinking water abstraction.

The quality standards in the context of the WFD refer to the absence of any impact on community structure of aquatic ecosystems. Hence, not the potential to recover after transient exposure, but long-term undisturbed function is the protection objective under the WFD. Recovery in a test situation, after a limited exposure time, is therefore not included in the derivation of the MPC and MAC.

1.2 Current MPCs

The current MPCs for phenanthrene are $0.3~\mu g.L^{-1}$ for water, $1~mg.kg_{dwt}^{-1}$ for suspended matter and $0.5~mg.kg_{dwt}^{-1}$ for sediment. The derivation of these values is reported by Kalf et al. (1995). For air there is an indicative MPC of $9.6~ng.m^{-3}$. Derivation of this value is described by Hansler et al. (2008).

1.3 Sources of phenanthrene

There is no production of phenanthrene as a pure product. Phenanthrene, like most other polycyclic aromatic hydrocarbons (PAHs), is however present in fossil fuels and derived products; human use of these products is one of the main sources of phenanthrene in the environment. Other important anthropogenic sources are industrial processes, such as iron steel works, coke manufacturing, asphalt production, wood preservation, ship protection and petroleum cracking. Most of these industries have however improved their processes or reduced or stopped the use of PAH containing products and current emissions are highly reduced as compared to the past. Apart from anthropogenic sources, there are also natural sources like vegetation fires and volcanic emissions.

1.4 Methodology

The methodology for risk limit derivation is described in detail in the INSguidance document (Van Vlaardingen and Verbruggen, 2007), which is further referred to as the INS-Guidance. The methodology is based on the Technical Guidance Document (TGD), issued by the European Commission and developed in support of the risk assessment of new notified chemical substances, existing substances and biocides (EC, 2003) and on the Manual for the derivation of Environmental Quality Standards in accordance with the Water Framework Directive (Lepper, 2005). The European guidance under the framework of WFD has been revised, and the updated guidance has been published recently (EC, 2011). The risk limits in this report will be used for setting water quality standards that will become effective after the new guidance has come in to force. Therefore, the terminology is harmonised as much as possible and the new guidance is followed in the case it deviates from the INS-guidance. This specifically applies to the derivation of the MAC (see section 3.5), for which the new methodology is used. This also holds for the MPC for surface waters intended for the abstraction of drinking water (MPC_{dw, hh}, see section 3.4). In the INS-guidance, this is one of the MPCs from which the lowest value should be selected as the general MPC_{water} (see section 3.1.6 and 3.1.7 of the INS-Guidance). According to the new guidance, the MPC_{dw, hh} is not taken into account for the derivation of the general MPC_{water} , but specifically refers to locations that are used for drinking water abstraction. For the derivation of ERLs for air, no specific guidance is available. However, as much as possible, the

basic principles underpinning the ERL derivation for the other compartments are followed for the derivation of an atmospheric ERL.

1.4.1 Data sources

The RIVM report "Environmental risk limits for polycyclic aromatic hydrocarbons (PAHs)" (Verbruggen, in prep.) is used as the source of physicochemical and (eco)toxicity data. Information given in this report is checked thoroughly and approved by the scientific committee of the project 'International and National Environmental Quality Standards for Substances in the Netherlands' (INS). Therefore, no additional evaluation of data is performed for the ERL derivation. Only valid data combined in an aggregated data table are presented in the current report. Occasionally, key studies are discussed when relevant for the derivation of a certain ERL. Since in the report of Verbruggen only ERLs for direct toxicity are reported, the additional ERLs to be derived according to the INS guidance are derived in this report.

1.4.2 Data evaluation

Ecotoxicity studies were screened for relevant endpoints (i.e. those endpoints that have consequences at the population level of the test species) and thoroughly evaluated with respect to the validity (scientific reliability) of the study. A detailed description of the evaluation procedure is given in section 2.2.2 and 2.3.2 of the INS-Guidance and in the Annex to the draft EQS-guidance under the WFD. In short, the following reliability indices were assigned, based on Klimisch et al. (1997):

Ri 1: Reliable without restriction

'Studies or data ... generated according to generally valid and/or internationally accepted testing guidelines (preferably performed according to GLP) or in which the test parameters documented are based on a specific (national) testing guideline ... or in which all parameters described are closely related/comparable to a guideline method.'

Ri 2: Reliable with restrictions

'Studies or data ... (mostly not performed according to GLP), in which the test parameters documented do not totally comply with the specific testing guideline, but are sufficient to accept the data or in which investigations are described which cannot be subsumed under a testing guideline, but which are nevertheless well documented and scientifically acceptable.'

Ri 3: Not reliable

'Studies or data ... in which there are interferences between the measuring system and the test substance or in which organisms/test systems were used which are not relevant in relation to the exposure (e.g., unphysiologic pathways of application) or which were carried out or generated according to a method which is not acceptable, the documentation of which is not sufficient for an assessment and which is not convincing for an expert judgment.'

Ri 4: Not assignable

'Studies or data ... which do not give sufficient experimental details and which are only listed in short abstracts or secondary literature (books, reviews, etc.).'

Citations

In case of (self-)citations, the original (or first cited) value is considered for further assessment, and an asterisk is added to the Ri of the endpoint that is cited.

All available studies are summarised in data-tables that are included as Annexes to this report. These tables contain information on species characteristics, test conditions and endpoints. Explanatory notes are included with respect to the assignment of the reliability indices. In the aggregated data table only one effect value per species is presented. When for a species several effect data are available, the geometric mean of multiple values for the same endpoint is calculated where possible. Subsequently, when several endpoints are available for one species, the lowest of these endpoints (per species) is reported in the aggregated data table.

1.5 Status of the results

The results presented in this report have been discussed by the members of the scientific advisory group for the INS-project (WK-INS). It should be noted that the ERLs in this report are scientifically derived values, based on (eco)toxicological, fate and physicochemical data. They serve as advisory values for the Dutch Steering Committee for Substances, which is appointed to set the Environmental Quality Standards (EQSs). ERLs should thus be considered as advisory values that do not have an official status.

Substance properties, fate human toxicology and trigger values

2.1 Identity

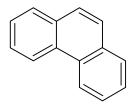


Figure 1. Structural formula of phenanthrene

Table 2. Identification of phenanthrene

rable 21 Identification of prichartement			
Parameter	Name or number		
Chemical name	phenanthrene		
Common/trivial/other name	phenanthrene; o-diphenyleneethylene		
CAS number	85-01-8		
EC number	201-581-5		
Molecular formula:	$C_{14}H_{10}$		
SMILES code	c12ccccc1c3cccc3cc2		

2.2 Physicochemical properties

Table 3. Physicochemical properties of phenanthrene from Verbruggen (in prep.)

Parameter	Unit	Value	Remark
Molecular weight	[g.mol ⁻¹]	178.23	
Water solubility	[µg.L ⁻¹]	1034	Geometric mean of seven values by the generator-column method
log K _{OW}	[-]	4.502	Geometric mean of three values by the slow-stirring method
log K _{OC}	[L.kg ⁻¹]	4.292	QSAR
Vapour pressure	[Pa]	0.018	Geometric mean of five values by the gas saturation method
Melting point	[°C]	99.2	-
Boiling point	[°C]	340	
Henry's law constant	[Pa.m ³ .mol ⁻¹]	3.8	Geometric mean of seven values by the gas stripping method, one by the headspace method and one by the wetted-wall method

2.3 Bioconcentration and biomagnification

Bioconcentration data (based on lab studies) and bioaccumulation data (based on field studies) for phenanthrene are given in Table 4. The data in this table are based on studies reviewed by Bleeker and Verbruggen (2009) according to the Ri classification of Klimisch et al. (1997) and considered reliable (Ri1 or 2). A full overview of these studies is given in Appendix 1

Table 4. Overview of bioaccumulation data for phenanthrene

Parameter	Unit	Value	Remark
BCF (fish)	[L.kg ⁻¹]	1750	Geometric mean of most reliable BCF value for for <i>Cyprinodon variegatus</i> (1149) and <i>Pimephales promelas</i> , all data normalized to 5% fat. Species geometric mean for <i>Pimephales promelas</i> was first calculated from five values.
BCF (molluscs)	[L.kg ⁻¹]	1260	Geometric mean of the BCF values for <i>Mya</i> arenaria and <i>Mytilus edulis</i> . These BCFs could not be normalized to 5% fat
BCF (crustaceans - human consumption)	[L.kg ⁻¹]	210	BCF for <i>Crangon septemspinosa</i> , this is the only BCF for a crustacean suitable for human consumption
BCF (crustaceans)	[L.kg ⁻¹]	648	Geometric mean of BCF values for all crustacean species. Species geometric mean was calculated first for <i>Diporeia</i> sp. Only one of the BCF values could be normalized to 5% fat
BCF (insects)	[L.kg ⁻¹]	1340	Geometric mean of all BCF values for <i>Hexagenia limbata</i> normalized to 5% fat
BCF (worms)	[L.kg ⁻¹]	1616	Geometric mean of the BCF values for Stylodrilus heringianus and Nereis virens. These BCFs could not be normalized to 5% fat
BCF (plants)	[L.kg ⁻¹]	30	Geometric mean of all BCF values for <i>Lemna</i> gibba not normalized to 5% fat
BMF	[kg.kg ⁻¹]	1	Biomagnification has not been observed (Nfon et al., 2008, Wan et al., 2007, Takeuchi et al., 2009) ^a

^a In a study into foodweb distribution of PAHs (Vives et al., 2005), a biomagnification factor of 1.5 kg.kg⁻¹ is reported. This value was based on phenanthrene concentrations in the fish diet and a fat normalized concentration in the fish liver. Since the phenanthrene concentration in the fish liver is in general higher than in the whole fish, this reported value supports the use of a BMF of 1.

A wide range of BCFs is available. In addition, BAFs are available. These BAFs (derived from field samples) indicate that the BCFs (derived from laboratory studies) are comparable to the bioaccumulation in the field. The BAF values are presented in Appendix 1. The low lipid content of most organisms from the field carries some additional uncertainty in comparison to the laboratory BCF values. Therefore, the BCF data normalized to 5% lipids will be used in the calculation of the MPCs for secondary poisoning of mammals and birds (MPCfw, secpois and MPC_{sw, secpois}) and the MPC for human food consumption (MPC_{water, hh food}). When deciding which BCF should be used for calculation of the MPCs for secondary poisoning of mammals and birds (MPCfw, secpois and MPCsw, secpois) and the MPC for human food consumption (MPC_{water, hh food}), it should be considered that humans have a more specific food choice (fishery products) than mammals and birds, for which diets can vary considerably amongst different species. Therefore different BCFs should be used when deriving the different MPCs. For the MPC_{water, hh food}, the relative human consumption of fish, molluscs and crustaceans is used to determine the BCF. The human food consumption pattern used to determine the BCF is based on the Dutch food consumption survey for 1998 (Anonymous, 1998). The relative consumption of fish, molluscs and crustacean is 90%: 7%: 3% for fish: molluscs: crustaceans. On the basis of this relative consumption, a weighted average is calculated from the BCFs for fish, molluscs and the crustacean that is suitable for human consumption. The calculated BCF is: 1664 L.kg⁻¹ based on geometric mean values for fish and molluscs, normalized to 5% lipids if possible. It should be noted that this approach is not the most conservative. A person having an equal daily consumption of fish only might not entirely be protected by this BAF. On the

other hand, the derivation of the MPC $_{\text{water, hh food}}$ is already precautionary for the general Dutch population, because of the relatively high daily intake (115 g fishery products) and the fact that the contribution of the consumption of fishery products to the total daily exposure is only 10%. Therefore, a large risk for such a person is considered unlikely.

For the BCF to calculate the $MPC_{fw, \, secpois}$, it is presumed that some predatory species have strong preference for the one of the three groups (fish, crustaceans or molluscs) or another group (e.g. worms) for their diet. The selected BCF for the $MPC_{fw, \, secpois}$ is the highest of the available groups and is the geometric mean of the BCF values for fish which is 1750 L.kg⁻¹.

2.4 Human toxicological threshold limits and carcinogenicity

Phenanthrene has not been classified in EU framework. The U.S. EPA (IRIS) concluded that phenanthrene is not classifiable for human carcinogenicity. RIVM concluded that phenanthrene is probably carcinogenic, but the relative carcinogenic potential is extremely low. For oral toxicity, a Tolerable Daily Intake (TDI) of 0.040 mg.kg $_{bw}^{-1}.day^{-1}$ was derived based on a threshold approach (Baars et al., 2001). This value is adopted as TDI in this report. For inhalation toxicity no individual TCA (Tolerable Concentration in Air) is available for phenanthrene. A limit value of 0.01 ng.m⁻³ has been proposed by the EU working group on PAHs (EC, 2001) for a lifetime exposure risk of 10⁻⁶ for benzo[a]pyrene (BaP) as indicator for the total PAHs and this value has been adopted in EU legislation (EU, 2004). To obtain an limit value for benzo[a]pyrene as an individual substance, the limit value can be increased with a factor of 10 (a factor that is used to estimate the risk of total PAHs on the risk of BaP only) to 0.1 ng.m⁻³. TCAs for other PAHs can be derived from this value on the basis of their relative carcinogenic potency. The relative carcinogenic potency of phenanthrene is considered to be negligible and has been set at <0.001 (Baars et al., 2001). With this value the TCA for phenanthrene will be > 0.1 µg.m⁻³, no bound value can be derived. With a daily breathing volume of about 10,000 L air, it is can be calculated that this value of 0.1 µg.m⁻³ would still be very stringent compared to the TDI.

2.5 Trigger values

This section reports on the trigger values for ERL_{water} derivation (as demanded in WFD framework) as reported in Verbruggen (in prep.).

Table 5. Phenanthrene: collected properties for comparison to MPC triggers

Parameter	Value	Unit	Method/Source
Log K _{p,susp-water}	3.29	[-]	$K_{\rm OC} \times f_{\rm OC,susp}^{\rm a}$
BCF	1664 / 1750 ^b	[L.kg ⁻¹]	
BMF	1	[kg,kg ⁻¹]	
Log K _{OW}	4.50	[-]	
R-phrases	n.a.	[-]	
A1 value	n.a.	[µg.L ⁻¹]	
DW standard	n.a.	[µg.L ⁻¹]	

 $^{^{}a} f_{OC,susp} = 0.1 \text{ kg}_{OC}.\text{kg}_{solid}^{-1} \text{ (EC, 2003)}.$

o phenanthrene has a log $K_{p, susp-water} > 3$; derivation of MPC_{sediment} is triggered.

 $^{^{\}rm b}$ Different BCF values are given to be used separately for calculation of the MPC $_{\rm water,\ hh\ food}$ and the MPC $_{\rm fw,\ secpois}$ respectively.

n.a. = not available.

- o phenanthrene has a log $K_{\rm p,\,susp-water}$ > 3; expression of the MPC as MPC as MPC is required.
- phenanthrene has a BCF > 100 L.kg⁻¹; assessment of secondary poisoning is triggered.
- o phenanthrene is only marginally carcinogenic but given the low TDI and the fact that the BCF is above 100 $L.kg^{-1}$, an MPC_{water} for human health via food (fish) consumption (MPC_{water}, hh food) should be derived.

3 Toxicity data and derivation of ERLs for water

3.1 **Toxicity data**

The selected freshwater toxicity data for phenanthrene as reported by Verbruggen (in prep.) are given in Table 6 and marine toxicity data are shown in Table 7.

Table 6. Phenanthrene: selected freshwater toxicity data for ERL derivation

Chronic	NOEC/EC ₁₀	Acute	L(E)C ₅₀
Taxonomic group	(µg.L ⁻¹)	Taxonomic group	(µg.L ⁻¹)
		Cyanophyta	
		Anabaena flos-aqua	1300
Algea		Algea	
Pseudokirchneriella subcapitata	15 ^a	Nitzschia palea	870
Scenedesmus vacuolatus	150	Pseudokirchneriella subcapitata	233 ^f
		Scenedesmus vacuolatus	590
Crustacea		Crustacea	
Ceriodaphnia dubia	13	Daphnia magna	700
Daphnia magna	18 ^b	Daphnia pulex	100
Daphnia pulex	13 ^c	Diporeia spp.	74 ^g
Hyalella azteca	155	Gammarus minus	460
		Insecta	
Pisces		Chironomus riparius	41 ^h
Danio rerio	14 ^d	•	
Micropterus salmoides	11		
Oncorhynchus mykiss	23 ^d		
Oryzias latipes	93 ^e		

 $^{^{}a}$ Geometric mean of 10 and 24 μ g.L $^{-1}$ for the growth rate (most relevant parameter) under optimal growth conditions (2 d, pH restricted to 7.0-7.3).

Table 7. Phenanthrene: selected marine toxicity data for ERL derivation

Chronic Taxonomic group	NOEC/EC ₁₀ (μg.L ⁻¹)	Acute Taxonomic group	L(E)C ₅₀ (μg.L ⁻¹)
		Bacteria	
		Vibrio fischeri	310 ^d
Mollusca		Mollusca	
Mytilus galloprovincialis	29 ^a	Mytilus edulis	148
		Annelida	
		Neanthes arenaceodentata	187
Crustacea		Crustacea	
Acartia tonsa	69 ^b	Acartia tonsa	422
		Artemia salina	520
		Oithona davisae	522 ^e
		Palaemonetes pugio	360

b Most sensitive parameter (reproduction) determined under most reliable exposure regime (intermittent flow).

^c Most sensitive parameter (reproduction).

d Most sensitive parameter (weight).
e Most sensitive parameter (malformations).

^f Geometric mean of 180 and 302 μ g.L⁻¹ for the growth rate (most relevant parameter) under optimal growth conditions (2 d, pH restricted to 7.0-7.3).

⁹ Longest exposure time of 5 d.

h Most sensitive life-stage (1st instar) illuminated with a mercury light source 330-800 nm, including some UV-A.

Chronic	NOEC/EC ₁₀	Acute	L(E)C ₅₀
Taxonomic group	(µg.L ⁻¹)	Taxonomic group	(µg.L ⁻¹)
Echinodermata			
Arbacia punctulata	164		
Paracentrotus lividus	105 ^c		
Urochordata			
Ciona intestinalis	262 ^c		

a determined in the dark.

3.1.1 Mesocosm studies

No mesocosm studies are available.

3.2 Treatment of fresh- and saltwater toxicity data

As stated by Verbruggen (in prep.), there is no significant difference between the freshwater and marine acute toxicity data. Therefore the datasets can be combined. Chronic data for marine organisms appeared to be significantly higher than data for freshwater organisms. This was considered due to the inclusion of typically marine taxonomic groups (Echinodermata and Urochordata), which appeared to be relatively tolerant to phenanthrene. Therefore, the chronic data sets were combined as well.

3.3 Derivation of MPC_{fw} and MPC_{sw}

3.3.1 $MPC_{fw, eco}$ and $MPC_{sw, eco}$

The following derivation of the MPC $_{fw,\,eco}$ and MPC $_{sw,\,eco}$ is cited from Verbruggen (in prep.). Because acute and chronic toxicity data are available for algae, Daphnia, and fish, an assessment factor of 10 can be applied to the lowest NOEC or EC $_{10}$. This is the EC $_{10}$ of 11 $\mu g.L^{-1}$ for $Micropterus\, salmoides$. The resulting MPC $_{fw,\,eco}$ is 1.1 $\mu g.L^{-1}$. Because chronic data are available for additional taxonomic groups for the marine environment, the same assessment factor can be applied for the MPC $_{sw.\,eco}$, which is 1.1 $\mu g.L^{-1}$ too.

3.3.2 MPC_{fw, secpois} and MPC_{sw, secpois}

Phenanthrene has a BCF > 100 L.kg $^{-1}$, thus assessment of secondary poisoning is triggered. Therefore, toxicological data on birds and mammals should be used to derive an MPC $_{\text{oral, min}}$ from which the MPC $_{\text{fw, secpois}}$ and MPC $_{\text{sw, secpois}}$ can be derived. However no studies with population relevant endpoints for mammals and birds could be found. The EPA ECOTOX Database does contain NOELs for birds and mammals, but the underlying studies did not examine population relevant endpoints and/or only applied the PAH in a single dose and mostly only one concentration was tested.

As an alternative approach the $MPC_{fw,\;secpois}$ is derived in a similar way as the TDI for phenanthrene.

The TDI of phenanthrene was based on the toxicity of the aromatic C_{9-16} fraction of TPH (Total Petroleum Hydrocarbon) because phenanthrene is part of this fraction (Baars et al., 2001). On the same grounds, it can be presumed that the underlying studies can be used to derive an MPC_{oral, min} for phenanthrene. The

^b most sensitive parameter (recruitment) determined under most reliable exposure regime (intermittent flow).

 $^{^{}c}$ determined with a photoperiod 14:10 h light:dark by cool daylight lamps (380-780nm, PAR) with an intensity of 70 μE.m $^{-2}$.s $^{-1}$.

^d Geometric mean of 530, 530, 510, 520, 144, 142, and 182 μ g.L⁻¹ for standard exposure time (15 min).

^e Most sensitive endpoint (mortality).

TDI is based on studies with 90 days administration of naphthalene, fluorene, anthracene, fluoranthene and pyrene (TPHCWG, 1997). Considering population relevant endpoints, the lowest NOEL for rats from these studies was 50 mg.kg_{bw} $^{-1}$.d $^{-1}$ for changes in body weight from a study with naphthalene. With a conversion factor of 20 to calculate a NOEC_{oral} in mg.kg_{food} $^{-1}$ and an assessment factor of 90 for 90 days studies the MPC_{oral} is 11 mg.kg_{food} $^{-1}$. The lowest NOEL for mice was 250 mg.kg_{bw} $^{-1}$.d $^{-1}$ for changes in body weight and food consumption from a study with fluoranthene. With a conversion factor of 8.3 to calculate a NOEC_{oral} in mg.kg_{food} $^{-1}$ and an assessment factor of 90, the MPC_{oral} is 23 mg.kg_{food} $^{-1}$. The MPC_{oral}, min of 11 mg.kg_{food} $^{-1}$ is used to calculate the MPC_{fw}, secpois and MPC_{sw}, secpois. With the BCF of 1750 L.kg $^{-1}$ and BMF1 and BMF2 of 1, the MPC_{fw}, secpois and MPC_{sw}, secpois are both 6.3 μ g.L $^{-1}$.

3.3.3 MPC_{water, hh food}

Derivation of MPC_{water, hh food} for phenanthrene is triggered (Table 5). This derivation is based on the TDI of 0.040 mg.kg_{bw} $^{-1}$.day $^{-1}$. MPC_{hh, food} = 0.1 x TL_{hh} x BW / 0.115 = 2.4 mg.kg_{food} $^{-1}$, where the TL_{hh} is the TDI, BW is a body weight of 70 kg, 0.115 kg is the daily consumption of fishery products. With a BCF of 1664 L.kg $^{-1}$ and a BMF1 of 1, the resulting MPC_{water, hh food} is then: 2.4 / (1664*1) = 1.5 μ g.L $^{-1}$. The MPC_{water, hh food} is valid for the freshwater and saltwater compartment.

3.3.4 Selection of the MPC_{fw} and MPC_{sw}

The MPC_{fw} and the MPC_{sw} are determined by the lowest MPC_{fw/sw} derived. Therefore, both the MPC_{fw} and the MPC_{sw} are 1.1 μ g.L⁻¹.

Phenanthrene has a log $K_{p, susp-water} \ge 3$; expression of the MPC_{water} as MPC_{susp, water} is required. The MPC_{susp, water} is calculated according to:

 $MPC_{susp, water} = MPC_{water, dissolved} X K_{p, susp-water, Dutch standard}$

For this calculation, $K_{p,susp-water,Dutch\ standard}$ is calculated from the log Koc of 4.292 as given in Table 3. With an $f_{OC,susp,\ Dutch\ standard}$ of 0.1176 the $K_{p,\ susp-water,\ Dutch\ standard}$ can be calculated to 2305 L.kg⁻¹. With this value, both the MPC_{susp,\ fw} and the MPC_{susp,\ sw} are 2.5 mg.kg_{dwt}⁻¹.

3.4 Derivation of MPC_{dw, hh}

No A1 value and DW standard are available for phenanthrene. With the TDI of 0.040 mg.kg_{bw}⁻¹.day⁻¹ an MPC_{dw, hh, provisional} can be calculated with the following formula: MPC_{dw, hh, provisional} = 0.1 x TL_{hh} x BW / uptake_{dw} where the TL_{hh} is the TDI, BW is a body weight of 70 kg, and uptake_{dw} is a daily uptake of 2 L. As described in section 2.2 water treatment is currently not taken into account. Therefore the MPC_{dw, hh} = the MPC_{dw, hh, provisional} and becomes: 0.1 x 0.040 x 70 / 2 = 0.14 mg.L⁻¹.

3.5 Derivation of MAC_{eco}

The following derivation of the MAC_{eco} is cited from Verbruggen (in prep.). There are no reliable acute toxicity data for fish or other vertebrates and for aquatic plants. However, from two fish species tested and one aquatic plant (i.e. studies that were considered unreliable because only nominal concentrations were reported), these groups do not appear particularly sensitive. Therefore, the $MAC_{fw, eco}$ and the $MAC_{sw, eco}$ can be derived from a species sensitivity distribution (Figure 2). The HC5 of the acute toxicity data is 67 μ g.L⁻¹, which is above the

lowest value of 41 μ g.L⁻¹ for *Chironomus riparius*. The HC₅₀ is 307 μ g.L⁻¹. The goodness-of-fit is accepted at all significance levels. The number of toxicity data and the taxonomic diversity is high and the differences in species sensitivity are low, which is characteristic of narcotic effects. The MAC_{fw, eco} should be protective of any acute effects. However, the values used in the SSD are 50% effective concentration. Therefore, an assessment is made between the 50% and 10% effective concentrations (EC₅₀ and EC₁₀). A direct comparison can be made for eight species from four taxonomic groups (Table 8). The ratio between the EC₅₀ and EC₁₀ varies widely. Moreover, such data have not been generated for the most sensitive taxonomic group, which are the insects. Therefore, an assessment factor of 10 is applied to the HC5 (acute) to derive the MAC_{fw, eco}. The MAC_{fw, eco} is thus 6.7 μ g.L⁻¹. Because of the number of marine data, including non standard species such as annelids and molluscs, an extra assessment factor for the MAC_{sw, eco} is not necessary. The MAC_{sw, eco} is 6.7 μ g.L⁻¹ too.

Table 8 Ratio of acute no effect levels (10% cut-off by means of EC_{10}) versus

50% effect levels (EC₅₀) for phenanthrene

Taxon	Species	EC ₅₀ /EC ₁₀ or LC ₅₀ /LC ₁₀
Algae	Pseudokirchneriella subcapitata	6.5-18
Algae	Scenedesmus vacuolatus	3.9
Bacteria	Vibrio fischeri	3.7-24
Crustacea	Daphnia magna	1.3-2.6
Crustacea	Daphnia pulex	2.5
Crustacea	Acartia tonsa	1.3
Crustacea	Oithona davisae	2.1-2.7
Cyanophyta	Anabaena flos-aqua	2.5

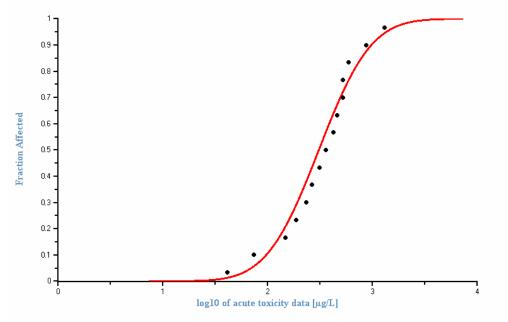


Figure 2 Species sensitivity distribution for the acute toxicity of phenanthrene to aquatic organisms

3.6 Derivation of the NC

Negligible concentrations are derived by dividing the MPCs by a factor of 100. This gives an NC_{fw} of 11 $ng.L^{-1}$ and an NC_{sw} of 11 $ng.L^{-1}$

3.7 Derivation of the SRC_{water, eco}

The following derivation of the $SRC_{water, eco}$ is cited from Verbruggen (in prep.). The $SRC_{water, eco}$ is equal to the geometric mean of the chronic toxicity data and is 43 μ g.L⁻¹. The $SRC_{water, eco}$ is valid for the salt- and freshwater environment.

3.8 Lipid approach

In Verbruggen (in prep.), ERLs were also calculated on the basis of internal lipid concentrations. In this approach all individual toxicity data for all examined PAHs were recalculated to internal lipid concentrations and concentrations were expressed on a molar basis. The obtained dataset was set out in a species sensitivity distribution and the values for HC_5 and HC_{50} have been recalculated to concentrations for the individual PAHs in water, sediment and soil. More details on this approach can be found in Verbruggen (in prep.). With this method an $MPC_{fw, eco}$ of $0.68~\mu g.L^{-1}$ was calculated after application of an assessment factor of 5 to the HC_5 . The HC_{50} of $38~\mu g.L^{-1}$ was taken over as the $SRC_{water, eco}$. These values are comparable to the derived ERL values for freshwater.

4 Toxicity data and derivation of ERLs for sediment

4.1 Toxicity data

An overview of the selected sediment toxicity data for phenanthrene as reported by Verbruggen (in prep.) is given in Table 9. These values are recalculated to standard sediment with 10% organic matter. The crustaceans *Rhepoxynius abronius* and *Schizopera knabeni* are marine species while the annelid *Limnodrilus hoffmeisteri* inhabits mostly brackish sediments. The rest of the species live in freshwater sediments.

Table 9. Phenanthrene: selected chronic sediment toxicity data for ERL derivation

Chronic ^a	NOEC/EC ₁₀	
Taxonomic group	(mg.kg _{dwt} -1)	
Annelida		
Limnodrilus hoffmeisteri	168 ^a	
Lumbriculus variegatus	26	
Crustacea		
Hyalella azteca	167 ^b	
Rhepoxynius abronius	122 ^c	
Schizopera knabeni	7.8 ^d	
Insecta		
Chironomus riparius	91 ^e	

^a most sensitive parameter (sediment egestion).

4.2 Derivation of MPC_{sediment}

The following derivation of the MPC $_{\rm sediment}$ is cited from Verbruggen (in prep.). With six chronic data from three taxonomic groups equally distributed over freshwater and marine species, a minimum assessment factor of 10 can be applied to derive the MPC $_{\rm sediment, \ fw}$ and MPC $_{\rm sediment, \ sw}$. The resulting value is 0.78 mg.kg $_{\rm dwt}$ $^{-1}$ for Dutch standard sediment.

Both the MPC $_{\rm sediment,\;fw}$ and the MPC $_{\rm sediment,\;sw}$ are 0.78 mg.kg $_{\rm dwt}^{-1}$ for Dutch standard sediment.

4.3 Derivation of NC_{sediment}

The NC_{sediment, fw} is set a factor of 100 lower than de MPC_{sediment, fw} at 7.8 μ g.kg _{dwt}⁻¹ for Dutch standard sediment. The NC_{sediment, sw} is 7.8 μ g.kg _{dwt}⁻¹ for Dutch standard sediment.

4.4 Derivation of SRC_{sediment, eco}

The following derivation of the $SRC_{sediment, eco}$ is cited from Verbruggen (in prep.). The $SRC_{sediment, eco}$ is derived from the geometric mean of the benthic data and is 63 mg.kg_{dwt}⁻¹ for Dutch standard sediment.

b geometric mean of 339, 113, and 122 mg.kg_{dw, standard sed}⁻¹, recalculated to standard sediment with 10% organic matter, for the most sensitive parameter (length).

c geometric mean of 125 and 120 mg.kg_{dw, standard sed}-1, recalculated to standard sediment with 10% organic matter.

^d most sensitive parameter (reproduction).

^e geometric mean of 84, 114, and 79 mg.kg_{dw, standard sed} $^{-1}$, recalculated to standard sediment with 10% organic matter for the parameter emergence/mortality in a 28-d study.

The $SRC_{sediment, eco}$ is 63 mg.kg $_{dwt}^{-1}$ for Dutch standard sediment. The $SRC_{sediment, eco}$ is valid for the marine and the freshwater environment.

4.5 Lipid approach

With the lipid approach as briefly described in Section 3.8, Verbruggen (in prep.) calculated an MPC $_{\rm sediment,\ fw}$ of 0.78 mg.kg $_{\rm dwt}^{-1}$, after application of an assessment factor of 5 to the HC5. An HC50 of 44 mg.kg $_{\rm dwt}^{-1}$ was taken over as the SRC $_{\rm sediment,\ eco}$. Both values were normalised for Dutch standard sediment. These values are comparable to the derived ERL values for sediment.

5 Toxicity data and derivation of ERLs for soil

5.1 Toxicity data

An overview of the selected soil toxicity data for phenanthrene as reported by Verbruggen (in prep.) is given in Table 10.

Table 10. Phenanthrene: selected chronic soil toxicity data for ERL derivation

Table 10. I Heriantin ene. Sei	ceted emorne son toxicity data
Chronic ^a	NOEC/EC ₁₀
Taxonomic group	(mg.kg _{standard soil} -1)
Bacteria	
nitrification	154
Macrophyta	
Sinapsis alba	98 ^a
Trifolium pretense	88 ^a
Lolium perenne	645 ^a
Annelida	
Eisenia fetida	36 ^b
Eisenia veneta	92
Enchytraeus crypticus	87 ^c
Insecta	
Folsomia candida	37 ^d
Folsomia fimetaria	72 ^c

^a Most sensitive endpoint (fresh weight).

5.2 Derivation of MPC_{soil}

5.2.1 MPC_{soil}, eco

The following derivation of the MPC $_{soil, eco}$ is cited from Verbruggen (in prep.). Chronic toxicity data for phenanthrene in soil are available for annelids, collembola, terrestrial plants, and microbial processes. The EC $_{10}$ for reproduction of *Eisenia fetida* is the lowest EC $_{10}$ or NOEC. This value is almost equal to the geometric mean of 33 and 41 mg.kg $_{dwt, standard soil}^{-1}$ for the springtail *Folsomia candida*. Because chronic data are available for 8 species and 1 terrestrial process, covering all trophic levels, an assessment factor of 10 can be applied to derive the MPC $_{soil, eco}$. The MPC $_{soil, eco}$ is thus 3.6 mg.kg $_{dwt}^{-1}$ for Dutch standard soil.

The MPC_{eco} for soil is 3.6 mg.kg_{dwt}⁻¹ for Dutch standard soil.

5.2.2 MPC_{soil, secpois}

Phenanthrene has a BCF $> 100 \text{ L.kg}^{-1}$ and therefore secondary poisoning is triggered.

An indicative MPC_{soil}, secpois can be calculated from the indicative MPC_{oral} of $11~\text{mg.kg}_{\text{food}}^{-1}$ as calculated in Section 3.3.2. The MPC_{soil}, secpois, TGD can be calculated with the method as described in Van Vlaardingen and Verbruggen (2007). This calculation has been performed with an estimated BCF value of $380~\text{L.kg}^{-1}$, based on the log K_{ow}. The calculated MPC_{soil}, secpois, TGD is: $11.4~\text{mg.kg}_{\text{dwt}}^{-1}$. Conversion to Dutch standard soil gives $33.7~\text{mg.kg}_{\text{dwt}}^{-1}$ for

b Most sensitive endpoint (total offspring) derived from presented data based on time weighted average concentrations.

^c Most sensitive endpoint (reproduction) corrected for time weighted average concentrations.

^d Geometric mean of 33 and 41 mg.kg_{dwt}-¹ for most sensitive endpoint (reproduction) corrected for time weighted average concentrations.

Dutch standard soil. Jager (Jager, 1998) reported BCF values for earthworms ranging from 9 to 40 L.kg⁻¹. This indicates that the calculated MPC_{soil, secpois} is most likely a worst case estimate.

5.2.3 MPC_{soil, hh food}

For the derivation of the MPC_{soil, hh food}, the TDI of 0.040 mg.kg_{bw}- 1 .day- 1 can be used as TL_{hh}. With the method as described in van Vlaardingen and Verbruggen (2007), specific human intake routes are allowed to contribute 10% of the human toxicological threshold limit. Four different routes contributing to human exposure have been incorporated: consumption of leafy crops, root crops, milk and meat. Uptake via root crops was determined to be the critical route. The calculated MPC_{soil, hh food} is 1.85 mg.kg_{dwt}- 1 for Dutch standard soil.

5.2.4 Selection of the MPC_{soil}

The lowest MPC_{soil} is the MPC_{soil}, $_{hh\ food}$, this sets the MPC_{soil} to 1.9 mg. kg_{dwt}^{-1} for Dutch standard soil.

5.3 Derivation of NC_{soil}

The NC_{soil} is set a factor of 100 lower than de MPC $_{soil}$ at 0.019 mg.kg $_{dwt}$ $^{-1}$ for Dutch standard soil.

5.4 Derivation of SRC_{soil, eco}

The following derivation of the $SRC_{soil, eco}$ is cited from Verbruggen (in prep.). The $SRC_{soil, eco}$ is derived from the geometric mean of the data for the eight species and is 90 mg. kg_{dwt}^{-1} for Dutch standard soil.

5.5 Lipid approach

With the lipid approach as briefly described in Section 3.8, Verbruggen (in prep.) calculated an MPC $_{soil,\ eco}$ of 0.76 mg.kg $_{dwt}^{-1}$ was calculated, after application of an assessment factor of 5 to the HC5. The HC50 of 44 mg.kg $_{dwt}^{-1}$ was taken over as the SRC $_{soil,\ eco}$. Both values are normalised for Dutch standard soil. These values are comparable to the derived ERL values for soil.

6 Derivation of ERLs for groundwater

6.1 Derivation of MPC_{gw}

6.1.1 $MPC_{gw, eco}$

Since groundwater-specific ecotoxicological ERLs are absent, the surface water MPC_{fw, eco} is taken as a substitute. Thus the MPC_{gw, eco} = MPC_{fw, eco} = 1.1 μ g.L⁻¹.

6.1.2 MPC_{gw, hh}

The $MPC_{gw, hh}$ is set equal to the $MPC_{dw, hh}$: 0.14 mg.L⁻¹.

6.1.3 Selection of the MPC_{gw}

The lowest MPC_{gw} sets the MPC_{gw} this is the MPC_{gw, eco}: $1.1 \mu g.L^{-1}$.

6.2 Derivation of NC_{gw}

The NC_{gw} is set a factor 10 lower than the MPC_{gw} : 11 ng.L⁻¹.

6.3 Derivation of SRC_{gw, eco}

The $SRC_{gw, eco}$ is set equal to the $SRC_{water, eco}$: 43 $\mu g.L^{-1}$.

7 Derivation of ERLs for air

7.1 Derivation of MPC_{air}

7.1.1 MPC_{air, eco}

No data are available to derive an $\mbox{MPC}_{\mbox{\scriptsize air, eco}}.$

7.1.2 MPC_{air, hh}

Since only an unbound TCA for phenanthrene in air is available, an $\mbox{MPC}_{\mbox{\scriptsize air, hh}}$ cannot be derived.

7.1.3 Selection of the MPC_{air}

No MPC_{air} can be derived.

7.2 Derivation of NC_{air}

Since there is no MPC $_{\mbox{\scriptsize air}}$ an NC $_{\mbox{\scriptsize air}}$ can also not be derived.

8 Comparison of derived ERLs with monitoring data

Surfacewater

The RIWA (Dutch Association of River Water companies) reports monitoring data for phenanthrene in the Rhine and Meuse basins. The total concentrations for the years 2006-2010 are given in Table 11. These values cannot be directly compared with the ERLs derived in this report that expressed as dissolved concentrations. However, presuming a concentration of suspended matter in surface water varying between 15 and 30 mg.L $^{-1}$ and the $K_{p,\;susp-water,\;Dutch\;standard}$ given in Section 3.3.4, the fraction of the total concentration sorbed to suspended matter is only 3 to 6%. The total concentration is therefore representative for the dissolved fraction. It can be concluded that none of these values exceeds the newly derived MPC or MAC for freshwater, but the NC $_{\rm water}$ of 11 ng.L $^{-1}$ is exceeded in many cases.

Table 11 Total concentrations ($\mu g.L^{-1}$) of phenanthrene in surface water of the Rhine and Meuse for the vears 2006-2010. Source: RIWA

location	2006		2007		2008		2009		2010	
	aa.c	max	aa.	max	aa.	max	aa.	max	aa.	max
Rhine										
Lobith	< d	<	0.0896	1	0.015	0.04	<	<	<	0.01
Nieuwegein ^a	0.0319	0.07	0.0147	0.02	0.0187	0.03	0.03	0.05	0.0107	0.03
Nieuwersluis ^b	0.0223	0.06	0.0137	0.04	<	0.02	0.0137	0.02	0.0233	0.08
Meuse										
Eijsden	_ e	-	0.02	0.05	0.0154	0.04	-	-	-	-
Heel	<	<	<	<	<	<	<	0.0062	0.0129	0.05
Brakel	<	0.02	<	003	<	0.02	<	0.02	<	0.02
Keizersveer	0.01	0.04	0.01	0.03	<	0.02	<	0.01	0.0235	0.11
Stellendam	<	<	<	<	<	<	<	<	<	<

^a Lek canal.

The Dutch Ministry of Infrastructure and Environment does present monitoring data for total concentrations of phenanthrene in water and sediment on their website (www.waterbase.nl). For the years 2001 to 2010 maximum peak values for surface water were reported up to 3.1 μ g.L⁻¹. These values do not exceed the MAC_{eco} derived in this report. For suspended matter, the average of the concentrations reported for 2001 to 2010 did not exceed the MPC_{susp, fw} or MPC_{susp, sw}.

For remote mountain lakes in the Pyrenees, alps and central Norway, dissolved water concentration for phenanthrene are reported ranging from 0.096 to 0.176 ng.L⁻¹ (Vilanova et al., 2001). In these water samples, phenanthrene counted for 31 to 45% of the total PAH concentration. For the marine environment, background concentrations have been agreed for several regions of the North-East Atlantic. The background concentration of phenanthrene ranges from 0.262 to 0.636 ng.L⁻¹ (OSPAR, 2005). These values are lower than the NCs for fresh- and saltwater.

^b Amsterdam-Rhine canal.

^c aa. = annual average.

d < = below limit of detection/quantification.

e - = not reported.

Sediment

For sediment, over the years 2001 to 2010 the reported concentrations exceeded the new derived MPC $_{\rm sediment}$ in four occasions: 3.39 mg.kg $_{\rm dwt}^{-1}$ (Sas van Gent, 2001), 1.2 mg.kg $_{\rm dwt}^{-1}$ (Sas van Gent 2006), 2.7 mg.kg $_{\rm dwt}^{-1}$ (Sas van Gent, 2009) and 0.79 mg.kg $_{\rm dwt}^{-1}$ (Bovensluis, 2002). Most of the other reported values exceed the newly derived NC $_{\rm sediment}$. Concentrations in North Sea sediment are also collected for the OSPAR convention. Actual concentrations are not report for phenanthrene but in the assessment report for 2008/2009 (OSPAR, 2009b) it can be seen that the concentration in almost all samples exceed the OSPAR "Background Assessment Concentration" of 32 µg.kg $_{\rm dwt}^{-1}$ normalised to 2.5% TOC (OSPAR, 2009a). Normalised to Dutch standard sediment, this value would be about a factor 10 lower than the MPC $_{\rm sediment}^{-1}$ derived in this report but exceeding the NC $_{\rm sediment}^{-1}$. The trends for concentrations of phenanthrene in North Sea sediment over the period 2003-2007 are in general stable and rarely declining.

Soil

In the year 2000, the AW2000 project examined the concentrations of many contaminants in agricultural soil and soils in nature reserves in the Netherlands, which were not exposed to local sources of contamination, in order to determine their background values in the Netherlands (Lamé et al., 2004b). The median concentration of phenanthrene in the upper soil (0-0.1 m) was determined at 10 μg.kg_{dwt}-1 for Dutch standard soil. In the lower soil (0.5-1.0 m) the median was determined at 0.004 µg.kg_{dwt}⁻¹ for Dutch standard soil. These values are comparable to the estimated natural background concentration of 1-10 µg.kg_{soil}-1 for individual PAHs as determined by Wilcke (2000). It seems in contradiction that soils in European high mountain areas, recently examined on their PAH concentration (Quiroz et al., 2011) showed higher concentrations. For phenanthrene, the average concentrations were 117 µg.kg⁻¹, 72 µg.kg⁻¹, 256 μg.kg⁻¹ and 120 μg.kg⁻¹ for Montseny (Spain), Pyrenees (French-Spanish border), Alps (Austria) and Tatras (Slovania) respectively. However, the actual concentration is correlated to the altitude and these high concentrations are attributed to condensation effects at higher altitudes caused by the lower temperatures. When this correlation is extrapolated to sea level, the estimated value is comparable to those determined within the AW2000 project (Lamé et al., 2004a) and by Wilcke (2000). The maximum concentrations monitored in the AW2000 project are 1.16 mg.kg_{dwt}⁻¹ and 0.093 mg.kg_{dwt}⁻¹ for the upper and lower soil respectively normalised to Dutch standard soil. These values do not exceed the derived MPC value for soil but are higher than the NC_{soil}. The 80% level in the upper soils was 0.022 mg.kg_{dwt}-1 meaning that at least 20% of the uppers soil samples exceed the NC_{soil} . In the lower soils, the 90% level of 0.018 mg.kg_{dwt}⁻¹ was close to the NC_{soil}. From this can be concluded that the newly derived NC_{soil} will be exceeded in many areas with a relatively low exposure of PAHs.

Sum of PAHs

The observations reported above are based on the reported concentrations for phenanthrene alone. It should be considered that phenanthrene will not occur on its own but as part of the mixture of PAHs. Therefore, the occurrence of mixture toxicity should be considered when performing a risk assessment. PAHs are a large group of substances of which the mechanisms of toxicity are comparable. Therefore, the risk assessment for every environmental compartment should be based on concentration addition for every PAH determined and not on a single PAH like phenanthrene alone.

9 Conclusions

In this report, the risk limits Negligible Concentration (NC), Maximum Permissible Concentration (MPC), Maximum Acceptable Concentration for ecosystems (MAC_{eco}), and Serious Risk Concentration for ecosystems (SRC_{eco}) are derived for phenanthrene in water, groundwater, sediment and soil. The MPC for water and suspended matter are higher than the current valid ERLs. The MPC for sediment is also higher than the current valid ERL. These differences are due to a more extensive dataset on ecotoxicology in combination with the more recent methodology for derivation of ERLs. Monitoring data suggests that currently the NC_{water} derived in this report will be exceeded in the Dutch surface waters. Also, the MPC_{sediment} could be exceeded in some cases and the NC_{sediment} is likely to be exceeded in many cases. Besides that, it should be mentioned that phenanthrene will not occur on its own but as part of the mixture of PAHs. For a substance group like PAHs, additive effects (mixture toxicity) should not be ruled out and the total group of PAHs should be assessed by application of concentration addition, at least for ecotoxic effects. The ERLs that were obtained are summarised in the table below. For the soil compartment, it can be concluded that the NC_{soil} will be exceeded in many cases, including soils with a relatively low exposure to PAHs.

Table 12. Derived MPC, NC, MAC_{eco}, and SRC_{eco} values for phenanthrene

ERL	unit	value	•		
		MPC	NC	MACeco	SRC _{eco}
freshwater ^a	μg.L ⁻¹	1.1	1.1 x10 ⁻²	6.7	43
freshwater susp. matter ^b	mg.kg _{dwt} ⁻¹	2.5			
drinking water human health ^c	mg.L ⁻¹	0.14			
saltwater	μg.L ⁻¹	1.1	1.1 x10 ⁻²	6.7	43
saltwater susp. matter	mg.kg _{dwt} -1	2.5			
freshwater sediment ^d	mg.kg _{dwt} -1	0.78	7.8 x10 ⁻³		63
saltwater sediment ^d	mg.kg _{dwt} -1	0.78	7.8 x10 ⁻³		63
soil ^e	mg.kg _{dwt} -1	1.9	1.9 x10 ⁻²		90
groundwater	μg.L ⁻¹	1.1	1.1 x10 ⁻²		43
air	mg.m ⁻³	n.d.			

 $[^]a$ From the MPC $_{\rm fw,\,eco}$, MPC $_{\rm fw,\,secpois}$ and MPC $_{\rm fw,\,hh\,food}$ the lowest one is selected as the 'overall' MPC $_{\rm water}$.

^b Expressed on the basis of Dutch standard suspended matter.

 $[^]c$ As stated in the new WFD guidance, the MPC $_{dw,\,hh}$ is not included in the selection of the final MPC $_{fw}$. Therefore, the MPC $_{dw,\,hh}$ is presented as a separate value.

 $^{^{\}mbox{\scriptsize d}}$ Expressed on the basis of Dutch standard sediment.

e Expressed on the basis of Dutch standard soil.

n.d. = not derived.

References

- Anonymous. 1998. Basisrapportage derde voedselconsumptiepeiling. Zeist: TNO. Baars A, Theelen R, Janssen P, Hesse J, van Apeldoorn M, Meijerink M, Verdam L, Zeilmaker M. 2001. Re-evaluation of human-toxicological maximum permissible risk levels. Bilthoven, The Netherlands: National Institute of Public Health and the Environment (RIVM). Report no. 711701025.
- Baussant T, Sanni S, Jonsson G, Skadsheim A, Børseth JF. 2001a.

 Bioaccumulation of polycyclic aromatic compounds: 1. Bioconcentration in two marine species and in semipermeable membrane devices during chronic exposure to dispersed crude oil. Environ Toxicol Chem. 20: 1175-1184.
- Baussant T, Sanni S, Skadsheim A, Jonsson G, Børseth JF, Gaudebert B. 2001b. Bioaccumulation of polycyclic aromatic compounds: 2. Modeling bioaccumulation in marine organisms chronically exposed to dispersed oil. Environ Toxicol Chem. 20: 1185-1195.
- Bihari N, Fafandel M, Piskur V. 2007. Polycyclic aromatic hydrocarbons and ecotoxicological characterization of seawater, sediment, and mussel *Mytilus galloprovincialis* from the gulf of Rijeka, the Adriatic Sea, Croatia. Arch. Environ. Contam. Toxicol. 52: 379-387.
- Bleeker EAJ, Verbruggen EMJ. 2009. Bioaccumulation of polycyclic aromatic hydrocarbons in aquatic organisms. Bilthoven: RIVM. Report no. 601779002.
- Burkhard LP, Lukasewycz MT. 2000. Some bioaccumulation factors and biotasediment accumulation factors for polycyclic aromatic hydrocarbons in lake trout. Environmental Toxicology and Chemistry. 19: 1427-1429.
- Cailleaud K, Budzinski H, Le Menach K, Souissi S, Forget-Leray J. 2009. Uptake and elimination of hydrophobic organic contaminants in estuarine copepods: An experimental study. Environ.Toxicol.Chem. 28: 239-246.
- Carlson RM, Oyler AR, Gerhart EH, Caple R, Welch KJ, Kopperman HL, Bodenner D, Swanson D. 1979. Implications to the aquatic environment of polynuclear aromatic hydrocarbons liberated from Northern Great Plains coal. Duluth, MN: U.S. EPA Report no. U.S. EPA report EPA-600/3-79-093.
- Casserly DM, Davis EM, Downs TD, Guthrie RK. 1983. Sorption of organics by Selenastrum capricornutum. Water Research. 17: 1591-1594.
- Cheikyula JO, Koyama J, Uno S. 2008. Comparative study of bioconcentration and EROD activity induction in the Japanese flounder, red sea bream, and Java medaka exposed to polycyclic aromatic hydrocarbons. Environ. Toxicol.: 354-362.
- De Maagd PG-J. 1996. Polycyclic aromatic hydrocarbons: fate and effects in aquatic environment. Utrecht, Utrecht University.
- Djomo JE, Garrigues P, Narbonne JF. 1996. Uptake and depuration of polycyclic aromatic hydrocarbons from sediment by the zebrafish (*Brachydanio rerio*). Environ Toxicol Chem. 15: 1177-1181.
- Duxbury CL, Dixon DG, Greenberg BM. 1997. Effects of simulated solar radiation on the bioaccumulation of polycyclic aromatic hydrocarbons by the duckweed *Lemna gibba*. Environ.Toxicol.Chem. 16: 1739-1748.
- Eastmond DA, Booth GM, Lee ML. 1984. Toxicity, accumulation, and elimination of polycyclic aromatic sulfur heterocycles in *Daphnia magna*. Arch. Environ. Contam. Toxicol. 13: 105-111.
- EC. 2001. Ambient air pollution by polycyclic aromatic hydrocarbons (PAH).

 Position paper prepared by the working group on polycyclic aromatic

- hydrocarbons. Luxembourg: Office for official publications of the european communities. Report no. KH-41-01-373-EN-N.
- EC. 2003. Technical Guidance Document on risk assessment in support of Commission Directive 93/67/EEC on risk assessment for new notified substances, Commission Regulation (EC) No 1488/94 on risk assessment for existing substances and Directive 98/8/EC of the European Parliament and of the Council concerning the placing of biocidal products on the market. Ispra, Italy: European Commission Joint Research Centre.
- EC. 2011. Common implementation strategy for the Water Framework Directive (2000/60/EC). Guidance document No. 27. Technical guidance for deriving environmental quality standards. Brussels: European Commission.
- EU. 2004. Directive 2004/107/EC of the European Parliament and of the Council of 15 December 2004 relating to arsenic, cadmium, mercury, nickel and polycyclic aromatic hydrocarbons in ambient air. Official Journal of the European Union. European Union.
- Fan CW, Reinfelder JR. 2003. Phenanthrene accumulation kinetics in marine diatoms. Environmental Science & Technology. 37: 3405-3412.
- Frank AP, Landrum PF, Eadie BJ. 1986. Polycyclic aromatic hydrocarbon rates of uptake, depuration, and biotransformation by Lake Michigan *Stylodrilus heringianus*. Chemosphere. 15: 317-330.
- Freitag D, Ballhorn L, Geyer H, Korte F. 1985. Environmental hazard profile of organic chemicals. An experimental method for the assessment of the behaviour of organic chemicals in the ecosphere by means of simple laboratory tests with ¹⁴C labelled chemicals. Chemosphere. 14: 1589-1616.
- Geyer H, Politzki G, Freitag D. 1984. Prediction of ecotoxicological behaviour of chemicals: Relationship between n-octanol/water partition coefficient and bioaccumulation of organic chemicals by alga Chlorella. Chemosphere. 13: 269-284.
- Halling-Sørensen B, Nyholm N, Kusk KO, Jacobsson E. 2000. Influence of nitrogen status on the bioconcentration of hydrophobic organic compounds to *Selenastrum capricornutum*. Ecotoxicology and Environmental Safety. 45: 33-42.
- Hansler RJ, Van Herwijnen R, Posthumus R. 2008. Indicatieve milieukwaliteitsnormen voor prioritaire stoffen. Bilthoven: RIVM. Report no. 601782012.
- Jager T. 1998. Mechanistic approach for estimating bioconcentration of organic chemicals in earthworms (Oliochaeta). Environmental Toxicology and Chemistry. 17: 2080-2090.
- Jonker MTO, Van der Heijden SA. 2007. Bioconcentration factor hydrophobicity cutoff: An artificial phenomenon reconstructed. Environmental Science & Technology. 41: 7363-7369.
- Jonsson G, Bechmann RK, Bamber SD, Baussant T. 2004. Bioconcentration, biotransformation, and elimination of polycyclic aromatic hydrocarbons in sheepshead minnows (*Cyprinodon variegatus*) exposed to contaminated seawater. Environ.Toxicol.Chem. 23:: 1538-1548.
- Kalf DF, Crommentuijn GH, Posthumus R, Van de Plassche EJ. 1995. Integrated environmental quality objectives for polycyclic aromatic hydrocarbons (PAHs). Bilthoven: RIVM. Report no. 679101018.
- Klimisch HJ, Andreae M, Tillman U. 1997. A systematic approach for evaluating the quality of experimetnal toxicological and ecotoxicological data. Regulatory Toxicology and Pharmacology. 25: 1-5.

- Lamé FPJ, Brus DJ, Nieuwenhuis RH. 2004a. Achtergrondwaarden 2000 Bijlage Rapport 1 AW2000: Datasheets voor de geanalyseerde stoffen. Utrecht: Nederlands Instituut voor Toegepaste Geowatenschappen. Report no. NITG 04-2420A.
- Lamé FPJ, Brus DJ, Nieuwenhuis RH. 2004b. Achtergrondwaarden 2000 -Hoofdrapport AW2000 fase 1. Utrecht: Nederlands Instituut voor Toegepaste Geowatenschappen. Report no. NITG 04-2420-A.
- Landrum PF. 1988. Toxicinetics of organic xenobiotics in the amphipod, *Pontoporeia hoyi*: Role of physiological and environmental variables. Aquat. Toxicol. 12: 245-271.
- Landrum PF, Lotufo GR, Gossiaux DC, Gedeon ML, Lee JH. 2003.

 Bioaccumulation and critical body residue of PAHs in the amphipod,

 Diporeia spp.: Additional evidence to support toxicity additivity for PAH mixtures. Chemosphere 51: 481-489.
- Landrum PF, Poore R. 1988. Toxicokinetics of selected xenobiotics in *Hexagenia limbata*. J. Great Lakes Res. 14: 427-437.
- Lee JH, Landrum PF, Koh CH. 2002. Toxicokinetics and time-dependent PAH toxicity in the amphipod *Hyalella azteca*. Environmental Science & Technology. 36: 3124-3130.
- Lepper P. 2005. Manual on the Methodological Framework to Derive Environmental Quality Standards for Priority Substances in accordance with Article 16 of the Water Framework Directive (2000/60/EC). Schmallenberg, Germany: Fraunhofer-Institute Molecular Biology and Applied Ecology.
- Lijzen JPA, Baars AJ, Otte PF, Rikken MGJ, Swartjes FA, Verbruggen EMJ, van Wezel AP. 2001. Technical evaluation of the intervention values for soil/sediment and groundwater Human and ecotoxicological risk assessment and derivation of risk limits for soil, aquatic sediment and groundwater. Bilthoven, the Netherlands: National Institute of Public Health and the Environment (RIVM). Report no. 711701023.
- Lu PY, Metcalf RL, Plummer N, Mandel D. 1977. The environmental fate of three carcinogens: Benzo-(a)-pyrene, benzidine, and vinyl chloride evaluated in laboratory model ecosystems. Arch. Environ. Contam. Toxicol. 6: 129-142
- McLeese DW, Burridge LE. 1987. Comparative accumulation of PAHs in four marine invertebrates. In: Capuzzo JM, Kester DR (Eds.) Oceanic Processes in Marine Pollution. pp. 110-117. Malabar, FL, Robert E. Krieger.
- Neff JM, Anderson JW, Cox BA, Laughlin JRB, Rossi SS, Tatem HE. 1976. Effects of petroleum on survival, respiration and growth of marine animals. Sources, Effects and Sinks of Hydrocarbons in the Aquatic Environment. pp. 515-539. Arlington, USA, American Institute of Biological Sciences.
- Newsted JL, Giesy JP. 1987. Predictive models for photoinduced acute toxicity of polycyclic aromatic hydrocarbons to *Daphnia magna*, Strauss (Cladocera, crustacea). Environ.Toxicol.Chem. 6: 445-461.
- Nfon E, Cousins IT, Broman D. 2008. Biomagnification of organic pollutants in benthic and pelagic marine food chains from the Baltic Sea. Science of the Total Environment. 397: 190-204.
- Niimi AJ, Palazzo V. 1986. Biological half-lives of eight polycyclic aromatic hydrocarbons (PAHs) in rainbow trout (*Salmo gairdneri*). Water Research. 20: 503-507.
- Okay OS, Karacik B. 2008. Bioconcentration and phototoxicity of selected PAHs to marine mussel *Mytilus galloprovincialis*. Journal of Environmental Science and Health Part A. 43: 1234-1242.

- OSPAR. 2005. Agreement on background concentrations for contaminants in seawater, biota and sediment. London: OSPAR commission.
- OSPAR. 2009a. Background document on CEMP assessment criteria for QSR 2010. London, UK: OSPAR commission.
- OSPAR. 2009b. CEMP assessment report: 2008/2009 Assessment of trends and concentrations of selected hazardous substances in sediments and biota. London, UK: OSPAR commission.
- Palmork KH, Solbakken JE. 19981. Distribution and elimination of [9-14C]phenanthrene in the horse mussel (*Modiola modiolus*). Bull. Environ. Contam. Toxicol. 26: 196-201. Bull. Environ. Contam. Toxicol. 26: 196-201.
- Petersen GI, Kristensen P. 1998. Bioaccumulation of lipophilic substances in fish early life stages. Environ Toxicol Chem. 1385-1395.
- Quiroz R, Grimalt JO, Fernandez P, Camarero L, Catalan J, Stuchlik E, Thies H, Nickus U. 2011. Polycyclic aormatic hydrocarbons in soils from European high mountain areas. Water Air and Soil Pollution. 215: 655-666.
- Southworth GR, Beauchamp JJ, Schmieder PK. 1978. Bioaccumulation potential of polycyclic aromatic hydrocarbons in *Daphnia pulex*. Wat.Res. 12: 973-977.
- Takeuchi I, Miyoshi N, Mizukawa K, Takada H, Ikemoto T, Omori K, Tsuchiya K. 2009. Biomagnification profiles of polycyclic aromatic hydrocarbons, alkylphenols and polychlorinated biphenyls in Tokyo Bay elucidated by ¹³C adn ¹⁵N isotope ratios as guides to trophic web structure. Marine Pollution Bulletin. 58: 663-671.
- Telli-Karakoc F, Tolun L, Henkelmann B, Klimm C, Okaya O, Schramm K-W. 2002. Polycyclic aromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs) distributions in the Bay of Marmara sea: Izmit Bay. Environmental Pollution. 119: 383-397.
- TPHCWG. 1997. Development of fraction specific reference doses (RfDs) and reference concentrations (RfCs) for total petroleum hydrocarbons (TPH). Amherst: Amherst Scientific Publishers.
- Trucco RG, Engelhardt FR, Stacey B. 1983. Toxicity, accumulation and clearance of aromatic hydrocarbons in *Daphnia pulex*. Environmental Pollution (Series A). 31: 191-202.
- Van Hattum B, Cid Montañes JF. 1999. Toxicokinetics and bioconcentration of polycyclic aromatic hydrocarbons in freshwater isopods. Environmental Science & Technology. 33: 2409-2417.
- Van Vlaardingen PLA, Verbruggen EMJ. 2007. Guidance for the derivation of environmental risk limits within the framework of "International and national environmental quality standards for substances in the Netherlands" (INS). Bilthoven, The Netherlands: National Institute of Public Health and the Environment (RIVM). Report no. 601782001.
- Verbruggen EMJ. in prep. Environmental risk limits for polycyclic aromatic hydrocarbons (PAHs). Bilthoven, The Netherlands: National Institute of Public Health and the Environment (RIVM). Report no. 711701XXX.
- Vilanova RM, Fernandez P, Martinez C, Grimalt JO. 2001. Polycyclic aromatic hydrocarbons in remote mountain lake waters. Water Research. 35: 3916-3926.
- Vives I, Grimalt JO, Ventura M, Catalan J. 2005. Distribution of polycyclic aromatic hydrocarbons in the food web of a high mountain lake, Pyrenees, Catalonia, Spain. Environ.Toxicol.Chem. 24: 1344-1352.
- VROM. 1999. Environmental risk limits in the Netherlands. A review of environmental quality standards and their policy framework in the Netherlands. The Hague, The Netherlands: Ministry of Housing, Spatial Planning and the Environment.

- VROM. 2004. (Inter)nationale Normen Stoffen. Den Haag, the Netherlands: Ministerie van Volkshuisvesting, Ruimtelijke Ordening en Milieubeheer.
- Wan Y, Jin X, Hu J, Jin F. 2007. Trophic dilution of polycyclic aromatic hydrocarbons (PAHs) in a marine food web from Bohai Bay, North China. Environ.Sci.Technol. 41: 3109-3114.
- Wilcke W. 2000. Polycycilc aromatic hydrocarbons (PAHs) in soil a Review. J. Plant Nutr. Soil Sci. 163: 229-248.
- Witt G. 2002. Occurrence and transport of polycyclic aromatic hydrocarbons in the water bodies of the Baltic sea. Marine Chemistry. 79: 49-66.

Appendix 1 Detailed BCF data

Table A1.1. Bioconcentration factors for phenanthrene adopted from RIVM report 601779002 (Bleeker and Verbruggen, 2009)

Species	Species properties	Purity	Analysis	Test Test p type water	H Hardnes Salinity	s/ Temp	. Exposure time	Exp. concn.	lipid content	Uptake rate constant	Depuration rate constant	BCF	BCF type	Norm. BCF	Method	Ri Notes	Ref
		[%]			[g.L ⁻¹]	[°C]	[d]	[µg.L ⁻¹]	[%]	[h ⁻¹]	constant	[L.kg _{ww} -1]		[L.kg _{ww} -1]			
Algae					-					<u> </u>							
Chlorella fusca				S								1760	wet weight		equi.	3 22	Geyer et al. (1984)
Pseudokirchneriella				S								10620	wet weight		equi.	3 22	Casserly et al. (1983)
subcapitata																	
Pseudokirchneriella			C14	S								4.22 - 4.47	dry weight		equi.	3 22,25	Halling-Sørensen et al. (2000
subcapitata																	
Thalassiosira pseudonana			C14	S								17	dry weight		equi.		Fan and Reinfelder (2003)
Thalassiosira pseudonana			C14	S								38.3	dry weight		equi.	3 22,25	Fan and Reinfelder (2003)
Macrophyta																	
Lemna gibba		>95	C14	S			3+1	25	-	18.4	0.268	69	whole plant	-	kinetic	2 1	Duxbury et al. (1997)
Lemna gibba		>95	C14	S			3+1	25	-	8.66	0.35	25		-	kinetic		Duxbury et al. (1997)
Lemna gibba		>95	C14	S			3+1	25	-	6.55	0.453	14	whole plant	-	kinetic	2 1	Duxbury et al. (1997)
Lemna gibba		>95	C14	S			3+1	35	-	19.6	0.212	92	whole plant	-	kinetic	2 1	Duxbury et al. (1997)
Lemna gibba		>95	C14	S			3+1	35	-	9.55	0.314	30	whole plant	-	kinetic	2 1	Duxbury et al. (1997)
Lemna gibba		>95	C14	S			3+1	35	-	6.09	0.546	11	whole plant	-	kinetic	2 1	Duxbury et al. (1997)
Mollusca																	
Modiola modiolus				S								3.1	wet weight		equi.	3 20	Palmork and Solbakken (19981)
Mya arenaria			HPLC	F		10	4+14	4.3 ± 0.4	-	0.0064	8.2	1280	whole animal	-	kinetic	1 1	McLeese and Burridge (1987)
Mytilus edulis			HPLC	F		10	4+14	4.3±0.4	-	0.013	16.1	1240	whole animal	-	kinetic	1 1	McLeese and Burridge (1987)
Mytilus edulis				F								2932	wet weight		equi.	4 26	Baussant et al. (2001a)
Mytilus galloprovincialis	field collected	HPLCgrade	flu	R nw	22‰	22	7d	100	-			100	whole animal	-	equi.	3 13	Okay and Karacik (2008)
Mytilus galloprovincialis	field collected	HPLCgrade	flu	R nw	22‰	22	7d	400	-			279	whole animal	-	equi.	3 13	Okay and Karacik (2008)
Rangia cuneata												240	wet weight		kinetic	3 21	Neff et al. (1976)
Annelida																	
Capitella capitata				F								30.7	wet weight		egui.	4 28	Lu et al. (1977)
Lumriculus variegatus				S								34700	lipid weight		equi.	3 23	Jonker and Van der Heijden
5															•		(2007)
Nereis virens			HPLC	F		10	4+14	4.3±0.4	-	3	0.006	500	whole animal	-	kinetic	2 1	McLeese and Burridge (1987)
Stylodrilus heringianus		>98	C14	F		4	0.25 + 8	-	-	94.0±12.9	0.018 ± 0.003	5222	whole animal	-	kinetic	2 1	Frank et al. (1986)

Species	Species properties	Purity	Analysis	Test Test type water	Hardness/ Salinity	Temp.	Exposure time	Exp. concn.	lipid content	Uptake rate constant	Depuration rate constant	BCF	BCF type	Norm. BCF	Method Ri Notes	Ref
		[%]			[g.L ⁻¹]	[°C]	[d]	[µg.L ⁻¹]	[%]	[h ⁻¹]	constant	[L.kg _{ww} -1]		[L.kg _{ww} -1	1	
Crustacea Assellus aquaticus				S								1300	wet weight		equi. 3 22	Van Hattum and Cid Montane
Crangon septemspinosa Daphnia magna Daphnia magna Daphnia pulex	< 24h		HPLC HPLC C14 C14	F R S		10 23±1	4+14 1	4.3±0.4 40.1	-	6.8	0.032	210 324 600 1165-1424	whole animal whole animal wet weight wet weight		kinetic 1 1 equi. 2 1 equi. 4 27 equi. 4 27,29	McLeese and Burridge (1987) Newsted and Giesy (1987) Eastmond et al. (1984) Trucco et al. (1983)
Daphnia pulex			flu.	S		25	1	30	-			325	whole animal	-	equi. 2 1	Southworth et al.(1978)
Diporeira sp.	juvenile 5-11 months	>98	C14	R		4	28+nr	57.1	-	62.5±12.8	0.006±0.002	10417	whole animal	-	kinetic 2 1	Landrum et al. (2003)
Diporeira sp.	juvenile 5-11 months	>98	C14	R		4	28+nr	104.6	-	79.4±23.2	0.009 ± 0.003	8822	whole animal	-	kinetic 2 1	Landrum et al. (2003)
Diporeira sp.	juvenile 5-11 months	>98	C14	R		4	28+nr	214.4	-		0.005 ± 0.001	11440	whole animal	-	kinetic 2 1	Landrum et al. (2003)
Diporeira sp.	juvenile 5-11 months	>98	C14	R		4	28+nr	383	-		0.005±0.0042	10380	whole animal		kinetic 2 1	Landrum et al. (2003)
Diporeira sp.	juvenile 5-11 months	>98	C14	R		4	28+nr	637.8	-	44.1±9.4	0.008±0.003	5513	whole animal		kinetic 2 1	Landrum et al. (2003)
Eurytemora affinis Hyalella azteca			GC/MS C14	CF nw/dw R	15	10	3.6	0.06	-			530 440-504	whole animal wet weight		equi. 4 27,29	Cailleaud et al. (2009) Lee et al. (2002)
Pontoporeia hoyi		>98	C14	F		4	0.25+14	0.07-7.1	9.4	129.0±31	0.0046±0.0027	28043	whole animal	14893	kinetic 1 2	Landrum (1988)
Insecta																
Hexagenia limbata		>98	C14	F		10	0.25+14	0.07-7.1	7.8±1.9	131.1±46.8	0.032±0.004	4097	whole animal	2626	kinetic 2 1	Landrum and Poore (1988)
Hexagenia limbata		>98	C14	F		15		0.07-7.1		43.3±12.0	0.0076±0.0016		whole animal		kinetic 2 1	Landrum and Poore (1988)
Hexagenia limbata		>98	C14	F		15		0.07-7.1	9.1±3.4		0.029±0.002	1983	whole animal		kinetic 2 1	Landrum and Poore (1988)
Hexagenia limbata		>98	C14	F		20		0.07-7.1	6±2.4	56.3±6.8	0.032±0.004	1759	whole animal		kinetic 2 1	Landrum and Poore (1988)
Hexagenia limbata		>98	C14	F		20		0.07-7.1	3.7±1.2	33.0±8.0	0.067±0.008	493	whole animal		kinetic 2 1	Landrum and Poore (1988)
Hexagenia limbata		>98	C14	F		10	0.25+14	0.07-7.1	6±1.4	34.2±7.2	0.026±0.002	1315	whole animal	1096	kinetic 2 1	Landrum and Poore (1988)
Pisces																
Brachidanio rerio Brachidanio rerio				S R								11446 7943-9120	wet weight dry weight		kinetic 3 14 equi. 3 15	Djomo et al. (1996) Petersen and Kristensen (1998)
Culpea harengus				R								20893	dry weight		equi. 3 15	Petersen and Kristensen (1998)
Cyprinodon variegatus	2.5±1.2 g, 4.7±0.8 cm	n	GC-MS	CF sw	34	25	36+8	0.12±0.030		680	0.84	810	whole fish ww	418	kinetic 2 3	Jonsson et al. (2004)
Cyprinodon variegatus	2.5±1.2 g, 4.7±0.8 cm		GC-MS	CF sw	34	25	36+8	0.12±0.030	9.7			8351	whole fish lw	418	kinetic 2 3	Jonsson et al. (2004)
Cyprinodon variegatus	2.5±1.2 g, 4.7±0.8 cm	n	GC-MS	CF sw	34	25	36+8	1.12±0.32		1783	0.8	2229	whole fish ww	1149	kinetic 2	Jonsson et al. (2004)
Cyprinodon variegatus	2.5±1.2 g, 4.7±0.8 cm	n	GC-MS	CF sw	34	25	36+8	1.12±0.32	9.7			22977	whole fish lw	1149	kinetic 2	Jonsson et al. (2004)
Cyprinodon variegatus	2.5±1.2 g, 4.7±0.8 cm		GC-MS	CF sw	34	25	36+8	0.12±0.030				700	whole fish ww	361	egui. 2 3,9	Jonsson et al. (2004)
Cyprinodon variegatus	2.5±1.2 g, 4.7±0.8 cm	n	GC-MS	CF sw	34	25	36+8	1.12±0.32				1623	whole fish ww	837	equi. 2 9	Jonsson et al. (2004)
Gadus morhua				R								10715	dry weight		equi. 3 15	Petersen and Kristensen (1998)
Leuciscus idus melanotus Oncorhynchus mykiss Oryzias javanicus Pagrus major Pagrus major				S D F F								1760 613 150 180 173, 737	wet weight wet weight wet weight wet weight values for different tissues		equi. 3 16 kin 4 25 equi. 3 17 equi. 3 17 equi. 3 17,18	Freitag et al. (1985) Niimi and Palazzo (1986) Cheikyula et al. (2008) Cheikyula et al. (2008) Cheikyula et al. (2008)

Species	Species properties	Purity	Analysis	Test Test pH type water	Hardness Salinity	/ Temp.	. Exposure time	Exp. concn.	lipid content	Uptake rate constant	Depuration rate constant	BCF	BCF type	Norm. BCF	Method	Ri Note	es Ref
Parlychthys olivaceus Parlychthys olivaceus		[%]		F F	[g.L ⁻¹]	[°C]	[d]	[µg.L ⁻¹]	[%]	[h ⁻¹]		[L.kg _{ww} ⁻¹] 75 10, 181	wet weight values for different tissues	[L.kg _{ww} ⁻¹]	equi.	3 17 3 17,1	Cheikyula et al. (2008) 8 Cheikyula et al. (2008)
Pimephales promelas	5-6 w, 4.8% lipid		GC-PID	(C)F nw		24±1	28+5	2.63±0.83	4.8			1733	whole fish wy	v 1805	kinetic	2 4,5, 0	1 Carlson et al. (1979)
Pimephales promelas Pimephales promelas	5-6 w, 3.8% lipid 5-6 w, 4.1% lipid		GC-PID GC-PID	(C)F nw (C)F nw		24±1 24±1	28+5 28+5	2.55±0.44 2.53±0.44	3.8 4.1			3611 2086	whole fish ww whole fish ww		kinetic kinetic		Carlson et al. (1979) 1 Carlson et al. (1979)
Pimephales promelas Pimephales promelas	5-6 w, 4.3% lipid 5-6 w, 4.4% lipid		GC-PID GC-PID	(C)F nw (C)F nw		24±1 24±1	28+5 28+5	2.34±0.54 2.20±0.25	4.3 4.4			2084 2240	whole fish ww whole fish ww			2 4,10 2 4,7, 0	Carlson et al. (1979) 1 Carlson et al. (1979)
Pimephales promelas Scophthalmus maximus Scophthalmus maximus Scophthalmus maximus	0.52±0.21 g	>96	HPLC-Flu	S tw F F R		20±1	4	626		2000	0.3	6761 10300 936 11220	whole fish ww wet weight wet weight dry weight	V	kinetic	3 8,11 3 19 3 19 3 15	De Maagd (1996) Baussant et al. (2001a) Baussant et al. (2001b) Petersen and Kristensen (1998)

Notes

- 1 BCF is based on the parent compound.
- 2 In this study lipid content was expressed only as percentage of dry weight (35%). In addition the ratio between total wet weight and dry weight was given (0.269). For lipid normalization it was assumed that the same ratio holds for lipids, resulting in a lipid content of 9.4% based on wet weight; BCF is based on the parent compound. This species is the same as *Diporeia* sp.
- 3 Curve fitting not significant.
- 4 16:8 photoperiod; extracted with C18 solid phase; MeOH 10 μl.L⁻¹.
- 5 Tested together with α-naphthaflavone.
- 6 Tested together with 9-chlorophenanthrene.
- 7 Tested together with dibenzofuran, fluorene, 1methyl-phenanthrene, fluoranthene, and pyrene.
- 8 12:12 photoperiod; corrected for control volatilisation; recovery from fish fitted to data.
- 9 BCF assessed at days 4, 7 and 36.
- 10 Simultaneous kinetic method used.
- 11 Kinetic adjusted Banerjee method.
- 12 BCF based on dry weight.

- 13 BCF based on nominal concentration, highest concentration and possibly both concentrations are toxic.
- 14 Exposure via sediment for less than 96 h.
- 15 BCF values based on dry weight; early life stages used.
- 16 No air or food was provided; exposure 72 h.
- 17 Exposure to oversaturated PAH mixture.
- 18 BCF values in different tissues.
- 19 Exposure to oil, PAH concentration appears to be above water solubility.
- 20 Low on experimental detail, exposure concentration unclear.
- 21 Low on experimental detail, exposure type not reported, steady state not reached.
- 22 Short, static exposure; steady state unlikely.
- 23 Static exposure; sediment present; steady state unlikely.
- 24 BCF value based on dry weight and total radioactivity.
- 25 Exposed via diet.
- 26 Exposed to oil.
- 27 Based on total radioactivity.
- 28 Exposed in the field.
- 29 Values represent (a range of) BCF values from (a range of) different exposure concentrations.

Table A1.2. Bioaccumulation factors for phenanthrene

Species	Species properties	Analysis	Test water	рН	Hardness/ Salinity		Exp. conc.	lipid content	BAF	BAF type	Norm. BAF	Ri Notes	Ref
					[g.L ⁻¹]	[°C]	[ng.L ⁻¹]	[%]	[L.kg _{ww} ⁻¹]		[L.kg _{ww} ⁻¹]		
Mollusca													
Crassostrea gigas	6.94 g	GC-MS	Tokyo Bay, Japan				12.3 (9.94-15.24)		595	whole body	2887	2	Takeuchi et al. (2009)
1ercenaria stimpsoni	7.07 g	GC-MS	Tokyo Bay				12.3 (9.94	0.38	167	whole body	2202	2	Takeuchi et al. (2009)
Mytilopsis sallei	0.38 g	GC-MS	Tokyo Bay				12.3 (9.94	1.28	466	whole body	1822	2	Takeuchi et al. (2009)
1ytilus galloprovincialis	3.35 g	GC-MS	Tokyo Bay				12.3 (9.94	1.41	622	whole body	2207	2	Takeuchi et al. (2009)
Perna viridis	4.83 g	GC-MS	Tokyo Bay				12.3 (9.94	0.73	549	whole body	3762	2	Takeuchi et al. (2009)
Kenostrobus securis	0.56 g	GC-MS	Tokyo Bay				12.3 (9.94	0.83	606	whole body	3651	2	Takeuchi et al. (2009)
Mytilus galloprovincialis	4-5 cm	HPLC UV-VIS fluorescence	Gulf of Rijeka, Adriatic Sea, Croatia		- 36.04- 36.53	14.2- 15.6	5		2000	whole body		3 7	Bihari et al. (2007)
Mytilus galloprovincialis	4-5 cm	HPLC UV-VIS fluorescence	Gulf of Rijeka, Adriatic Sea, Croatia	8.18	36.04- 35.28	14.3- 15.6	52		210	whole body		3 7	Bihari et al. (2007)
Mytilus galloprovincialis	4-5 cm	HPLC UV-VIS fluorescence	Gulf of Rijeka, Adriatic Sea, Croatia		- 18.05- 20.60	12.8- 13.5	<1		>7000	whole body		3 7	Bihari et al. (2007)
Mytilus galloprovincialis	4-5 cm	HPLC UV-VIS fluorescence	Gulf of Rijeka, Adriatic Sea, Croatia		- 36.11- 36.90	14.2- 16.0	<1		>15000	whole body		3 7	Bihari et al. (2007)
Mytilus galloprovincialis	4-5 cm	HPLC UV-VIS fluorescence	Gulf of Rijeka, Adriatic Sea, Croatia		- 20.80- 24.20	13.1- 14.2	6		3000	whole body		3 7	Bihari et al. (2007)
1ytilus galloprovincialis	4-5 cm	HPLC UV-VIS fluorescence	Gulf of Rijeka, Adriatic Sea, Croatia	8.20	- 36.53- 37.57	14.5- 16.4	5		1600	whole body		3 7	Bihari et al. (2007)
Mytilus galloprovincialis	6.6±0.1 cm, 0.48±0.02 g _{dw}	HPLC flu.	Ïzmit Bay, Turkey				0.28	0.87±0.26	8100	whole body	46000	3 7	Telli-Karakoc et al. (2002)
Aytilus galloprovincialis	3.8±0.3 cm, 0.21±0.02 q _{dw}	HPLC flu.	Ïzmit Bay, Turkey				0.47	1.41±0.20	21000	whole body	75000	3 7	Telli-Karakoc et al. (2002)
lytilus galloprovincialis	5.1±0.2 cm, 0.19±0.02 g _{dw}	HPLC flu.	Ïzmit Bay, Turkey				0.19	1.30±0.32	27000	whole body	103000	3 7	Telli-Karakoc et al. (2002)
Mytilus galloprovincialis	6.1±0.3 cm, 0.17±0.01 g _{dw}	HPLC flu.	Ïzmit Bay, Turkey				2.47	0.49±0.47	540	whole body	6000	3 7	Telli-Karakoc et al. (2002)
Aytilus galloprovincialis	5.6±0.3 cm, 0.20±0.03 g _{dw}	HPLC flu.	Ïzmit Bay, Turkey				< 0.01	0.37±0.28	>320000	whole body	>4.4E6	3 7	Telli-Karakoc et al. (2002)
Mytilus galloprovincialis	4.6±0.3 cm, 0.19±0.02 g _{dw}	HPLC flu.	Ïzmit Bay, Turkey				< 0.01	0.27±0.14	>27000	whole body	>5.0E5	3 7	Telli-Karakoc et al. (2002)
Mytilus galloprovincialis	5.1±0.4 cm, 0.18±0.02 g _{dw}	HPLC flu.	Ïzmit Bay, Turkey				1.31	0.64±0.21	2700	whole body	11000	3 7	Telli-Karakoc et al. (2002)
Radix ovata	, 5011	GC-MS	Lake Redon, Pyrenees, Spain				~0.096			whole body	3.5E6	4 4,5	Vives et al. (2005)
Pisidium sp.		GC-MS	Lake Redon, Pyrenees, Spain				~0.096			whole body	160000	4 4,5	Vives et al. (2005)
Crustacea			. ,							,		,	` ,
Daphnia pulicaria		GC-MS	Lake Redon, Pyrenees, Spain				~0.096			whole body	8900	4 1,5	Vives et al. (2005)
Hemigrapsus penicillatus	0.42 q	GC-MS	Tokyo Bay, Japan				12.3 (9.94-15.24)	2.76	438	whole body	793	2	Takeuchi et al. (2009)
Monoporeia affinis	3	HPLC flu. (water) GC-MS (biota)	Baltic Sea, Bothnian Sea		7.5 ± 0.5	2.0±0.5	0.381±0.070	1.13	1200	whole body	5400	3 8	Nfon et al. (2008); Witt (2003)
Mysis sp.		HPLC flu. (water) GC-MS (biota)	Baltic Sea, Bothnian Sea		7.5 ± 0.5	2.0±0.5	0.381±0.070	0.51	990	whole body	9700	3 8	Nfon et al. (2008); Witt (200
Saduria entomon Pisces		HPLC flu. (water) GC-MS (biota)	Baltic Sea, Bothnian Sea		7.5±0.5	2.0±0.5	0.381±0.070	0.21	200	whole body	4900	3 8	Nfon et al. (2008); Witt (200
Acanthogobius flavimanus	9.12 g	GC-MS	Tokyo Bay, Japan				12.3 (9.94-15.24)	0.3	63	whole animal	1043	2	Takeuchi et al. (2009)
Clupea harengus	- 3	HPLC flu. (water) GC-MS (biota)	Baltic Sea, Bothnian Sea		7.5±0.5	2.0±0.5		0.58	480	whole body	4100	3 8	Nfon et al. (2008); Witt (2002
Salmo trutta	field sampled, 286±26mm, 230±58 g, 11±4 years	GC-MS	Lake Redon, Pyrenees, Spain			2.0-0.0	~0.096	1.2	110000	liver	470000		Vives et al. (2005)
Salvelinus namaycush siscowet	527±18 mm, 1.3±0.1 kg, 9.2±0.9 years		Lake Superior, USA				3.49±1.71	20.5	18	fillet	4.4	3 6	Burkhard and Lukasewycz (2

Notes

- 1 Lipid normalized BAF read from figure.
- 2 Lipid content of 4.6% is for the liver based on dry weight, the lipid content in the muscles was 3%.
- 3 Average water content in brown trout tissue of 74.2% used to recalculate to fresh weight BAF (not normalized).
- 4 Based on ratios of reported concentrations in organisms and lipid contents and the BAF for brown trout.
- Not clear if biota and water were sampled at the same time. Water concentrations are averages over 1.5 year. Concentrations show some (possibly seasonal) variation (Vilanova et al., 2001).
- 6 Trout sampled in 1991, water sampled in 1986. Sampling location in Lake Superior were not the same as well.
- 7 Samples were collected and extracted unfiltered. Therefore, the aqueous concentrations do not represent dissolved concentrations. This may explain the variable and sometimes very high water concentrations and BAFs.
- 8 Biota samples collected in 1991–1993, water sampled from 1992-1998 (Witt, 2002). Water samples not exactly the same location as the biota samples. Nevertheless, water concentrations seem rather constant over time and over water. Total water concentrations monitored, but particulate organic carbon is low (~0.25 mq.L⁻¹).

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